Hydroxyakalone, a Novel Xanthine Oxidase Inhibitor Produced by a Marine Bacterium, Agrobacterium aurantiacum

Hitoshi Izumida, Kyoko Adachi, Akira Mihara, Tohru Yasuzawa and Hiroshi Sano

Marine Biotechnology Institute, Shimizu Laboratory, 1900 Sodeshi-cho, Shimizu-shi, Shizuoka 424, Japan
1Kansai Paint Co., Ltd., Technical Research Laboratory, 4-17-1 Higashi-Yawata, Hiratsuka 254, Japan
Pharmaceutical Research Laboratories, Kyowa Hakko Kogyo Co., Ltd., 1188 Shimotogari, Nagaizumi-cho, Shizuoka 411, Japan
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A new xanthine oxidase inhibitor named hydroxyakalone was isolated from the culture broth of a marine bacterium Agrobacterium aurantiacum N-81106. Structure of hydroxyakalone was determined to be 4-amino-1H-pyrazolo[3,4-d]pyrimidine-3-one-6-ol by the spectral studies of hydroxyakalone and its permethyl derivative. The concentration to induce 50% inhibition (%I) was 4.6 μM against xanthine oxidase.

Xanthine oxidase (XOD, EC 1.2.3.2) catalyzes the oxidation of hypoxanthine into uric acid via xanthine. It is well known that an excessive accumulation of uric acid in the blood, called hyperuricacidemia, causes gout. A number of XOD inhibitors such as anthraquinones, flavonoids, xanthones and caffeic ester have been found from plant sources.

In a previous paper, we reported a new XOD inhibitor, akalone from a marine bacteria, Agrobacterium aurantiacum N-81106. When cultivation conditions of A. aurantiacum N-81106 were changed using a 1000 liters fermenter, akalone was not produced, and another strong XOD inhibitor was found to be produced instead of akalone. In the present paper, we describe the fermentation, isolation and structural determination of this XOD inhibitor named hydroxyakalone.

Materials and Methods

General
The UV absorption spectrum was recorded on a Shimadzu UV-2000S recording spectrophotometer in distilled water, while NMR spectra were recorded with a Varian Unity 500 spectrometer using D2O and DMSO-d6 solvents. The FAB mass spectra (FAB-MS) were measured with a JEOL JMS-SX102 mass spectrometer.

Fermentation
Pre-incubated Agrobacterium aurantiacum N-81106 in a 500 ml Erlenmeyer flask was inoculated into a 30-liter jar fermenter containing 18 liters of E3 medium composed of artificial seawater 500 ml, deionized water 500 ml, peptone 5.0 g (Kyokuto kogyo), yeast extract 1.0 g (Oriental yeast industry), glucose 2.0 g, FePO4 0.01 g (pH 7.7 before sterilization) and was incubated for 4 days at 20°C. The resulting fermentation broth was inoculated into a 2000-liter large scale fermenter containing 1000 liters of the same medium and was incubated for 4 days at 20°C.

Assay Method
Supernatant obtained from the fermentation broth by centrifuging at 18,600 g for 5 minutes was provided as a test solution. The bioassay for XOD inhibitory activity was performed according to a modified version of Noro's method, using XOD from cow's milk that was obtained from Sigma Chemical Co. A mixture consisting of 0.3 ml of test solution and 1.0 ml of XOD solution, which was prepared at a concentration of 0.22 units/ml with 50 mM phosphate buffer, was pre-incubated for 15 minutes at 25°C. The enzymatic reaction was initiated by adding to the mixture 1.7 ml of substrate solution containing xanthine at a concentration of 1.74 μg/ml. The optical density of the mixture was recorded at 290 nm by time scanning while incubating at 25°C. The enzyme activity was determined using the expression (1 - B/A) x 100, where A and B are the levels of XOD activity without and with test solution, respectively.
Isolation of XOD Inhibitor

A 1000 liters sample of the fermentation broth of the *Agrobacterium aurantiacum* N-81106 was filtered to remove solid materials. The solution was diluted by 2.5 times with distilled water and adjusted to pH 4.0 with acetic acid. The solution was applied to an ion-exchange column [DIAION SK104 (H+), 50 liters] and eluted with 2n NH4OH. Active fraction was concentrated to 120 liters and was further purified on an ODS column (YMC ODS-AQ, 10 liters) eluting with water, followed by gel filtration (Sephadex G-25 column, 1 liter) eluting with ammonium acetate (pH 8.5). The active fraction was evaporated to dryness to give 158 mg of hydroxyakalone as a colorless powder.

Methylation of Hydroxyakalone

To a suspension of hydroxyakalone (10 mg) in DMF (1 ml), silver (I) oxide (460 mg) and methyl iodide (95 μl) were added, and the mixture was stirred vigorously at 25°C for 4 days. The reaction mixture was filtered, washing the solids with chloroform. The filtrate was evaporated to give residue. An aqueous solution of the residue was applied to an ODS HPLC column (Cosmosil C18 AR, Nacalai Tesque, HPLC conditions: column size, 20 mm i.d. × 250 mm; mobile phase, 50% CH3CN with 0.1% TFA; flow rate 5 ml/minute; detection UV 210 nm). Evaporation of the major absorbing fraction gave 0.5 mg of methylated hydroxyakalone as a colorless powder.

Results and Discussion

Structure of Hydroxyakalone

Physico-chemical properties of hydroxyakalone are summarized in Table 1. Hydroxyakalone is soluble in aqueous alkali and DMSO and insoluble in methanol, chloroform and n-hexane. Negative FAB-MS of hydroxyakalone showed a pseudo molecular ion (M−H)^− at m/z 166, and the molecular formula was determined to be C5H5N5O2 on the basis of HR negative FAB-MS. The UV absorption spectrum showed a maximum at 298 nm, and the 13C NMR spectrum (Table 2) exhibited five sp^2 carbons, implying the existence of a conjugated heterocyclic system. The 1H NMR spectrum in DMSO-d6 showed five exchangeable protons, that is, there were no carbon-bonded protons. These data suggested that hydroxyakalone was an analog of akalone. However, no more information could be obtained from the spectra of the intact compound, and further investigation was carried out using the methyl derivative.

Methylation of hydroxyakalone with methyl iodide and silver oxide afforded several products. One of the products had the molecular formula C10H15N5O2 which was indicated by HR FAB-MS (Table 1). The difference of molecular formulas between hydroxyakalone and its methylate suggested that five methyl groups were introduced into hydroxyakalone.

The five methyl groups introduced were assigned to two N-CH3 (δH 3.36, δC 31.20 and δH 3.21, δC 26.93), one N-(CH3)2 (δH 2.93, δC 40.36) and one O-CH3 (δH 3.82, δC 53.91) groups by 1H NMR, 13C NMR and HSQC data. The HMBC spectrum showed long range couplings between the two N-CH3 protons and the 6-carbonyl carbon, and between the 7-NCH3 protons and the 7a-carbon (Fig. 1), which confirmed the partial structure from N-5 to the 7a-carbon. A NOE was observed between N(CH3)2, which was long range coupled with C-4, and 5-NCH3, implying the bonding between N-5 and C-4. Since the neighboring atoms of the sp^2 quarternary carbon at δC 104.35 were carbons

| Appearance | Colorless powder | Colorless powder |
| Molecular formula | C5H5N5O2 | C10H15N5O2 |
| HR/AB-MS (M−H)^− (m/z) | 166.0339 | 166.0365 |
| IR (KBr) cm^-1 | 3300, 3088, 1711, 1616 | 3418, 2926, 2858, 1702, 1632, 1466, 1377 |
| UV λmax (H2O, pH 12.6) | 298 nm | 282 nm (MeOH) |
| Solubility | Soluble in: H2O, MeOH, CHCl3 | MeOH, CHCl3, DMSO |
| Insoluble in: | H2O | MeOH, CHCl3, DMSO |
Based on its chemical shift, the neighboring atoms must be C-4, C-7a and C-3. Thus, the connectivity of C-3 to C-4 was established. A NOE between N(CH$_3$)$_2$ and 3-OCH$_3$, which was long range coupled with C-3, also assisted this connectivity. The partial structure C-3 through C-7a contained all elements except two nitrogens. Thus the whole structure of methylated hydroxyakalone was determined to be as shown in Fig. 2, introducing the remaining two nitrogens between the 3- and 7a-carbons. The chemical shifts of C-3, C-3a and C-7a were reasonable for this structure. In the HMBC study, $^4$J$_{CH}$ coupling was observed between 5-NCH$_3$ protons and the 3a-carbon instead of $^3$J$_{CH}$ coupling with the 4-carbon. The total assignments of $^1$H and $^{13}$C NMR signals of methylated hydroxyakalone are listed in Table 3.

Owing to the establishment of the structure of methylated hydroxyakalone, the novel structure of hydroxyakalone was determined to be 4-amino-1H-pyrazolo[3,4-d]pyrimidine-3-one-6-ol as shown in Fig. 2 where the 6-position of akalone is substituted by a hydroxy group. In its structure, a contribution of 6-one and 3-ol forms was thought to exist.

**XOD Inhibitory Activity of Hydroxyakalone**

Hydroxyakalone showed XOD inhibitory activity stronger than akalone. The 50% inhibition concentration (IC$_{50}$) of hydroxyakalone was 4.6 $\mu$m, compared with 16.9 $\mu$m for akalone and 4.0 $\mu$m for allopurinol.

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**References**

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