Lymphostin (LK6-A), a Novel Immunosuppressant from *Streptomyces* sp. KY11783:

Structural Elucidation

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Lymphostin (LK6-A), a novel immunosuppressant, was isolated from the culture broth of *Streptomyces* sp. KY11783. Lymphostin showed potent inhibitory activity against the lymphocyte kinase, Lck (p56^{lck}). The structure of lymphostin was determined by spectroscopic methods to be a novel tricyclic aromatic alkaloid. A key feature of the structure is the pyrrolo[4,3,2-*de*]quinoline moiety which is rare among natural products.

We have isolated a potent immunosuppressant, lymphostin (LK6-A¹, Fig. 1), from the culture broth of *Streptomyces* sp. KY11783. Lymphostin inhibited the activity of the lymphocyte kinase, Lck (p56^{lck}), with an IC₅₀ value of 0.05 μ M. In the preceding paper²⁾, fermentation, purification, and biological properties of lymphostin were reported. In this paper, we describe the details of its structure determination.

Structural Elucidation

The physico-chemical properties of lymphostin are summarized in Table 1. The UV spectrum indicated the presence of a polyaromatic system. The IR spectrum suggested the existence of a secondary amide and a ketone conjugated with double bonds. The molecular formula for lymphostin was established as C₁₆H₁₄N₄O₃ by the HR-FAB-MS measurement, implying twelve degrees of unsaturation in the molecule.

In the ¹H NMR spectrum in dimethylsulfoxide-*d*₆ (DMSO-*d*₆), one methyl singlet (δ_{H} 2.35), one *O*-methyl (δ_{H} 3.94), three aromatic methine singlets (δ_{H} 8.14, δ_{H} 8.39, δ_{H} 8.62), and a pair of coupled olefinic protons (δ_{H} 7.87, δ_{H} 7.64, *J* = 12.6 Hz) were observed. Also observ-

ed were two additional broad signals at δ_{H} 8.21 (two protons) and at δ_{H} 10.14 (one proton). According to the molecular formula and the IR spectrum, they were assigned to an amino and an amide proton respectively.

The ¹³C NMR (DMSO-*d*₆) spectrum showed all sixteen signals and the DEPT experiments revealed the presence of two methyls, five methines and nine quaternary carbons. Direct ¹³C-¹H connectivities were established by HSQC (Heteronuclear Single Quantum Coherence) experiment. The quaternary carbons at δ_{C} 188.7 and δ_{C} 169.7 should be assigned to a conjugated carbonyl carbon and an amide carbonyl carbon respectively, considering the results of the UV and the IR spectra. One of the methines was assumed to be adjacent to a nitrogen atom in a pyrrol ring based on its chemical shifts (δ_{C} 145.8, δ_{H} 8.39) and a large ¹*J*_{CH} coupling constant (190 Hz).

Complete ¹H and ¹³C NMR assignments obtained from analyses of one and two dimensional NMR experiments are exhibited in Table 2. Connectivities between structural elements were mainly established with

Fig. 1. Structure of lymphostin.

Table 1. Physico-chemical properties of lymphostin.

MP	275~277°C
Molecular weight	310
Molecular formula	C ₁₆ H ₁₄ N ₄ O ₃
FAB-MS (<i>m/z</i>)	311 [M+H] ⁺
HRFAB-MS (<i>m/z</i>)	
Observed	311.1137 [M+H] ⁺
Calcd	311.1144
UV $\lambda_{\text{max}}^{\text{CH}_3\text{CN}}$ nm (ϵ)	467 (8,200), 412 (6,000), 344 (11,900), 284 (16,000), 256 (19,300), 218 (15,200)
IR ν_{max} (KBr) cm ⁻¹	3367, 1671, 1659, 1633, 1595, 1506, 1362, 1350, 1236, 1128, 1076

Table 2. ^{13}C and ^1H NMR data of lymphostin in $\text{DMSO}-d_6$.

Position	δ_{C} (m) [125 MHz]	δ_{H} (m, J in Hz) [500 MHz]
2	145.8 (d)	8.39 (s)
2a	136.6 (s)	
3	117.6 (d)	8.62 (s)
4	147.7 (s)	
5a	133.5 (s)	
6	139.5 (s)	
7	110.8 (d)	8.14 (s)
8	150.3 (s)	
8a	124.9 (s)	
8b	124.4 (s)	
9	188.7 (s)	
10	101.9 (d)	7.64 (d, 12.6)
11	163.9 (d)	7.87 (d, 12.6)
12	58.5 (q)	3.94 (s)
13	169.7 (s)	
14	24.5 (q)	2.35 (s)
6-NH-		10.14 (s)
8-NH ₂		8.21 (s)

δ : ppm from TMS, m: multiplicity.

Fig. 2. ^1H - ^{13}C long range couplings of lymphostin.

Arrows are directing H to C.

Fig. 3. Partial structure of lymphostin.

Numbers are ^{13}C NMR chemical shifts. Underlined numbers are ^{13}C NMR chemical shifts of deuterized lymphostin.

HMBC experiment (Fig. 2). The *O*-methyl proton H-12 (δ_{H} 3.94) showed long range ^{13}C - ^1H coupling with the olefinic methine carbon C-11 (δ_{C} 163.9) and the two olefinic methine protons H-10 and H-11 (δ_{H} 7.64, δ_{H} 7.87) had long range couplings with the carbonyl carbon C-9 (δ_{C} 188.7), indicating the presence of a β -methoxy enone structure. The geometry of the double bond was determined to be *E*-configuration from the result of NOESY experiment, in which *O*-methyl proton H-12 had a correlation to H-10 as well as to H-11. Also indicated was the presence of an acetamido structure, from the observation of correlations of the amide carbonyl carbon C-13 (δ_{C} 169.7) to the methyl proton H-14 (δ_{H} 2.35) and to the amide proton (δ_{H} 10.14). These two structures were supposed to be the side chains of the polyaromatic moiety of lymphostin. Although the rest of the HMBC information (Fig. 2) allowed definition of several other partial structures, it was not enough to determine the whole structure because of the ambiguity of three *versus* four bond couplings in the HMBC experiments. Therefore, we added CD_3OD - CD_3OH (1:1) solvent to the $\text{DMSO}-d_6$ solution of lymphostin in order to obtain deuterium shift information in ^{13}C NMR spectra. In that experiment, several carbon signals showed a characteristic high field shift³⁾, which led us to construct the structure in the vicinity of labile protons as shown in Fig. 3. The remained partial structures were connected according to other long range ^{13}C - ^1H couplings (Fig. 2). The structure of lymphostin in Fig. 1 was finally determined

after conducting a differential NOE experiment, in which we found an important NOE correlation between aromatic methines H-2 (δ_{H} 8.39) and H-3 (δ_{H} 8.62).

Discussion

Lymphostin was determined to be a novel tricyclic aromatic alkaloid with a pyrrolo[4,3,2-*de*]quinoline skeleton. The high unsaturation of the structure needed analyses of the various NMR spectroscopic experiments until the structure determination was achieved.

This fused aromatic ring system is rare among natural products. The plakinidines^{4,5)} are the only natural compounds reported to possess the pyrrolo[4,3,2-*de*]quinoline system. Plakinidine A was reported to have reverse transcriptase inhibiting activity, anti-parasite activity and cytotoxicity to L1210. Plakinidine B and C were also reported to have one or two of these activities. However, they have not been reported to inhibit any protein kinase activities. Further investigation of the activities of the novel compound, lymphostin, will definitely provide substantial information for the development of protein kinase inhibitors.

Experimental

The melting point was determined with a Yanagimoto melting point apparatus. The FAB-MS spectrum was measured with a JEOL JMS-HX/HX110A mass spectrometer. The UV spectrum was recorded on a Shimadzu UV-2200 spectrometer. The IR spectrum was taken on a JEOL JIR-RFX3001 spectrometer. The NMR spectra were recorded on a Bruker AM500 spectrometer or a JEOL A-400 using TMS as an internal standard.

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