Histochemical Demonstration of NADPH-Diaphorase Activity in the Pineal Organ of the Frog (*Rana esculenta*), But Not in the Pineal Organ of the Rat

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Received June 3, 1989

Summary. Using the histochemical method for the demonstration of NADPH-diaphorase activity, the pineal organ of the frog and rat was investigated in serial sections. A positive NADPH-diaphorase activity was demonstrated in pinealocytes and nerve cells in the pineal organ of the frog, but not in the rat. An intense activity existed in the apical portion of the photosensitive pinealocytes of the frog. Large NADPH-diaphorase positive nerve cells (15-20 μm in diameter) were located within the parenchyma of the pineal organ in the frog. Large NADPH-positive nerve cells were more numerous in the rostral than in the caudal portion of the organ, but the intensely stained cells, counting 25-35 in number, showed almost equal distribution and number in the ventral and the dorsal aspect of the pineal organ. In their staining ability, NADPH-diaphorase positive pineal nerve cells resembled retinal amacrine cells.

The results in the pineal organ of the frog are discussed in light of previous morphological findings using the acetylcholinesterase reaction, and with electrophysiological results.

The presence of different types of neurons in the pineal organ of anurans has been demonstrated by supravital methylene blue staining and silver impregnation techniques (PAUL et al., 1971), and by the acetylcholinesterase (AChE) method (WAKE et al., 1974). However, the precise spatial and functional interrelationship of the cells is difficult to disentangle; unlike the organization of the retina, the wall of the pineal organ does not show a distinct pattern of layers.

The pineal organ of *Rana ridibunda* and *Rana esculenta* comprises two different types of nerve cells — multipolar and pseudounipolar nerve cells — as shown by the AChE-method according to Karnovsky and Roots (WAKE et al., 1974), the former cells being closely connected to photosensitive pinealocytes, whereas the latter are scattered along the pineal tract, sending one process toward the brain. Therefore, it has been suggested that the former, multipolar cells are interneurons. A similar morphological differentiation of AChE-positive neurons has also been observed in the trout (KORF, 1974; EKSTRÖM and KORF, 1985), in the pike (FALCON, 1979), the European minnow (VIGH-TEICHMANN et al., 1982) and in several species of birds (UECK and KOBAYASHI, 1972; SATO and WAKE, 1983, 1984).

Recent investigations have achieved a selective demonstration of amacrine cells in the retina of several mammalian species using NADPH-diaphorase histochemistry (SANDELL, 1985; SAGAR, 1986; MÜLLER and PEICHL, 1987); in a previous paper I reported similar results in the frog and pigeon (SATO, 1990). The present paper aims at an initial investigation as to whether the pineal organ might contain NADPH-diaphorase positive cells. The study was carried out in a photosensitive pineal organ of the frog and in a merely secretory pineal organ of the rat.

MATERIALS AND METHODS

Ten adult Wistar rats (200-250 g) and 20 frogs, *Rana esculenta*, were investigated. The frogs were imported from Yugoslavia over the period from August till
November. All animals were kept under natural light conditions. The animals were sacrificed between 9:00 and 12:00 a.m. as described below.

After opening the thorax under Ketavet anesthesia (i.m. 20 mg/kg body weight), the animals were perfused via the left ventricle or the aorta with 25-50 ml of ice-cold saline followed by an aldehyde fixative solution. The perfusion was done with 100-250 ml of an ice-cold mixture of 4% paraformaldehyde and 0.5% glutaraldehyde in 0.1 M phosphate buffer (pH 7.4). After perfusion, the brain was quickly removed and left in 4% phosphate-buffered (pH 7.4) paraformaldehyde fixative for 2 h at 4°C. The whole brains were stored overnight in 0.1 M phosphate buffer (pH 7.4) containing 15% sucrose at 4°C. Frontal, horizontal and sagittal sections (20 μm thick) were serially cut on a cryostat (American Optical) and incubated for the demonstration of NADPH-diaphorase activity using the method of SCHERER-SINGLER et al. (1983) (see SATO, 1990). From the serial sagittal sections of two frogs, the NADPH-diaphorase positive nerve cells were plotted at the light-microscopic level. Some in-toto preparations were also incubated for the demonstration of NADPH-diaphorase activity.

RESULTS

The NADPH-diaphorase method stains a distinct population of nerve cells in the brain—as is shown in the frog’s striatum (Fig. 1). In the frog’s pineal organ, NADPH-diaphorase positive nerve cells are visualized; additionally, an intense enzyme activity occurs in the apical protrusion of the photosensitive pinealocytes (Figs. 2-4), whereas the nuclei are unstained and the basal portion of the sensory cells seems to be faintly stained dependent on the thickness of the sections and the intensity of the staining. However, the pinealocytes are easy to distinguish from the visualized nerve cells: the latter are larger, spherical in shape and located at the basal of the parenchyma, while the former are smaller, elongated and contact the pineal lumen (Figs. 5-8).

Additional to the visualized photosensitive pinealocytes which contact the pineal lumen are demonstrated cells containing a spherical, NADPH-diaphorase positive region. These cells are located near the basal lamina (Figs. 3, 4).

The diaphorase-positive large nerve cells (15-20 μm in diameter) were counted in serial sections of two pineal organs (Fig. 9) at 25 and 35 in number. These cells predominate in the rostral part of the pineal organ, but a few cells are visible also in the caudal part. The stained nerve cells are seen similar in number in the ventral and the dorsal wall of the pineal organ. They are intramurally localized (Figs. 5-7): strong enzyme activity occurs in the cytoplasm of the perikarya, but the processes of the nerve cells are only faintly stained in comparison to the strong results in nerve cells of the brain (Fig. 1) and in some types of amacrine cells of the retina (SAITO, 1990). In addition to the large intramural nerve cells, some smaller nerve cells can be visualized in the dorsal wall (Fig. 8); the total number of the latter cell type is difficult to count because of the diminished intensity of the staining and the extremely thin cytoplasmic layer surrounding the unstained nucleus.

Contradictory to the staining of photosensitive pinealocytes in the frog’s pineal organ is the result in the rat’s pineal organ. Here, no recognizable NADPH-diaphorase activity could be recognized in the secretory pinealocytes of the rat. The visualization of distinct neurons in adjacent brain areas (Fig. 10) supports the view that the pinealocytes of rats are NADPH-diaphorase negative.

DISCUSSION

The NADPH-diaphorase-method provided a demonstration of pinealocytes and nerve cells in the pineal organ of Rana esculenta. The reaction product of the diaphorase activity is localized in the cytosol, according to an ultrastructural study by VINCENT and JOHANSSON (1983). Neurohistological studies using methylene blue, Bodian or Golgi techniques in the pineal organ (STUDNIČKA, 1905; HOLMGREN, 1918, 1919; OKSCHE, 1955; PAUL et al., 1971) visualized only part of the nervous apparatus. The best results to date were obtained by using the AChE-reaction (WAKE et al., 1974). Two different types of AChE-reactive nerve cells have been distinguished: multipolar cells and smaller pseudounipolar cells. The former cells are localized within the parenchyma, while the perikarya of the latter often protrude into the perivascular connective tissue. Concerning the function of these two main types of nerve cells, it has been suggested from the results of WAKE et al. (1974) that the pseudounipolar cells give rise to the pineal tract, while the multipolar elements have been interpreted as interneurons. From their size and localization, we compare the large NADPH-diaphorase positive cells with the multipolar cells shown with the AChE-method. Moreover, from the intimate spatial relationship between the photosensitive pinealocytes and the large NADPH-diaphorase positive neurons
Figs. 1-4. *Rana esculenta*, NADPH-diaphorase method. **Fig. 1.** Sagittal section of an area from the torus semicircularis of the frog brain. A distinct population of neurons demonstrated in a Golgi-like image: perikarya, dendritic trees, and axons are stained. ×130. **Fig. 2.** Mid-sagittal section of the frog pineal organ. Adjacent to the pineal lumen (L), the apical portion of the photosensitive pinealocytes is intensely stained. Arrowheads indicate positive neurons. *EP* epithalamus. ×210. **Fig. 3.** Frontal section of the pineal organ. Innersegments of photosensitive pinealocytes (*asterisks*) are initially stained after a short incubation time (60 min). *L* pineal lumen. ×560. **Fig. 4.** A higher powered view of an area marked by the rectangle in Fig. 3. Beside the photosensitive pinealocytes (*asterisks*) adjacent to the pineal lumen (*L*), cells (*solid stars*) are stained containing a spherical, NADPH-diaphorase positive product; these cells are located near the basal lamina; they obviously have no contact with the pineal lumen. *PT* pineal tract. ×1,150
(Sato and Ueck, 1989; Ueck et al., 1989), and, on the other hand, between the multipolar cells and pseudounipolar nerve cells (Wake et al., 1974), it has been emphasized that the multipolar cells of the frog pineal organ might be interneurons. The NADPH-diaphorase method selectively stains types of amacrine cells in the inner nuclear layer of the frog's retina (Sato, 1990). Multipolar nerve cells in the pineal organ of the frog, Rana esculenta, are similar to retinal amacrine cells in respect to their NADPH-diaphorase- and AChE-activity.

Cells characterized by a spherical, NADPH-diaphorase positive region in their cytoplasm and by their location near the basal lamina (Figs. 3, 4) are obviously without contact with the pineal lumen; they are assumed to be modified pinealocytes because of the similarity of their stained area with that of the inner segment of the photosensitive cell. Modified
pinealocytes have been described in other lower vertebrates, i.e., the lamprey (MEINEL, 1981).

The direct photosensitivity of the pineal complex of Anura is clearly demonstrated by electrophysiological studies. Direct illumination of the frontal organ is followed by chromatic and achromatic responses, whereas the pineal organ proper usually responds achromatically (DODT and HEERD, 1962; MORITA, 1965; DODT et al., 1971; DODT, 1973). A detailed description of morphological characteristics of the frontal organ shown by the NADPH-diaphorase method is in preparation.

Conventional synapses have been observed beside the ribbon synapses in the pineal organ of the frog. Their functional significance is obscure, but it has been suggested that they may be linked to interneurons (BAYRHUBER, 1972; KORF, 1976). Therefore, it is relevant to investigate the synapse formation using the NADPH-diaphorase method at the ultrastructural level.

Distinct populations of neurons have been confirmed with the NADPH-diaphorase histochemistry in the rat brain (Fig. 10), similar to the previous observations by SCHERER-SINGLER et al. (1983), VINCENT et al. (1983), and VINCENT and JOHANSSON (1983). However, no NADPH-diaphorase activity could be recognized in the pineal organ. Considering that the pineal organ of the rat is free of neuronal perikarya (ARIEŃ S KAPPERS, 1960, 1965), the absence of diaphorase-activity should be ascribed to the feature of pinealocytes. This difference in staining ability of photosensitive pinealocytes and retinal photoreceptors on one hand and secretory mammalian pinealocytes on the other hand is interesting in respect to the general discussion of retinal and pineal relationships (for literature see O'BRIEN and KLEIN, 1986).

Acknowledgement. This work was supported by grants from the Alexander von Humboldt Foundation. The author is indebted to Professor M. UECK for his patient support and valuable discussions and thanks Mrs. A. HACH for her efficient technical assistance.

REFERENCES


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