Inhibitory Effects of Saururus chinensis (Lour.) Bail on the Development of Atopic Dermatitis-Like Skin Lesions in NC/Nga Mice

Myoung Suk Choi, Eui Chul Kim, Hyung Suk Lee, Sun Kwang Kim, Hyun Myung Choi, Jung Hyuk Park, Jae-Bok Han, Hyo Jin An, Jae Young Um, Hyung Min Kim, Ah-Reum Han, Moo Chang Hong, Hyunsu Bae, and Byung-Il Min

Department of East-West Medicine, Graduate School, Kyung Hee University; College of Oriental Medicine, Institute of Oriental Medicine, Oriental Medical Science Center, Kyung Hee University; Department of Physiology, College of Oriental Medicine, Kyung Hee University; and Department of Physiology, College of Medicine, Kyung Hee University;

#1 Hoegi-dong, Dongdaemoon-gu, Seoul 130-701, Republic of Korea.

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Atopic dermatitis (AD) is a common inflammatory skin disorder that is both uncomfortable and distressing to patients as a result of its associated intense pruritus and unsightly lesions. When severe, AD can be extremely disabling, causing major psychological problems, and, in the case of a young child, be overwhelming to the entire family. AD is a genotypic diathesis in which minor skin stimulation is perceived as an itching that, when scratched or rubbed, elicits a heightened immune response. The heightened immune response leads to the development of eczema, which is the clinical syndrome of eczematous dermatitis group of inflammatory skin conditions characterized by pruritus, pale, erythematous and violaceous hues, vesiculation, erosion, scaling, exudation, crusting, lichenification, and excoriation. It is well established that AD is a model of AD.21,22) The NC/Nga strain originated from Japan and China. Only a few reports have documented the biological activities of SC herb indicating that the extract and lignans isolated exhibited hepato-protective, anti-inflammatory or antioxidative activity.14—17) SC contains flavonoids, including quercetin, quercitrin, isoquercitrin and rutin, as active components. It has been reported that flavonoids possess a number of biological effects such as antiallergic, anti-inflammatory, antiviral, antiproliferative and anticarcinogenic activities.18,19) Antiinflammatory effect of these flavonoids suggests the possibility of their therapeutic efficacy in various inflammatory diseases.20) However, it has not yet been studied if and how SC suppresses the development of AD. In this study we investigated therapeutic effects of SC on AD-like skin lesions in NC/Nga mice.

The present study was performed to examine whether the leaves of Saururus chinensis (Lour.) Bail (SC), an herb used for the management of various skin diseases including atopic dermatitis (AD) in Eastern countries, inhibited the development of AD-like skin lesions in NC/Nga mice which was induced by repeated application of picryl chloride (PIC). The efficacy of SC was judged by measurement of skin severity, itching behavior, histological study, serum IgE levels, IL-4 and IFN-γ in lymph nodes. Oral administration of SC extract to the PIC-treated NC/Nga mice for 8 weeks (5 d per week) inhibited significantly the development of AD-like skin lesions macroscopically. Histologically, SC inhibited dermatitis changes like hypertrophy, hyperkeratosis, and infiltration of inflammatory cells into epidermis and dermis. The itching behavior and serum IgE level decreased significantly after SC administration. SC administration enhanced IFN-γ mRNA expression but did not have an effect on IL-4 mRNA expression. These results suggest that SC could inhibit the development of AD-like skin lesions in NC/Nga mice possibly through modulating the Th1/Th2 imbalance by the promoting of Th1 cell response. Thus, SC may be an alternative substance for the management of AD patients.

Key words:
- atopic dermatitis
- Saururus chinensis
- NC/Nga mouse
- interferon gamma
- interleukin-4
- Th1/Th2

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MATERIALS AND METHODS

Animals Female 4-week-old NC/Nga mice were purchased from SLC (Shizuoka, Japan) and were maintained for 2 weeks before the start of the experiments. They were housed in an air-conditioned animal room with a 12-h light/dark cycle (08:00—20:00 h light, 20:00—08:00 h dark), at 23 ± 2 °C and a humidity of 50 ± 10%. The study was approved by the Institutional Animal Care and Use Committee of Kyung Hee University and all procedures were conducted in accordance with the U.S. National Institutes of Health guidelines. Mice were divided into three groups: (1) control group (0.9% normal saline administered NC/Nga mice, 0.2 cc/d, n=7), (2) SC100 group (100 mg/kg SC extract administered NC/Nga mice, 0.2 cc/d, n=8), (3) SC250 group (250 mg/kg SC extract administered NC/Nga mice, 0.2 cc/d, n=8).

Reagents 1-Chloro-2,4,6-trinitrobenzene (Picryl Chloride, purchased from Toctis Japan) was used after recrystallization with ethanol. Picryl chloride (PiCl)-induced AD-like skin lesion was developed in NC/Nga mice according to standard instructions provided by Charls River Japan. Briefly, the abdomen and ears of mice were sensitized epicutaneously with 150 μl of 5% PiCl dissolved in an ethanol and acetone mixture (4:1). On day 5 after sensitization, the dorsal skin of mice was challenged with 150 μl of 1% PiCl dissolved in olive oil. After the first challenge, 1% PiCl solution was repeatedly applied to the dorsal skin of mice for further 8 times at 1-week intervals.

Preparation of SC Extract The leaves of *Saururus chinensis* (Lour.) BAILL. (Saururaceae) as a dried herb was purchased from Omniherb (Youngchun, South Korea). It has grown in Andong of South Korea. *Saururus chinensis* was authenticated by H. Bae, College of Oriental Medicine, Kyung Hee University. A voucher specimen was deposited at the Herbarium of the College of Oriental Medicine, Kyung Hee University. SC (600 g) was steeped in boiling distilled water for 30 min and filtered through a nylon mesh. After centrifugation at 5000 g for 20 min 4 °C, the supernatant was freeze-dried, providing SC extract (90 g, 15% w/w).

Quantitative Analysis of Rutin in SC SC extract was accurately weighed to 15 mg and dissolved in 1 ml of 50% methanol. For the quantitative analysis to SC, rutin, one of the known flavonoid constituents of SC, was purchased from Sigma-Aldrich, Inc., U.S.A., as the standard material. Rutin was weighed to 1 mg and dissolved in 1 ml of 50% acetonitrile in methanol. The dissolved standard solution was diluted to 0.1, 0.25, and 0.5 mg/ml, respectively. HPLC analysis was performed to authenticate SC. HPLC analysis were conducted using Waters system (Waters Co., Milford, MA, U.S.A.) with a 717+ autosampler, 2487 dual λ absorbance detector, and 1525 binary HPLC pump, and Waters Millenium^32^ System (Waters Co., Milford, MA, U.S.A.) was used for data acquisition and integration. The samples were analyzed by reverse phase (C₁₈) analysis HPLC (XTerra™ RP₁₈, 4.6×250 mm i.d., 5 μm, flow rate: 1 ml/min, UV detection: 250 nm) using a gradient solvent system of acetonitrile–methanol–water (1:1:2, 30 min). The relationship between the concentration and the peak-area was observed by the minimum square method (R² value). The standard calibration curve for rutin is Y=30146120X+282562.5 (R²=0.999). The average content of rutin in SC extract was calculated 54.6±1.2 mg/g (n=3) by the above formula (Fig. 1).

Administration of SC SC extract of different concentration (100, 250 mg/kg, 0.2 cc/d) was orally administered by gastric intubation with an animal-feeding needle, 5 d per week for 8 weeks starting from 1 week after sensitization with 5% PiCl. 0.9% normal saline (0.2 cc/d) was orally administered as a control.

Evaluation of Skin Severity The severity of dermatitis was assessed macroscopically in a blinded fashion according to the following scoring procedure on the 63rd day after sensitization. The total scores of the skin severity are defined as the sum of the individual score grades from score 0 (no symptoms), score 1 (mild), score 2 (moderate), and score 3 (severe) for each of the following 4 signs and symptoms, erythema (hemorrhage), edema, excoriation (erosion), and dryness (scaling).
Histological Study Mice were sacrificed on the 63rd day after sensitization. The auricular skin was removed, fixed in 10% formalin, embedded into paraffin, cut in 10-μm sections, stained with hematoxylin–eosin solution.

Measurement of Itching Behavior The mouse was individually put into a clear plastic cage and the behaviors were videotaped for 30 min on the 63rd day after sensitization. The playback served for the observation of scratching of rostral back and biting of caudal back; a series of scratching and biting behaviors were counted as one bout of event. The first 10 min was excepted from measurement as the adaptation period, the itching for the latter 20 min was counted as itching behavior.

Measurement of Serum IgE Level Blood was collected from the retro-orbital plexus of NC/Nga mice while under ether anaesthesia on the 63rd day after sensitization. Serum samples were obtained by centrifugation and stored at −80°C until assay. Serum IgE level was measured by a sandwich murine IgE ELISA kit (R&D system Inc., U.S.A.). The OD (optical density) was determined by using a microplate reader.

Detection of IFN-γ and IL-4 Levels in Lymph Nodes Lymph nodes were collected from the whole body of NC/Nga mice while under ether anaesthesia on the 63rd day after sensitization. Excised lymph nodes were homogenized in Isogen using HG 30 Homogenizer. One milliliter of homogenate was mixed with 200 μl of chloroform vigorously, and centrifuged at 13000 rpm for 15 min at 4°C using a microcentrifuge. The aqueous phase was separated and RNA in the phase was precipitated by mixing 0.5 ml of 2-propanol. The precipitate was washed with 75% ethanol and dried, and then dissolved in diethyl pyrocarbonate (DEPC)-treated water. The total RNA content was calculated based on the absorbance at 260 nm and the quality was confirmed by electrophoresis.

Reverse transcriptase-polymerase chain reaction (RT-PCR, Bioneer, Korea) was employed for the detection of mRNA. The primer sequence from 5’ to 3’ for β-Actin was, forward primer, GTG GGC CGC TAG GCA CCA, and reverse primer, CCG TTT GCC TTA GGG TTC AGG GGG G. The primer sequence from 5’ to 3’ for IFN-γ was forward primer, TAC TGCG CAC GAC GTC ATT GAA, and reverse primer, GCA GCG ACT CCT TTT CCG CTT CCT. The primer sequence from 5’ to 3’ for IL-4 was, forward primer, ACG GAG ATG GAT GTG CCA AAC GTC, and reverse primer, CGA GTA ATC CAT TTG CAT GAT GC. A mixture of 11 μl containing 1 μg RNA, DEPC-treated water and random primer was heated at 70°C for 10 min and then mixed with 4 μl of 5×First Strand Buffer, 1 μl of 10 mm deoxynucleoside triphosphate (dNTP) and 2 μl of 0.1 μM dithiothreitol. After 5 min at 25°C, 1 μl of reverse transcriptase (Superscript II) was added and RT, at 25°C for 10 min, 42°C for 50 min and then 70°C for 15 min, was performed on Tri- Thermoblock. Then, cDNA at a volume of 1 μl was mixed with 100 mM Tris–HCl (pH 8.3), 500 mM KCl, 1.5 mM MgCl2, 0.01% gelatin, 10 mM dNTP, 5 U/ml ampiTaqDNA polymerase and 1 μM primers (Stratagene) and PCR (denaturation at 94°C for 1.5 min, annealing at 62°C for 1.5 min and extension at 72°C for 1.5 min, 35 cycles) was performed on Tri-Thermoblock. Products were electrophoresed on 2% agarose gel containing ethidium bromide. The bands were recorded by Polaroid camera. Results were normalized by β-actin expression.

Comparison of Skin Severity The NC/Nga mice have been shown to develop AD-like skin lesions by repeated application of PiCl. In accordance with this previous finding, the clinical skin severity in control group increased gradually depending on the times of challenge with PiCl and reached a maximum on the 63rd day after the sensitization. All of the mice in control group exhibited AD-like skin lesions including eczema, erythema, excoriation and hemorrhage in the face, dryness, scaling and alopecia in the neck and dorsal skin (Fig. 2A). But, oral administration of SC suppressed the development of AD-like skin lesions (Figs. 2B, C).

Comparison of Serum IgE Level Serum IgE level in SC250 group decreased significantly compared to control group (<0.001) (Fig. 3). Although there was no significant difference in the clinical skin severity scores between SC100 group and SC250 group, the efficacy of SC 250 mg/kg administration was better than that of SC 100 mg/kg administration.

Comparison of Itching Behavior The total counts of itching behavior for 20 min were decreased significantly in SC administered groups compared to control group (<0.001). The itching behavior in SC250 group was significantly lowered than that in SC100 group (<0.001) (Fig. 5). Serum IgE level in SC250 group decreased significantly compared to control group (<0.001) and SC100 group (<0.05) (Fig. 6).

Comparison of IFN-γ and IL-4 in Lymph Nodes IFN-γ mRNA expression was detected in lymph nodes was detected very slightly in control group. IFN-γ mRNA expression in SC100 group increased compared to control group. IFN-γ mRNA expression in SC250 group increased compared to SC100 group (Fig. 7B). IL-4 mRNA expression in lymph nodes didn’t show significant difference between SC administered groups and control group (Fig. 7C).

Comparison of IFN-γ and IL-4 in Lymph Nodes
DISCUSSION

AD is a chronic eczematous skin disease accompanied by severe itching.\textsuperscript{30,31} We previously reported that oral administration of \textit{Rumex japonicus} HOUTT (RJH), a herb used in Eastern countries for the management of cutaneous diseases including AD, inhibits the development of AD-like symptoms in NC/Nga mice with no apparent side-effects, by suppressing the Th2 cell response.\textsuperscript{32}

SC is also an herb used for the management of various skin diseases including AD in Eastern countries. SC contains flavonoids, including quercetin, quercitrin, isoquercitrin and rutin, as pharmacologically active components. Recently, it was reported that the flavonoids addressed above could suppress inflammatory reactions \textit{in vivo}. The anti-inflammatory effect of these flavonoids suggests the possibility of their therapeutic efficacy in various inflammatory diseases.\textsuperscript{20}

In the present study, we investigated the inhibitory effects of SC on the development of AD-like skin lesions in NC/Nga

![Fig. 2. Comparison of AD-Like Skin Lesions in NC/Nga Mice after Administration of Normal Saline and SC](image1)

The photographs were taken on the 63rd day after sensitization. (A) Control group (0.9% normal saline administered, \(n=7\)), (B) SC100 group (100 mg/kg SC extract administered, \(n=8\)), (C) SC250 group (250 mg/kg SC extract administered, \(n=8\)).

![Fig. 3. Comparison of Skin Severity Score](image2)

The severity of dermatitis was assessed macroscopically in a blinded fashion according to the following scoring procedure on the 63rd day after sensitization. The total scores of the skin severity were defined as the sum of the individual score grades from score 0 (no symptoms), score 1 (mild), score 2 (moderate), and score 3 (severe) for each of the following 4 signs and symptoms, erythema (hemorrhage), edema, excoration (erosion), and dryness (scaling). Values represent the means \(\pm\) S.E.M. of each group. \(* * * p<0.001\) when compare SC100 group \((n=8)\) or SC250 group \((n=8)\) with control group \((n=7)\).

![Fig. 4. Histological Comparison of SC Administered Groups and Control Group](image3)

The auricular skins from (A) control group and (B) SC administered groups were stained with hematoxylin–eosin. (A) Surface hemorrhage, hypertrophy, intercellular edema, liquefaction degeneration of the basal layer in the epidermis and the infiltration of inflammatory cells in the dermis were observed \((100\times)\). (B) The inflammatory changes were not found in SC administered group \((100\times)\).
mice. NC/Nga mice are useful as an animal model for human AD. Skin lesions with increased numbers of eosinophils, mast cells, CD4+ T cells, and macrophages, which were clinically and histologically very similar to human AD, spontaneously developed in NC/Nga mice with a marked elevation in plasma levels of total IgE when raised in air-uncontrolled specific pathogen-free conditions. In this study we used PiCl induced AD-like skin lesions in NC/Nga mice because AD induced by repeated application of PiCl in NC/Nga mice is more valuable in its high reproducibility than spontaneous AD in NC/Nga mice.

There was a significant difference in the skin severity between SC administered groups and the control group. All of the mice in the control group exhibited AD-like skin lesions characterized by erythema, hemorrhage, excoriation, scaling, alopecia and dryness, whereas SC administered groups showed little change in the skin macroscopically (Fig. 2). In addition, the total skin severity scores were significantly reduced in SC administered groups (Fig. 3). Histologically, the control group showed hypertrophy, hyperkeratosis, intercellular edema, the liquefaction degeneration of the basal layer and infiltration of inflammatory cells in the skin, whereas SC administered groups showed a decrease in these changes (Fig. 4).

The itching behavior decreased significantly in SC administered groups compared to the control group (Fig. 5). Itching is one of the major diagnostic criteria of AD, because the itching is the most important problem for atopic patients and scratching worsens the dermatitis itself. Scratching their skin with toenails seemed to be the most important factor leading to dermatitis through increasing various immunological responses such as the elevation of serum IgE concentration and the number of mast cells. SC may play an important role in controlling the pathology of AD because it significantly suppressed the itching behavior.

In this study, the serum IgE in SC administered groups was significantly lower compared to the control group. Many studies reported that the normal value of serum IgE of NC/Nga mice in SPF condition remained under maximum 500 ng/ml, usually under 250 ng/ml, and the serum IgE level increased in NC/Nga mice with AD-like skin lesions. In NC/Nga mice, the constitutive and enhanced Jak3 phosphorylation in B cells was attributed to the high-sensitivity of CD40L and IL-4 signaling. In general, IgE synthesis by B cells is primarily regulated by cytokines. Th2 cytokines like IL-13, IL-4 play a key role for the hyperproduction of IgE, while Th1 cytokines, especially IFN-γ, are strong inhibitors of IgE synthesis, proliferation of Th2 cells, and expression of IL-4 receptors on T cells. It is well established that the elevation of serum IgE in AD may be due to the Th1/Th2 imbalance skewed to Th2, which plays important roles in the pathology of AD. Previous studies from other researchers reported that natural substances from plants or herbal therapy could inhibit the development of AD-like skin lesions in NC/Nga mice by modulating Th1 and/or Th2 cell response. Thus, we measured the levels of IFN-γ and IL-4 after SC treatment. Oral administration of SC enhanced IFN-
production, but did not inhibit IL-4 production. These results imply that the SC treatment can reduce the serum levels of total IgE by promoting Th1 cell response, especially IFN-γ. Because up-regulation of IFN-γ did not lead down-regulation of IL-4, it could be considered that IFN-γ might directly affect B cells and inhibit IgE production. Anyway, we can conclude that SC exhibits the activity that modulates the Th1/Th2 imbalance skewed to Th2 in NC/Nga mice.

Although patients with AD may benefit from herbal therapy in Oriental medicine, the possibility of hepatic toxic effects or other side effects remains a concern. However, the weight gain in NC/Nga mice during the experiment was not disturbed by SC extract administration (Table 1). In addition, the administration of SC extract appeared neither to cause tissue damage on macroscopic analysis nor to induce liver dysfunction (data not shown).

In conclusion, we demonstrated that the oral administration of SC inhibited the development of AD-like skin lesions in NC/Nga mice. The results from the present study suggest that SC could inhibit the development of AD-like skin lesions in NC/Nga mice, possibly through modulating the Th1/Th2 imbalance by the promotion of Th1 cell response. Since human is the genetically heterogeneous population, these results from the homogeneous inbred strain, NC/Nga, would be difficult to imply every human therapeutic usages. But the modulating activity of SC for the Th1/Th2 imbalance may play a role in their effectiveness for treating AD, thus SC may be an alternative substance for the management of AD. Investigations to identify the major components of SC that are responsible for the inhibitory effects on AD are currently in progress.

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REFERENCES

### Table 1. Body Weight Change

<table>
<thead>
<tr>
<th>Treatment</th>
<th>1st day (g)</th>
<th>2nd day (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>25.9±0.4</td>
<td>34.0±0.8</td>
</tr>
<tr>
<td>SC100</td>
<td>25.4±0.6</td>
<td>31.6±1.0</td>
</tr>
<tr>
<td>SC250</td>
<td>25.1±0.6</td>
<td>32.6±0.9</td>
</tr>
</tbody>
</table>

All groups were homogeneous in body weight in the beginning of experiment and the mean body weights on the day of sacrifice were not different between groups.