

Re-evaluation of Protease Activity of Reelin

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Received February 27, 2010; accepted March 4, 2010; published online March 24, 2010

Reelin is a very large secreted glycoprotein that is essential for brain formation and function, but the mechanism by which it affects the dynamics and morphology of neuronal cells remains unsolved. One previous study claimed that Reelin has a proteolytic activity against extracellular matrix proteins, which might explain many of the actions of Reelin. Therefore, in this study wild-type Reelin protein and its mutant in which a supposedly critical serine residue was replaced were expressed and tested for their self-degrading and laminin-degrading activities. We found that both of these proteins generated totally the same cleaved fragments and that neither of them is capable of degrading laminin. It is thus likely that Reelin is not a serine protease and is unable to degrade extracellular matrix.

Key words Reelin; protease; brain; extracellular matrix

Reelin is a large glycoprotein consisting of 3461 amino acid residues in mouse.¹⁾ Reelin plays central roles in brain development, such as neuronal cell migration²⁾ and dendritic growth.^{3,4)} Reelin is also implicated in adult brain function and diseases.⁵⁾ Therefore, it is important to understand how Reelin exerts its biological activity and how it is regulated.

Reelin protein is cleaved at two sites (between Reelin-repeat 2 (RR2) and RR3; N-t site, between RR6 and RR7; C-t site) *in vivo* and *in vitro*.^{6,7)} These cleavages give rise to five kinds of specific fragment, as shown in Fig. 1A. The amount of the fragment from the N-terminus to RR2 (NR2, Fig. 1A) is decreased in schizophrenia and mood disorder patients,⁸⁾ while it was increased in Alzheimer's disease patients.⁹⁾ It is therefore of physiological significance to investigate the mechanism of Reelin cleavage. Identification of the protease(s) in charge of Reelin cleavage is particularly important. So far two hypotheses have been proposed for the protease. One is that a protease(s) of the metalloprotease family other than Reelin itself cleaves Reelin.^{6,7,10)} The other is that Reelin is a serine protease and cleaves itself as well as other proteins.¹¹⁾ Most of the recent reports favor the former scenario, but the latter one has not been tested in detail and could even be compatible with the former one. We thus set out to investigate if Reelin has a serine protease activity against itself and an extracellular matrix protein laminin.

MATERIALS AND METHODS

Antibodies The mouse monoclonal anti-Reelin antibodies G10 and E5 were purchased from Chemicon (Temecula, CA; 1:2000) and from Santa Cruz Biotechnology (Santa Cruz, CA; 1:500), respectively.

Vector Construction The expression vector pCrl¹²⁾ was used to express Reelin wild-type (WT) and as a template for polymerase chain reaction. The expression vector for the Reelin mutant in which serine1283 was replaced with cysteine (Reelin1283SC) was constructed as follows. A polymerase chain reaction product harboring the point mutation was obtained by using primers CCTGTACAAGAAGAAATGCC and CAAACCGGTCTCCATCACACTTTC (the underlined codon corresponds to cysteine), and subcloned into pBluescriptII vector (Stratagene, La Jolla, CA, U.S.A.) that

had a BsrGI/AgeI fragment from pCrl. After the sequence was confirmed, the mutant fragment was subcloned into pCrl.

Cell Culture and Transfection Human embryonic kidney (HEK) 293T cells were cultured as previously described.¹⁰⁾ Transfection was performed with Lipofectamine 2000 (Invitrogen, Carlsbad, CA, U.S.A.) according to the manufacturer's instructions. After 5 h of transfection, the culture medium was replaced with serum-free Opti-MEM (Invitrogen), and cultured for another 43 h. The supernatants were filtrated through a 0.45 μ m filter (Millipore, Bedford, MA, U.S.A.). The concentration of Reelin prepared by this procedure is approximately 0.5 nM.¹³⁾

Western Blotting Samples were prepared in the sampling buffer (62.5 mM Tris, pH 6.8, 2% sodium dodecyl sulfate (SDS), 5% β -mercaptoethanol, 10% glycerol, 0.05% bromophenol blue), separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE), and transferred onto a polyvinylidene difluoride membrane (Millipore). The membrane was incubated with the indicated primary antibody and the appropriate horseradish peroxidase-conjugated secondary antibody. Images were captured using a chemiluminescent reagent (GE Healthcare, Piscataway, NJ, U.S.A.) and a LAS4000 system (Fuji, Tokyo, Japan).

Purification of Reelin The filtrated supernatants were applied to a Hitrap Heparin Sepharose HP column chromatograph set up in an AKTA system (GE Healthcare). The column was run at a flow rate of 0.25 ml/min, washed with 150 mM NaCl in Tris, pH 7.5 buffer, and eluted with a linear gradient of NaCl (150 mM to 1 M).

Laminin Degradation Assay Purified Reelin proteins were incubated with 5 μ g laminin (Invitrogen) for 0, 10, 30, 120 or 960 min at 37 °C. These samples were separated by SDS-PAGE. Separated gel was stained with silver staining II kit (Wako, Tokyo, Japan).

RESULTS

Serine1283 Is Dispensable for the Cleavage of Reelin In most serine proteases, the primary sequence surrounding the active serine residues (Gly-X-Ser-X-Gly) is conserved. Reelin contains this motif, GKSDG, at amino acid residues 1281–1285, which is located in RR3 (Fig. 1A). A previous

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- 3) Niu S., Renfro A., Quattrocchi C. C., Sheldon M., D'Arcangelo G., *Neuron*, **41**, 71—84 (2004).
- 4) Matsuki T., Pramatarova A., Howell B. W., *J. Cell Sci.*, **121**, 1869—1875 (2008).
- 5) Herz J., Chen Y., *Nat. Rev. Neurosci.*, **7**, 850—859 (2006).
- 6) Lambert de Rouvroit C., de Bergeyck V., Cortvrindt C., Bar I., Eeckhout Y., Goffinet A. M., *Exp. Neurol.*, **156**, 214—217 (1999).
- 7) Jossin Y., Gui L., Goffinet A. M., *J. Neurosci.*, **27**, 4243—4252 (2007).
- 8) Fatemi S. H., Kroll J. L., Strydom A. J., *Neuroreport*, **12**, 3209—3215 (2001).
- 9) Botella-Lopez A., Burgaya F., Gavin R., Garcia-Ayllon M. S., Gomez-Tortosa E., Pena-Casanova J., Urena J. M., Del Rio J. A., Blesa R., Soriano E., Saez-Valero J., *Proc. Natl. Acad. Sci. U.S.A.*, **103**, 5573—5578 (2006).
- 10) Kohno S., Kohno T., Nakano Y., Suzuki K., Ishii M., Tagami H., Baba A., Hattori M., *Biochem. Biophys. Res. Commun.*, **380**, 93—97 (2009).
- 11) Quattrocchi C. C., Wannenens F., Persico A. M., Ciafre S. A., D'Arcangelo G., Farace M. G., Keller F., *J. Biol. Chem.*, **277**, 303—309 (2002).
- 12) D'Arcangelo G., Nakajima K., Miyata T., Ogawa M., Mikoshiba K., Curran T., *J. Neurosci.*, **17**, 23—31 (1997).
- 13) Uchida T., Baba A., Pérez-Martínez F. J., Hibi T., Miyata T., Luque J. M., Nakajima K., Hattori M., *J. Neurosci.*, **29**, 10653—10662 (2009).
- 14) Tan K., Duquette M., Liu J. H., Lawler J., Wang J. H., *J. Mol. Biol.*, **381**, 1213—1223 (2008).
- 15) Ichihara H., Jingami H., Toh H., *Brain Res. Mol. Brain Res.*, **97**, 190—193 (2001).