Sauchinone Suppresses Pro-inflammatory Mediators by Inducing Heme Oxygenase-1 in RAW264.7 Macrophages

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Sauchinone, a biologically active lignan isolated from the roots of Saururus chinensis (Saururaceae), is reported to exert a variety of biological activities, such as hepatoprotective, anti-inflammatory actions and inhibitory effects on bone resorption. In this study, we investigated the effect of sauchinone in suppressing cyclooxygenase-2 (COX-2) and inducible nitric oxide synthase (iNOS) expression, leading to a reduction in COX-2-derived prostaglandin E2 (PGE2) and iNOS-derived nitric oxide (NO) production in lipopolysaccharide (LPS) stimulated RAW264.7 macrophages. Present study also demonstrates the effects of sauchinone in inducing heme oxygenase-1 (HO-1) expression and an increase in heme oxygenase (HO) activity in RAW264.7 macrophages. The effects of sauchinone on LPS-induced PGE2, NO, tumor necrosis factor-α (TNF-α) and interleukin-1β (IL-1β) production were partially reversed by the HO-1 inhibitor Tin protoporphyrin. We further provided evidence to support the role of HO-1 in mediation of the anti-inflammatory effects of sauchinone. Further this study provided evidence that HO-1 is a critical mechanism by which sauchinone exerts anti-inflammatory effects.

Key words sauchinone; heme oxygenase-1; pro-inflammatory mediator; RAW264.7 macrophage; extracellular signal-regulated kinase phosphorylation

MATERIALS AND METHODS

Chemicals and Reagents Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum (FBS), and other tissue culture reagents were purchased from Gibco BRL Co. (Grand, NY, U.S.A.) Tin protoporphyrin IX (SnPP IX), an inhibitor of HO activity, was obtained from Porphyrin Products (Logan, UT, U.S.A.). All other chemicals were obtained from Sigma Chemical Co. (St. Louis, MO, U.S.A.). Sauchinone (>95%) was deposited at the Standardized Material Bank for New Botanical Drugs (No. NNMBP014), Wonkwang University (Republic of Korea). Sauchinone was isolated from S. chinensis using a method described by Jeong et al. Primary antibodies, including HO-1, COX-2, iNOS and secondary antibodies were purchased from Santa Cruz Biotechnology (Heidelberg, Germany). Enzyme-linked immunosorbent

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assay (ELISA) kits for PGE$_2$, TNF-α and IL-1β were from R&D Systems, Inc. (Minneapolis, MN, U.S.A.).

**Cell Culture and Viability Assay** RAW264.7 macrophages were maintained at $5 \times 10^5$ cells per 1 ml in DMEM medium supplemented with 10% heat-inactivated FBS, penicillin G (100 unit/ml), streptomycin (100 mg/ml), and L-glutamine (2 mM) and was incubated at 37 °C in a humidified atmosphere containing 5% CO$_2$ and 95% air. For determination of cell viability, 50 mg/ml of 3-[4,5-di methylthiazol-2-yl]-2,5-diphenyltetrazolium bromide (MTT) was added to 1 ml of cell suspension ($1 \times 10^6$ cells per 1 ml in 96-well plates) for 4 h, and the formazan was dissolved in acidic 2-propanol; optical density was measured at 590 nm.

**Nitrite Assay** The nitrite concentration in the medium was measured as an indicator of NO production as per Griess reaction. One hundred microliters of each supernatant was mixed with the same volume of Griess reagent; absorbance of the mixture at 525 nm was determined with an ELISA plate reader.

**PGE$_2$ Assay** Macrophages were cultured in 24-well plates, pre-incubated for 12 h with different concentrations of sauchinone and then stimulated for 18 h with LPS. One hundred microliters of supernatant of culture medium was collected and the concentration of PGE$_2$ was determined using ELISA kit (R&D Systems, Minneapolis, MN, U.S.A.).

**TNF-α and IL-1β Assay** Macrophages were cultured in 24-well plates, pre-incubated for 12 h with various concentrations of sauchinone, and then stimulated for 18 h with LPS. Culture supernatants of culture medium were collected and the concentration of TNF-α and IL-1β was determined using ELISA kits (R&D Systems) as per manufacturer’s instructions.

**Western Blot Analysis** RAW264.7 cells were harvested and pelleted at 200 g for 3 min later washed with phosphate-buffered saline (PBS) and lysed with 20 mM Tris–HCl buffer (pH 7.4) containing protease inhibitor mixture (0.1 mM phenylmethanesulfonyl fluoride, 5 mM/ml aprotinin, 5 mg/ml pepstatin A, and 1 mg/ml chymostatin). Protein concentration was determined using the Lowry protein assay kit (P5626; Sigma). An equal amount of protein for each sample was resolved using 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and then electrophoretically transferred onto a Hybond enhanced chemiluminescence (ECL) nitrocellulose membrane (Bio-Rad, Hercules, CA, U.S.A.). The membrane was blocked with 5% skimmed milk and sequentially incubated with primary antibody (Santa Cruz Biotechnology, CA, U.S.A.) and horseradish peroxidase-conjugated secondary antibody followed by ECL detection (Amersham Pharmacia Biotech, Piscataway, NJ, U.S.A.).

**HO Activity** The method used for the determination of HO activity follows the protocol published by Motterlini et al. Briefly, after the incubation process, the cells were washed twice with PBS, gently scraped off the dish, and centrifuged (1000×g for 10 min at 4 °C). The cell pellet was suspended in MgCl$_2$ (2 mM) phosphate (100 mM) buffer (pH 7.4), frozen at −70 °C, thawed 3 times, and finally sonicated on ice before centrifugation at 18000×g for 10 min at 4 °C. The supernatant (400 μl) was added to a reduced nicotinamide adenine dinucleotide phosphate (NADPH)-generating system containing 0.8 mM NADPH, 2 mM glucose-6-phosphate, 0.2 unit glucose-6-phosphate-l-dehydrogenase, and 2 mg protein of rat liver cytosol prepared from the 15000×g supernatant fraction as a source of biliverdin reductase, potassium phosphate buffer (100 mM, pH 7.4), and hemin (10 μM) in a final volume of 200 μl. The reaction was conducted for 1 h at 37 °C in the dark and terminated by the addition of 1 ml chloroform. The extracted bilirubin was calculated by determining the difference in absorption between 464 and 530 nm.

**Statistical Analysis** Data are expressed as mean±S.D. of at least three independent experiments. To compare three or more groups, one-way analysis of variance (ANOVA) followed by Newman–Keuls post hoc test was used. Statistical analysis was performed with GraphPad Prism software, version 3.03 (GraphPad Software Inc., San Diego, CA, U.S.A.).

**RESULTS**

To determine the cytotoxic potential of sauchinone (Fig. 1A), its effect on viability of murine RAW264.7 macrophages was evaluated. Concentration of 30 μM revealed no cytotoxic effects in MTT assay. However, higher concentration showed reduced viability of the macrophages (Fig. 1B). At the non-cytotoxic concentrations (1—30 μM), RAW264.7 macrophages were challenged with LPS (1 μg/ml) in the presence or absence of the sauchinone and the levels of COX-2 and iNOS expression were measured. Pretreatment of the macrophages with sauchinone for 12 h resulted in decreased COX-2 expression (Fig. 2A) and COX-2-derived PGE$_2$ production (Fig. 2C). The PGE$_2$ production was found to have decrease by 5.1±1.7, 15.6±1.2, 46.3±0.9 and 73.4±1.1%. Under the same conditions, sauchinone also suppressed iNOS expression (Fig. 2B) and was also found to reduce the iNOS-derived NO (Fig. 2D) production (8.0±1.6, 29.1±1.2, 58.2±0.8, 88.7±0.7%). In this study, aspirin (>98% pure) was used as a positive control.

Moreover, the effects of sauchinone on the LPS-induced TNF-α and IL-1β production was examined by enzyme immunoassay in which RAW264.7 macrophages were pre-incubated with sauchinone for 12 h followed by LPS stimulation. As shown in Fig. 3, sauchinone also decreased TNF-α (17.3±1.5, 34.3±1.2, 58.0±1.0, 89.3±0.6%) and IL-1β (12.7±2.2, 27.9±2.3, 52.4±1.4, 77.6±1.1%) production in a concentration-dependent manner.
At non-cytotoxic concentrations (1—30 μM), sauchinone induced HO-1 expression in a concentration-dependent manner (Fig. 4A). In the macrophages treated with sauchinone (30 μM), HO-1 expression was first detected at 6 h, maximum increase was observed around 18 h, and reduced after 24 h (Fig. 4B). In accordance with the concentration-dependent HO-1 expression, sauchinone also increased HO activity (Fig. 4C). HO activity also showed maximal activity at 18 h after treatment (Fig. 4D).

As pre-incubation of the macrophages with sauchinone markedly inhibited LPS-induced pro-inflammatory enzymes and pro-inflammatory cytokines (Fig. 2) and since sauchinone was able to induce HO-1 expression (Fig. 4) in RAW264.7 macrophages, we examined whether sauchinone-mediated HO-1 induction could be responsible for the inhibition of COX-2-derived PGE₂, iNOS-derived NO, TNF-α and IL-1β production. RAW264.7 macrophages were pre-treated with sauchinone for 12 h in the presence of SnPP, a competitive inhibitor, followed by LPS stimulation. As shown in Fig. 5, SnPP treatment partially reversed the inhibitory effects of sauchinone on PGE₂, NO, TNF-α and IL-1β production.

We examined the effect of sauchinone on activation of MAPKs in RAW264.7 macrophages. At a concentration of 30 μM, which strongly induced the levels of HO-1, sauchinone activated the ERK pathway and increased ERK phosphorylation. As shown in Fig. 6A, phosphorylation of ERK was observed 15 min after sauchinone treatment, and was sustained up to 60 min after sauchinone treatment. In contrast, phosphorylation of JNK and p38 kinases was not seen at any time period. Furthermore, to investigate the role of MAPK in HO-1 expression, we examined the effects of specific inhibitors of ERK1/2 (PD98059), JNK (SP600125), and p38 (SB203580) on the levels of HO-1, by western blot. The ERK MAPK pathway inhibitor significantly reduced sauchinone-induced HO-1 expression, whereas the JNK and p38 inhibitors had no effect (Fig. 6B). The ERK inhibitor was not cytotoxic under our experimental conditions (data not shown).

**DISCUSSION**

Sauchinone is a unique and biologically active lignan found in *Saururus chinensis*, a perennial herbaceous plant with potential therapeutic utility in treatment of various diseases. The biological functions of sauchinone such as anti-inflammatory and immunosuppressive effects have been studied extensively. In previous studies, sauchinone is demonstrated to suppress the expression of pro-inflammatory mediators such as iNOS, COX-2 and tumor necrosis factor (TNF)
through the inhibition of nuclear factor-kappa B (NF-κB) and AP-1. However, the underlying mechanisms that could substantially explain the anti-inflammatory effect of sauchinone still remain to be elucidated. In this study, we aimed to examine the potency of sauchinone as an anti-inflammatory HO-1 inducer and its regulation of RAW264.7 macrophages. We investigated the possible involvement of HO-1 in the anti-inflammatory activities contributed by sauchinone and also examined whether sauchinone-mediated HO-1 expression correlates with inhibition of LPS-induced pro-inflammatory mediators, such as NO, PGE$_2$, TNF-α and IL-1β.

Inducible NOS (iNOS) is expressed predominantly in macrophages, that is implicated in organ destruction in some inflammatory and auto immune disorders. COX-2 enzyme is another important mediator which is the predominant isoform of cyclooxygenase at the site of inflammation. COX-2-derived PGE$_2$ plays a regulatory role in a variety of physiological and pathological processes following an immune response and inflammation. Pre-treatment with at 1—30 μM sauchinone suppressed the expression of COX-2 and iNOS in LPS-stimulated macrophages, thereby inhibiting COX-2-
derived PGE₂ as well as iNOS-derived NO production. In addition, sauchinone also inhibited LPS-induced TNF-α and IL-1β production. This finding suggests that sauchinone, at least in LPS-stimulated macrophages, exerts its anti-inflammatory effects by limiting the expression of the pro-inflammatory enzymes and inhibiting the secretion of the pro-inflammatory cytokines.

In a previous study, it was demonstrated that the protective effects of sauchinone is primarily by up-regulating heme oxygenase-1 in HepG2 cells. In this study, we found that sauchinone not only induces the HO-1 expression and increases HO activity but it also does not affect cell viability at 1—30 μM in RAW264.7 cells. The present study further examined whether sauchinone inhibited LPS-induced pro-inflammatory mediators, such as COX-2-derived PGE₂, iNOS-derived NO, TNF-α and IL-1β by increasing expression of HO-1. Our results indicate that the inhibition of HO activity by the HO inhibitor SnPP had partially reversed the inhibitory effects of sauchinone on PGE₂, NO, TNF-α and IL-1β production in LPS-stimulated macrophages (Fig. 5). Inhibition of HO-1 activity by SnPP abolished these effects, indicating that HO-1 expression plays a vital role in the inhibition of LPS-induced inflammatory responses by sauchinone. Thus, we could infer that sauchinone-induced HO-1 expression may be a key mechanism for the anti-inflammatory property exhibited by this compound.

MAPKs are a group of serine/threonine protein kinases that are activated in response to diverse extracellular stimuli that mediate signal transduction from the cell surface to the nucleus. In mammalian cells, there are three well-defined MAPK pathways: the extracellular signal-regulated kinase (ERK) pathway, the Jun N-terminal kinase (JNK) pathway and the p38 pathway. Previously, it has been observed that the MAPK pathways played a regulatory role in HO-1 gene expression. In this study, activation of the ERK pathway appeared to be involved in sauchinone-induced HO-1 expression (Fig. 6) as specific protein kinase inhibitors of ERK pathways appear to play key roles in the cyto-protection of HT22 cells. This may be due to the diverse, assortment and intensity of the complex signaling pathways activated by different inducers in diverse cell types. Sauchinone-induced HO-1 expression is directly related to the ERK pathway as ERK inhibitor PD98059 influenced the sauchinone-induced change in the HO-1 protein level, while the inhibitors of JNK and p38 pathways did not display any significant influence.

The by-products of the heme oxygenase reaction, which include free ferrous iron, carbon monoxide, and biliverdin/bilirubin, have also shown to exert a variety of biological activities. Carbon monoxide has been reported to suppress PGE₂, NO, TNF-α, and IL-1β production in LPS-stimulated macrophages through its interference of NF-κB activation. Studies have demonstrated that inflammatory stimuli induces the expression of HO-1 thereby suggesting that HO-1 expression is an adaptive cellular response to inflammation. Since sauchinone, as confirmed in this study, can actively induce the expression of the anti-inflammatory HO-1 in macrophages, possibility cannot be ruled out that anti-inflammatory effects of sauchinone could be mediated, partially, by the products of HO-1 enzyme reaction—namely, CO, bilirubin and/or biliverdin.

In conclusion, this study has demonstrated that non-cytotoxic concentration (1—30 μM) of sauchinone, actively induced HO-1 expression via ERK pathway in RAW264.7 macrophages leading to suppression of LPS-induced PGE₂, NO, TNF-α, and IL-1β production and mitigation of inflammatory process. This study strongly suggests that sauchinone may be a promising therapeutic agent for the treatment of inflammatory diseases.

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