6-Demethoxynobiletin, a Nobiletin-Analog Citrus Flavonoid, Enhances Extracellular Signal-Regulated Kinase Phosphorylation in PC12D Cells

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We previously demonstrated that nobiletin, a polymethoxylated flavone isolated from citrus peels, has the potential to improve cognitive dysfunction in patients with Alzheimer’s disease (AD). Recent studies suggest that the generation of intraneuronal amyloid-beta (Aβ) oligomers is an early event in the pathogenesis of AD. Aβ oligomers cause deficits in the regulation of the extracellular signal-regulated kinase (ERK) signaling which is critical for consolidation of the memory. Our previous studies revealed that nobiletin activated ERK signaling and subsequent cyclic AMP response element-dependent transcription. In this study, the effects of five nobiletin analogs, 6-demethoxynobiletin, tangeretin, 5-demethylnobiletin, sinensetin, and 6-demethoxytangeretin, isolated from citrus peels were assessed on ERK phosphorylation in PC12D cells, and the structure–activity relationships were examined. PC12D cells were treated with nobiletin or its analogs, and the cell extracts were analyzed by Western blotting using an antibody specific to phosphorylated ERK. 6-Demethoxynobiletin markedly enhanced ERK phosphorylation in a concentration-dependent manner. These results may be useful in developing drugs and functional foods using citrus peels for the treatment of dementia including AD.

Key words flavonoid; 6-demethoxynobiletin; nobiletin; extracellular signal-regulated kinase; Alzheimer’s disease

Alzheimer’s disease (AD) is an age-related neurodegenerative disorder characterized by progressive memory impairment and cognitive decline. Furthermore, intraneuronal soluble amyloid-beta (Aβ) oligomers have been shown to cause cognitive deficits in AD model animals. Moreover, it is suggested that the accumulation of Aβ oligomers is an early event in the pathogenesis of AD and it triggers neuronal cell dysfunction. Long-term potentiation (LTP), a form of synaptic plasticity, is believed to underlie learning and memory in the hippocampus. The cAMP-dependent signaling pathway plays a crucial role in hippocampal LTP. The extracellular signal regulated kinase (ERK) activates cAMP response element (CRE)-binding protein (CREB), a multifunctional transcription factor, which is known as the ERK/CREB cascade. Activation of ERK in mammalian hippocampus is important in consolidation of the memory. On the other hand, Aβ oligomers inhibit ERK activation and subsequently CREB in human neuroblastoma cells and primary neurons, which is prevented by an Aβ oligomer-specific antibody.

Previously, we successfully identified nobiletin, a polymethoxylated flavone found in citrus peels, as a natural compound with a unique mechanism of neurotrophic action, markedly activating protein kinase A/mitogen-activated protein kinase (MAPK)/ERK/CREB signaling and CRE-dependent transcription in cell culture systems including cultured rat hippocampal neurons. Our previous studies on the nobiletin analogs 6-demethoxynobiletin, tangeretin, 5-demethylnobiletin, sinensetin, and 6-demethoxytangeretin revealed that the compounds activate CRE-dependent transcription in the following order: nobiletin > 6-demethoxynobiletin > tangeretin. In addition, it has been reported that 6-demethoxynobiletin as well as nobiletin suppress the production of matrix metalloprotease-9/gelatinase B, which is involved in cartilage destruction and rheumatoid arthritis. However, the nobiletin analogs abilities to activate intracellular signaling in PC12D cells (a subclone of PC12 cells) are still unknown.

In this study, we examined whether nobiletin analogs enhance ERK phosphorylation and clarified the structure–activity relationships of these analogs using nobiletin as a reference compound. To the best of our knowledge, this is the first study to report that 6-demethoxynobiletin enhances ERK phosphorylation in PC12D cells.

MATERIALS AND METHODS

Preparation of Nobiletin and Its Analogs  Nobiletin and its analogs 6-demethoxynobiletin, tangeretin, 5-demethylnobiletin, sinensetin, and 6-demethoxytangeretin (Fig. 1) were isolated from the peels of Citrus depressa HAYATA as previously described previously. These compounds were dissolved in dimethyl sulfoxide (DMSO, Sigma-Aldrich Co., LLC, U.S.A.) and stored at −20°C.

Cell Culture  PC12D cells were cultured in high-glucose Dulbecco’s modified Eagle’s medium supplemented with 10% heat-inactivated horse serum (HS; Life Technologies Corp.), 5% heat-inactivated fetal bovine serum (FBS; MP Biomedicals LLC), 4 mM l-glutamine (Sigma-Aldrich Co., LLC), peni-

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cillin, and streptomycin (Meiji Seika Pharma Co., Ltd.). Cells were maintained in a humidified atmosphere containing 5% CO₂ at 37°C as described previously. 

**Western Blotting**

PC12D cells were plated at a density of 5 × 10⁵ cells/35-mm dish and incubated in the culture medium for 48 h. The culture medium was replaced with fresh low-serum medium (1% heat-inactivated FBS and 2% heat-inactivated HS) containing 30, 50, or 100 µM 6-demethoxynobiletin, tangeretin, 5-demethylnobiletin, sinensetin, and 6-demethoxytangeretin, or nobiletin (reference compound) or <0.3% DMSO (vehicle control). After stimulation, cells were lysed with lysis buffer as described previously. 

Protein assay was performed using the BCA Protein Assay Reagent (Thermo Fisher Scientific, Inc.). Western blotting was performed as described previously. 

The blotted membrane was blocked with TBST buffer (150 mM NaCl, 0.1% Tween 20, 10 mM Tris–HCl, pH 7.6) containing 5% skim milk for 1 h at room temperature. Thereafter, the membrane was incubated with phospho-ERK1/2 (Thr202/Tyr204) antibody in 5% BSA/TBST buffer overnight at 4°C and subsequently incubated with horseradish peroxidase (HRP)-conjugated anti-rabbit immunoglobulin G (IgG) for 1 h at room temperature. Following antibody stripping, the membrane was reprobed with p44/42 MAPK (Erk1/2) antibody in 5% BSA/TBST buffer overnight at 4°C and subsequently incubated with HRP-conjugated anti-rabbit IgG for 1 h at room temperature. All antibodies were obtained from Cell Signaling Technology, Inc., U.S.A. Immunoreactivity was detected using the SuperSignal® West Pico Chemiluminescent Substrate (Thermo Fisher Scientific, Inc.).

**Statistical Analysis**

Statistical analysis was performed using GraphPad Prism (GraphPad Software). Statistically significant differences were evaluated by one-way analysis of variance followed by Tukey’s multiple comparison test.

**RESULTS**

Nobiletin analogs were evaluated for their ability to stimulate ERK phosphorylation in PC12D cells (Fig. 2). 6-Demethoxytangeretin, 6-demethoxytangeretin, and tangeretin promoted ERK phosphorylation at 30 µM (Figs. 2a, b), whereas at 100 µM, 6-demethoxytangeretin and tangeretin markedly evoked ERK phosphorylation in the following order: nobiletin > tangeretin > 6-demethoxytangeretin (Figs. 2a, c).

The peak of the effect of nobiletin on ERK phosphorylation has been reported to occur at approximately 5 min. In this experiment, we examined the time-dependent effects of nobiletin analogs over a 15-min time course. 6-Demethoxytangeretin and tangeretin enhanced ERK phosphorylation in a concentration-dependent manner (Fig. 3a). Tangeretin induced ERK phosphorylation in a time-dependent manner, and this effect peaked at 5 min. On the other hand, the effect induced...
by 6-demethoxynobiletin was sustained until 15 min (Fig. 3b).

**DISCUSSION**

ERK, a type of MAPK, has been implicated as an important component in a large number of signaling systems involved in hippocampal synaptic plasticity and hippocampal-dependent memory formation, whereas Aβ oligomers are known to downregulate ERK and CREB phosphorylation in human neuroblastoma cells and primary neurons.

Nobiletin stimulates ERK phosphorylation, resulting in the stimulation of CREB phosphorylation and CRE-mediated transcription in PC12D cells. Moreover, we previously demonstrated that nobiletin improves learning and memory impairment in transgenic AD model mice, AD model rats, olfactory bulbectomized mice, and cerebral ischemia mice. Taken together, these findings suggest that nobiletin is a promising candidate for the treatment of AD.

Our previous experiments demonstrated that nobiletin induced ERK phosphorylation in a concentration- and time-dependent manner in PC12D cells. Furthermore, Furukawa et al. demonstrated the ability of tangeretin to activate ERK in rat neuron cultures. To the best of our knowledge, this is the first study to demonstrate that 6-demethoxynobiletin evokes ERK phosphorylation in a concentration-dependent manner in PC12D cells (Figs. 2, 3). It is possible that 6-demethoxynobiletin acts on the same pathway as that of nobiletin.

6-Demethoxynobiletin and its analogs evoke phosphorylate ERK at 100 µM in the following order: nobiletin > tangeretin > 6-demethoxynobiletin (Fig. 2c). Considering these results, methoxyl substituents at the 5, 6, 8, and 3' positions were proposed to be important for their ability to evoke ERK phosphorylation.

In addition, our previous studies demonstrated that PC12D cells treated with nobiletin or its analogs displayed the characteristic morphology of neurite outgrowth at 48 h after treatment in the following order: nobiletin >> 6-demethoxynobiletin. In this study, PC12D cells treated with nobiletin and 6-demethoxynobiletin for 24 h exhibited clear morphological changes (data not shown). These compounds probably exert a neurotrophic factor-like effect on PC12D cells within 24 h after treatment.

It is noteworthy that three compounds activated ERK phosphorylation and CRE-dependent transcription and that there was a correlation between them, suggesting that they may additively and/or synergistically activate CREB-mediated transcription. Although sinensetin activated CRE-dependent transcription, its ability to stimulate ERK phosphorylation was weak in PC12D cells (Fig. 2). This finding raises the possibility of an alternative pathway for CRE-dependent transcription in the action of sinensetin.

It has been demonstrated that the contents of nobiletin were 22 and 3.6 mg/kg, corresponding to approximately 55 and 8.9 µM, respectively, in the brains of mice that were intraperitoneally and orally administered 50 mg/kg nobiletin, respectively. These results suggest that nobiletin may quickly penetrate the blood–brain barrier of mice, and the in vivo concentrations of nobiletin in the brain are sufficient to activate ERK signaling. There is a possibility that the behavior of 6-demethoxynobiletin and other analogs is similar to that of nobiletin.

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**Fig. 3.** Nobiletin, Tangeretin, and 6-Demethoxynobiletin Enhanced ERK Phosphorylation in a Concentration-Dependent Manner (a)

PC12D cells were stimulated with nobiletin (10, 30, and 50 µM), tangeretin (10, 30, and 50 µM), 6-demethoxynobiletin (10, 30, and 100 µM), or vehicle control (0.3% DMSO; lane C) for 5 min. (b) Time-dependent effects of nobiletin, tangeretin, and 6-demethoxynobiletin on ERK phosphorylation. The cells were stimulated with 50 µM nobiletin, tangeretin, or 6-demethoxynobiletin for the indicated times. ERK phosphorylation was detected by Western blotting using anti-phospho-ERK1/2 and anti-ERK 1/2 antibodies.
Biological evaluation of 6-demethoxynobiletin activities such as its effects on the CREB signaling pathway is ongoing. Furthermore, it is necessary to demonstrate the effect of 6-demethoxynobiletin in animal models with memory impairment.

In conclusion, 6-demethoxynobiletin, similar to nobiletin, is a potentially useful compound in the development of functional foods for treating various forms of dementia including AD.

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