Ultrastructural Characteristics of an Established Hepatocyte Cell Line with Typical Epithelial Morphology

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ABSTRACT. An epithelial cell line (RLC1005) derived from normal fetal rat liver has been established. The cells are polygons and show intimate cell to cell adhesion. Electron microscopy showed that they are similar to hepatocytes, because of the presence of glycogen granules and of smooth surface ER, and the presence of intercellular structures similar to the bile canaliculi found in situ.

Mammalian livers have been utilized widely to test the toxicity of a variety of drugs, because detoxication is a liver-specific function. However, lack of controlled experimental conditions in vivo has restricted our understanding of the processes involved. In vitro cell culture systems, therefore, might be useful for analyzing the mechanism of drug actions. A number of successful cultures of liver cells, which retain part of the differentiative traits of livers, recently have been done (4, 6). We have focused our attention on establishing liver cell lines in vitro that retain specific characteristics of liver functions in situ. We here describe the morphology and ultrastructural characteristics of an epithelial cell line derived from fetal livers.

Liver cells were dissociated from 15-day-old fetuses of Wistar rats (5), then $10^5$ cells were inoculated into a 60 mm Falcon petri dish containing Ham's F12 medium (NISSUI) supplemented with four-fold concentrations of amino acids and 10% fetal bovine serum (Flow Laboratories). Cultures were maintained at 37°C in a humidified atmosphere of 5% CO₂ and 95% air. For the continuous subcultures, confluent cultures were trypsinized and seeded at a density of $2 \times 10^5$ cells every 10–14 days. For electron microscopy, cultures were fixed with 1.5% glutaraldehyde and 1.0% OsO₄.
Epithelial Culture from Fetal Rat Liver

in cacodylate buffer pH 7.2, and dehydrated through a series of graded ethanols, then embedded in Epon 812. The preparations for sectioning were the same as those of Eguchi and Okada (3).

Cells were polygons, granular, and adhered to each other. They proliferated actively as growing islands with extremely smooth peripheral contours that formed typical pavement-like sheets (Fig. 1). Growth curves indicated that the minimum doubling time during the logarithmic growth was about 24 h; the frequency of diploidy ($2n=42$) was about 50\%.

Electron microscopy revealed that the liver cells in these cultures had ultrastructural features similar to liver parenchyma in situ. The cells came in close contact with the entire junctional complexes of the zonula occludens and macula adherens types (Fig. 2, 6), as is typical of epithelial cells. Two characteristic features were the presence of glycogen granules and the smooth surface of the endoplasmic reticulum (Fig. 4, 5),

Fig. 2-4. 70-day cultures. Cells are rich in dilated rough ER (der) and smooth rough ER (ser), and free ribosomes are abundant. Intact Golgi complexes (gc) are located in the juxtanuclear region and a deposit of collagen-like materials (arrow) is in the intercellular spaces. ly, lysosome; m, mitochondria; mf, microfibrils. (2) $\times 21,000$, (3) $\times 20,000$, (4) $\times 16,700$

Fig. 5. 350-day culture. An accumulation of glycogen granules. $\times 24,000$
known to be ultrastructural markers of parenchyma liver cells (1). The SERs were sometimes continuous with rough surface ER and were rich in dilated rough ER filled with electron dense materials (2). They also had well-developed Golgi complexes located in the juxtanuclear region (Fig. 2, 3, 6). In addition the cytoplasm contained a large number of free ribosomes and polyribosomes (Fig. 3, 6). Besides these cell organelles, there were intercellular structures resembling the bile canaliculi (8) found in situ (Fig. 6) and a heavy deposit of collagen-like materials (7) in intercellular spaces (Fig. 3). Thus, the cell characteristics described above considered together suggest that it is that the epithelial cells in this study probably are derived from liver parenchyma.

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