Biased agonism: a novel paradigm in G protein-coupled receptor signaling observed in acquired hypocalciuric hypercalcemia

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Abstract. The classical model of G protein-coupled receptor (GPCR) activation is the two-state model, in which the GPCR exists in equilibrium between an active and inactive state. Based on this model, GPCR ligands have been classified as agonists, inverse agonists, or antagonists depending on their actions in shifting this equilibrium. Recently, however, accumulating evidence has indicated that GPCRs may exist in multiple active and inactive conformational states. In this situation, each ligand recognizes and stabilizes a specific conformation of the GPCR, leading to a set of specific biological effects. Based on this new model, a unique agonist or a combination of the usual agonist and an allosteric modulator may enable activation of a specific signaling pathway via a GPCR that activates multiple signals (biased agonism, functional selectivity). The calcium-sensing receptor autoantibody that we have identified in the serum of a patient with acquired hypocalciuric hypercalcemia (AHH) is the first example of a biased allosteric modulator of a GPCR working in a pathophysiological context. Our findings may indicate the presence of physiological allosteric modulators and provide new directions for the future drug development.

Key words: G protein-coupled receptor (GPCR), Biased agonism, Calcium-sensing receptor, Acquired hypocalciuric hypercalcemia (AHH), Protean agonism

G PROTEIN-COUPLED RECEPTORS (GPCRs) process a variety of extracellular signals related to cognition, sense, hormonal and metabolic control, cardiovascular regulation, and the immune system, and relay this information to intracellular signaling pathways [1-3]. This signaling system principally consists of GPCRs, G proteins, and effectors. GPCRs are encoded by about 1,000 genes and represent one of the largest families in the human genome. The G protein consists of Ga, Gβ, and Gγ subunits encoded separate genes from a pool of at least 16 Ga, 6 Gβ, and 12 Gγ genes. This pool potentially leads to about 1,000 variations of the G protein complex (Ga/Gβ/Gγ). G proteins regulate about 100 different effectors. In contrast to this diversity, the molecular mechanism underlying the GPCR-dependent activation of G proteins has been evolutionarily conserved among different species. For the past 20 years, many activating or inactivating mutations of GPCRs or G proteins have been reported as the underlying causes of different diseases [4-6]. Analyses of the mutants have revealed not only the molecular pathways of many diseases but also the physiological mechanisms of GPCRs.

GPCR signaling networks

GPCR signaling networks consists of GPCRs, G proteins, and effectors. In some cases, each GPCR-G protein-effector axis works independently (Fig. 1-A). In other cases, multiple GPCRs couple to one G protein thus leading to a signaling converging (Fig. 1-B). Conversely, many examples have been reported in which one GPCR couples to multiple G proteins i.e. divergent signaling (Fig. 1-C). In the classical model, it has been believed that when an agonist interacts with one GPCR that couples with multiple G proteins, multiple downstream signals related to these G proteins are activated to the same degree. To activate only one downstream signal in this model, an agonist needs to pair with a GPCR that specifically couples with only
one G protein.

A new concept, in which a unique agonist may activate a specific signaling pathway via a GPCR that involved in multiple signaling has emerged recently (Fig. 1-D). This concept is referred to as functional selectivity or biased agonism, depending on whether the focus is the receptor or the agonist [7-11]. Moreover, an allosteric modulator, which mediates the effects of an agonist by acting on a GPCR site that differs from the agonist interaction site, may augment or attenuate signals or in unique cases may activate a specific signaling pathway [12, 13] (Fig. 1-D).

If functionally selective activation is a true physiological mechanism, numerous agonist/GPCR pairs may not be required to activate specific signaling pathways and the fine-tuning of signals may be possible dependent on the cell type or context. However, this hypothesis is not compatible with the classical model of GPCR activation.

**Two-state versus multi-state model**

In the classical model of GPCR activation, it has been assumed that GPCRs exist in equilibrium between active and inactive states (Fig. 2-A). This two-state model is often used to explain many of the phenomena regarding GPCR activation and inactivation. In this model, for example, the GPCRs are activated at various frequencies without agonist stimulation. This explains why these receptors show basal activity when expressed. In this classical model, it is postulated that agonists shift the GPCR equilibrium toward the active form whereas inverse agonists do the opposite, and that antagonists, especially neutral antagonists, inhibit agonists competitively without affecting the equilibrium.

However, several lines of evidence now lend support to an alternative multi-state model for GPCRs, in which these receptors may spontaneously adopt multiple active and inactive conformational states (Fig. 2-B). In this new multi-state model, it is hypothesized that each ligand recognizes and stabilizes a specific conformation for the GPCR, leading to a set of unique and specific biological effects. A functionally selective agonist or biased agonist is defined in this case as a unique drug that recognizes and stabilizes a “chimeric” conformation that is “on” with respect to one signaling pathway and yet “off” to another. This enables the activation of a specific signaling pathway via a GPCR that activates multiple signals.

**Functional selectivity/biased agonism in GPCR pathways**

As an example that supports the multi-state model, it has been reported that in the PACAP receptor PACAP1-27 shows a higher relative efficacy for cAMP accumulation, but lower for inositol 1, 4, 5-trisphosphate (IP3) accumulation, than PACAP1-38 in LLC PK1 cells expressing PACAP receptors transiently [14]. This suggests that agonist activation of multiple signaling mechanisms is not uniform but is in fact often biased toward some but not all signaling pathways, and that agonist-selective states can produce biased agonism [15]. The findings of subsequent studies have supported this concept including ligand specific conformational changes of the β2-adrenergic receptor [16, 17], different types of agonists that induce selective coupling to a distinct second messenger pathway via the Drosophila D1-like dopamine receptor expressed in Xenopus oocytes [18], and receptor ligand specific dominancy between PLC-mediated inositol phosphate (IP) accumulation and PLA2-mediated arachidonic
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In the case of the PTH receptor that is known to activate Gs/adenyl cyclase and Gq/phospholipase C via PTH (1-34), PTH (3-34) cannot activate adenylyl cyclase even though it retains the ability to activate ERK1/2, possibly via Gq, in rat osteoblastic cells and UMR106-01 cells [20]. In tachykinin NK2 receptors expressed in HEK293 cells, NKA increases intracellular calcium and cAMP accumulation whilst the NKA (4-10) peptide increases only intracellular calcium without a cAMP response, suggestive of two distinct conformational states [21]. It must be noted in this context that it is very important to determine that both the normal agonist and the biased agonist act on the same single GPCR (Fig. 2-C, left). An alternative possibility that has to be ruled out is that the two agonists work on different GPCRs, such as closely related GPCR subtypes (Fig. 2-C, right). It will be particularly important to determine if biased agonism operates via a GPCR endogenously expressed in cells.

In the history of biased agonism in GPCR pathways, a seminal study was reported by the Bouvier group in 2003 [22]. In that study, it was demonstrated that the β2-adrenergic receptor ligands, ICI118511 and propranolol, function as inverse agonists in Gs-cAMP signaling and as partial agonists in ERK1/2 phosphorylation. The findings of this report are extremely interesting from the perspective that a unique biased ligand for one specific GPCR may work as both an agonist and an inverse agonist, depending on which signal transduction pathway is measured (Fig. 2-B).

There have been many reports showing that a GPCR activates plural G proteins. This seems to be at odds however with the aim of developing a specific signal in cells. In the past, it was believed that the localization of signaling machinery in cells may enable a specific signal event. As an alternative concept that may resolve this question, biased agonism may be a key mechanism. In a system expressing redundant and plural GPCRs, instead of expressing a specific GPCR/agonist pair which is time consuming and energy intensive, a specific biased agonist or biased allosteric modulator may enable a more rapid and flexible regulation of signals.

We have previously identified a unique autoantibody in a patient diagnosed with acquired hypocalciuric hypercalcemia (AHH) [23]. This autoantibody likely works as a biased allosteric modulator and stabilizes a unique conformation of the calcium sensing receptor (CaSR), which augments the Gq-PI turnover pathway and yet attenuates the Gi/o-ERK1/2 pathway. The characteristics of this autoantibody not only help to explain the mechanisms underlying AHH but also support the existence of biased agonism in GPCR signaling. At about the same time, several other examples of biased agonism in GPCR systems were reported including oxyntomodulin for the GLP-1 receptor [24], aripiprazole for the D2 receptor [25], carvedilol for the β2 receptor [26].

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acid release through human serotonin 2c (5-HT2c) in CHO cells [19].

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allosteric modulator, a conformation-specific allosteric antagonist has been described for the tachykinin NK2 receptor (NK2R) [27]. LPI805 promotes destabilization of the NKA-NK2R complexes that are in a conformation that triggers cAMP production, whereas the access of NKA to the conformation that triggers intracellular calcium elevation is unchanged. In contrast, among family C GPCRs such as metabotropic glutamate receptors, taste receptors and the GABA_B receptor, including a calcium sensing receptor, many positive and negative allosteric modulators have now been reported [28, 29]. In contrast to the autoantibodies we described in our AHH patient, however, most of these factors are not biased modulators.

Calcium sensing receptor — physiology and disease

Extracellular calcium ion controls its own concentration by stimulating the CaSR that is expressed in the parathyroid gland and renal tubular epithelium [30, 31]. When the serum calcium level increases, calcium-stimulated CaSR inhibits the secretion of parathyroid hormone (PTH) through a poorly defined mechanism, in which Gq/11 is believed to play a key role, and regulates the serum calcium level within a normal range. In contrast, when the serum calcium level decreases, PTH secretion is stimulated, followed by normalization of serum calcium. An antagonist acting on CaSR, a loss-of-function mutation of CaSR, and a blocking antibody to CaSR mimic hypocalcemia. A heterozygous loss-of-function mutation of CaSR causes familial hypocalciuric hypercalcemia (FHH) showing mild hypercalcemia with hypocalciuria. Brown group have reported that a blocking autoantibody to CaSR causes AHH which is a similar disease to FHH [32].

Characteristics of CaSR

CaSR belongs to the family C of GPCRs and contains a heptahelical domain, which plays a key role in G protein coupling, and a venous flytrap domain that traps the agonist. CaSR is assumed to constitutively dimerize through disulfide bonding between cystein residues 120 and 131, an event that underlies the proper functioning of this receptor. Allosteric regulation is one of the important characteristics of CaSR, similar to the taste receptors and metabotropic glutamate receptors [28, 33]. Allosteric modulators acting at six or seven transmembrane domains have been found to modulate the effects of calcium either positively or negatively. Positive allosteric modulators for CaSR are known as calciimetics, and negative allosteric modulators as calcilytics. Calciimetics in particular have been used clinically for the treatment of tertiary hyperparathyroidism in patients with end stage renal disease. Calciimetics do not activate CaSR in the absence of calcium ions, but augment the sensitivity to these ions.

An autoantibody to CaSR in an AHH patient is a biased allosteric modulator

As described earlier, we have identified and reported a unique autoantibody in a male patient, who showed mild hypercalcemia [23]. As this patient had showed normocalcemia in the past, this condition was suspected to be acquired. In one aspect in which his hypercalcemia was accompanied by hypocalciuria, his disease was similar to FHH. We suspected that he suffered from AHH caused by blocking autoantibody to CaSR, which only Brown group had reported to date. In terms of immunofluorescence, his serum was found to contain autoantibodies that reacted to CaSR expressed in HEK293 cells. We next investigated how these autoantibodies modulated calcium /CaSR-dependent signals. Surprisingly, however, the autoantibody in our patient was found to augment calcium-dependent phosphatidylinositol (PI) turnover, dependent on Gq/11, in contrast to blocking characteristics of the autoantibody reported by Brown and colleagues. Specifically, we found that the autoantibody from our patient did not stimulate PI turnover in the absence of calcium ions, but shifted the dose response curve of calcium-dependent IP accumulation to the left. This effect is similar to that exerted by calciimetics working as positive allosteric modulators of CaSR. Interestingly, the autoantibody in our patient was also found to inhibit calcium-dependent ERK1/2 phosphorylation, dependent on Gi/o (at least up to the 2.5mM calcium level). This contrasts with the effects of calciimetics that augment calcium-dependent ERK1/2 phosphorylation as well as calcium-dependent IP accumulation (Fig. 3A) (please also refer Fig. 4).

We thus revealed three main phenomena from our identification and characterization of the CaSR autoantibody in our AHH patient: (1) this CaSR autoantibody works as a positive allosteric modulator in Gq/11 signaling, but as negative allosteric modulator in Gi/o signaling, suggesting that it functions as a biased allo-
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Characteristics of newly diagnosed AHH patients

Many suspect patients of AHH have been referred to us to date, and four cases in addition to our seminal case described have been diagnosed as AHH by immunological and functional tests (unpublished, Fig. 4). Although these patients show a range of presentations from severe hypercalcemia with symptoms to mild

### Fig. 3

Autoantibodies of our AHH patient work as biased allosteric modulators of CaSR.

(A) Gq/11 vs. Gi/o. The AHH autoantibodies work as biased allosteric modulators to CaSR, positively regulating Gq/11-dependent PI turnover and yet negatively regulating Gi/o-dependent ERK1/2 phosphorylation. In contrast, a calcimimetic operates as a positive allosteric modulator to both signals. (B) Multi-states of CaSR. The data in (A) suggest that at least two different active conformations of CaSR exist. A calcium-bound form of CaSR activates both Gq/11 and Gi/o, whereas a calcium and AHH autoantibody-bound form of CaSR activates Gq/11 specifically. IPx, Inositol phosphates

### Fig. 4

The characteristics of AHH autoantibodies

Brown and colleagues reported in an AHH patient blocking autoantibodies that inhibit both PI turnover and ERK phosphorylation (2004). The autoantibodies in our patient were found to operate as biased allosteric modulators to CaSR, augmenting Gq/11-dependent PI turnover and yet inhibiting Gi/o-dependent ERK1/2 phosphorylation (2007). Brown group subsequently reported AHH autoantibodies that inhibit ERK phosphorylation without affecting PI turnover (2011). The characteristics of the autoantibodies of new AHH patients we analyzed were similar to that of the patient we reported in 2007 (unpublished).
hypercalcemia without symptoms, all of the autoantibodies in these cases were found to react to the extracellular domain (ECD) of CaSR, and to stimulate Gq/11 signals and inhibit Gi/o signals. All of our patients were elderly men with no other autoimmune disease. Moreover, in one case at least, the CaSR autoantibody was found to react to an epitope near to the flytrap domain in the ECD (unpublished data). We do not know currently whether Japanese patients are naïve to this disease or not. The key questions to be answered are in which part of the ECD is recognized by CaSR autoantibodies and how they modulate CaSR-coupling G proteins.

Protean agonism

A special case of GPCR-based functional selectivity is “protean” agonism, in which certain agonists may change or even reverse their effects depending on the states or systems adopted [9, 15, 34]. For example, in a quiescent system consisting mainly of receptors in an inactive state, some agonists may produce positive agonism. In contrast, in a constitutively active system involving a substantial spontaneously formed receptor active state, the same agonists may produce inverse agonism. This is because they convert the efficacious active state to a less efficacious ligand selective active state. Because the ligand effect changes in response to the system, these molecules were named protean agonists after the Greek sea-god Proteus (son of Poseidon), who could change shape at will, depending on his environment and needs [35]. As examples, the ligands for histamine H3 receptor (H3R) operate from full agonism to full inverse agonism, depending on the level of H3R constitutive activity [36]. Recently, we have reported that V2 receptor antagonists act as protean agonists, serving as pharmacological chaperones for inactivating V2 receptor mutants and also as inverse agonists of wild-type V2 receptor [37].

Summary

We have identified unique autoantibodies working as biased allosteric modulators of CaSR in a clinical context. Conceptually speaking, these autoantibody can switch CaSR coupling with both Gq/11 and Gi/o into CaSR coupling with Gq/11 specifically. This is therefore one clear example that the biased allosteric modulation of GPCR operates in disease. Biased agonism of GPCRs may therefore operate more generally in physiological and pathophysiological contexts. The concept of biased agonism, including protean agonism, may play a key role in developing unique new drugs as a biased agonist may switch a desirable signal “on” and an undesirable signal “off”.

References


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