A comparative analysis on the binding characteristics of various mammalian albumins towards a multitherapeutic agent, pinostrobin

Shevin R. FEROZ, Rumana A. SUMI, Sri N.A. MALEK, and Saad TAYYAB

Biomolecular Research Group, Biochemistry Programme, Institute of Biological Sciences, Faculty of Science, University of Malaya, 50603 Kuala Lumpur, Malaysia

Abstract: The interaction of pinostrobin (PS), a multitherapeutic agent with serum albumins of various mammalian species namely, goat, bovine, human, porcine, rabbit, sheep and dog was investigated using fluorescence quench titration and competitive drug displacement experiments. Analysis of the intrinsic fluorescence quenching data revealed values of the association constant, $K_a$ in the range of $1.49-6.12 \times 10^4 \text{M}^{-1}$, with 1:1 binding stoichiometry. Based on the PS–albumin binding characteristics, these albumins were grouped into two classes. Ligand displacement studies using warfarin as the site I marker ligand correlated well with the binding data. Albumins from goat and bovine were found to be closely similar to human albumin on the basis of PS binding characteristics.

Key words: animal model, drug–protein interaction, fluorescence spectroscopy, mammalian albumin, pinostrobin

Introduction

Animal models are immensely significant in drug discovery due to their importance in the characterization of disease pathophysiology, mechanism of drug action and identification of new biomarkers and drug targets [3, 23]. Information gathered from such animal studies have proved vital in establishing pharmacodynamic/pharmacokinetic relationships, determining clinical dosing regimens, as well as assessing the toxicity and safety margin of a particular drug [5, 11]. Results of in vitro and cell/tissue-based systems are not entirely representative of an in vivo response, as the metabolism and disposition properties of a drug are not taken into consideration. Thus, animal testing represents an essential intermediary phase in drug development before a potential drug can be used for human trials.

Flavonoids have emerged as one of the most promising drug candidates in recent times due to their medical significance in a wide range of diseases and ailments [9]. Furthermore, these flavonoids possess a large margin of safety as a result of their low toxicity in animals [14]. Pinostrobin (PS), a flavanone whose molecular structure is shown in Fig. 1, has been successfully tested as a potential anticancer agent based on its apoptotic activities on different cancer cell lines [20, 27, 28, 31]. In addition, PS has also been shown to confer other health benefits such as antiinflammatory [39], antiviral [38], antibacterial [22], antiulcerogenic [1] and antimutagenic [8] properties, to name a few.

As the most abundant plasma protein in the circulatory system, serum albumin functions as the main transport and depot protein for a vast variety of endogenous and exogenous ligands in the body [26]. It plays a critical role in the distribution, metabolism, efficacy and elimination of these molecules as they are significantly...
influenced by the nature and affinity of their interaction with albumin [26]. Thus, investigations into drug–albumin interactions are imperative in assessing the pharmacological potential of new drugs. Furthermore, before drug testing can be performed on human subjects, the safety and toxicity of a drug has to be evaluated using animal models that resemble as closely as possible to the human system in terms of the pharmacological aspects of drug–protein interaction. Species-dependent differences however, have been noticed with regard to the pharmacological response of various drugs/ligands [2, 4, 32, 34].

In the light of the above, we have investigated PS–albumin interaction with serum albumins from seven mammalian species, i.e., goat, bovine, human, porcine, rabbit, sheep and dog. The comparative data obtained from this study will be useful in developing an analogous model that could match certain attributes of the human system for clinical testing of new drugs.

**Materials and Methods**

**Materials**

Essentially fatty acid free serum albumins of goat (GSA), bovine (BSA), human (HSA), porcine (PSA), rabbit (RbSA), sheep (SSA) and dog (DSA); as well as warfarin (WFN) were purchased from Sigma-Aldrich Co. (St. Louis, MO). Pinostrobin (PS) was purified from *Boesenbergia rotunda* rhizomes following the published procedure [12] and its purity was established by NMR. All other chemicals used were of analytical grade purity.

**Analytical procedures**

Protein stock solutions were prepared in 10 mM sodium phosphate buffer, pH 7.4 and their concentrations except GSA and DSA were determined spectrophotometrically using \( E_{1cm}^M \) at 280 nm of 43,827 M\(^{-1}\) cm\(^{-1}\) (BSA), 35,700 M\(^{-1}\) cm\(^{-1}\) (HSA), 43,385 M\(^{-1}\) cm\(^{-1}\) (PSA and RbSA), and 42,925 M\(^{-1}\) cm\(^{-1}\) (SSA) [16]. The method of Lowry et al. [21] was employed to determine the concentrations of GSA and DSA stock solutions.

PS and WFN stock solutions were prepared by dissolving desired quantity of their crystals in 1 ml of ethanol and methanol, respectively; followed by dilution with 10 mM sodium phosphate buffer, pH 7.4 to 100 ml. A molar extinction coefficient, \( E_{1cm}^M \) of 13,610 M\(^{-1}\) cm\(^{-1}\) at 310 nm [36] was used to determine WFN concentration. All absorbance values were recorded on a Shimadzu UV-2450 double beam spectrophotometer (Shimadzu Corp., Kyoto, Japan) using quartz cuvettes of 1 cm path length.

**PS–albumin interaction**

The interactions between PS and various mammalian albumins were studied using fluorescence quench titration method as described earlier [12]. Increasing concentrations (1.5–24 \( \mu \)M) of PS were added to a fixed protein concentration (3 \( \mu \)M) in a total volume of 3 ml. 10 mM sodium phosphate buffer was used to make up the volume to 3 mL and the incubation mixture was kept for 1 h at 25 °C. Fluorescence measurements were carried out on a Jasco FP-6500 spectrofluorometer (Jasco Corp., Tokyo, Japan) using a 1 cm path length quartz cell. The protein samples were excited at 280 nm and the emission spectra were recorded in the wavelength range, 300–380 nm.

**Data analysis**

The values of the fluorescence intensity were first corrected for the inner filter effect according to the equation described by Lakowicz [19]:

\[
F_{\text{cor}} = F_{\text{obs}} \times 10^{(-A_{\text{ex}}+A_{\text{em}}/2)}
\]

where \( F_{\text{cor}} \) and \( F_{\text{obs}} \) refer to the corrected and the observed fluorescence intensity values; while \( A_{\text{ex}} \) and \( A_{\text{em}} \) are the changes in the absorbance of the samples at the excitation and emission wavelengths, respectively, produced by the addition of the ligand.

The corrected fluorescence data were then analyzed according to the following Stern-Volmer equation to determine the Stern-Volmer constant, \( K_{SV} \) and the quenching mechanism involved in PS–albumin interaction [19]:

![Molecular structure of pinostrobin.](image)
\[ \frac{F_0}{F} = K_{sv}[Q] + 1 = k_q \tau_0[Q] + 1 \]  

(2)

where \( F_0 \) and \( F \) are the fluorescence intensities in the absence and the presence of the quencher, respectively, \([Q]\) is the quencher (PS) concentration and \( k_q \) is the bimolecular quenching constant. The value of \( \tau_0 \), the fluorophore lifetime in the absence of quencher for proteins was taken as \( 10^{-8} \text{s} \) [19], which is considered as a reasonable estimate of \( \tau_0 \) for proteins in general and has been used in many previous studies on ligand–albumin interactions [10, 17, 30, 35].

The values of the association constant, \( K_a \) and \( n \), the number of binding sites were obtained using the following equation [6]:

\[
\log\left(\frac{F_0 - F}{F}\right) = n \log K_a - n \left(\log[1/([L_T] - (F_0 - F)/[P_T] / F_0)]\right)
\]

(3)

where \([L_T]\) and \([P_T]\) represent the total ligand concentration and total protein concentration, respectively. The values of \( K_a \), thus obtained, were used to calculate the free energy change of the binding reaction, \( \Delta G \) at temperature \( T \), with the help of the following relationship:

\[ \Delta G = -RT \ln K_a \]

(4)

A value of 8.3145 J mol\(^{-1}\) K\(^{-1}\) was used as the gas constant, \( R \).

**Warfarin displacement studies**

Competitive displacement experiments using WFN–albumin complexes were performed by recording the fluorescence spectra of equimolar (3 \( \mu \)M) WFN-albumin complexes in the wavelength range, 360–450 nm upon excitation at 335 nm, both in the absence and the presence of increasing PS concentrations (3–24 \( \mu \)M). The WFN-albumin mixtures were incubated for 1 h prior to the addition of PS, followed by additional 1 h incubation before emission spectral measurements.

**Results and Discussion**

PS-induced fluorescence quenching of serum albumins

Many efforts to characterize the binding of a fluorescent macromolecule to its ligand have relied on fluorescence spectroscopy to shed light on the nature of the interaction. In the case of proteins, the intensity of its fluorescence signal is significantly affected as a result of interaction with other small molecules [19].

Figure 2 shows the fluorescence spectra of seven mammalian serum albumins (3 \( \mu \)M each), namely, GSA, BSA, HSA, PSA, RbSA, SSA and DSA, recorded in the absence and the presence of increasing PS concentrations (1.5–24 \( \mu \)M). The fluorescence properties of various albumins in terms of intensity and emission maximum, in their native form are listed in Table 1. As can be seen, the fluorescence intensity varied greatly among the proteins, following the order: SSA > PSA > BSA > GSA > HSA > DSA > RbSA, whereas the emission maximum was found to lie in the range, 333–342 nm. These results were in line with the published results on the fluorescence characteristics of various serum albumins [16, 33].

It is well known that the main contributors to the fluorescence characteristics of a protein are the aromatic (mainly Trp and Tyr) amino acid residues [19]. Thus,
differences in the emission spectra of these albumins can be attributed to the number of Trp and/or Tyr residues and their location in the three-dimensional structure of the protein. However, in class B proteins, major contribution toward protein’s fluorescence is due to Trp residues [19]. The details of Trp residues in terms of their number and location in the primary structure of these albumins are given in Table 1. It is interesting to note that location(s) of Trp residues is (are) remarkably well conserved among various animal albumins. As can be seen, proteins with two Trp residues (SSA, PSA, BSA and GSA) produced higher fluorescence intensities along with higher emission maxima compared to those with single Trp residue (Fig. 2 and Table 1).

Addition of increasing PS concentrations to the protein solution produced significant quenching of the protein’s intrinsic fluorescence in all albumin species. A gradual decrease in the fluorescence intensity was observed in all albumins with the increase in PS concentration (Fig. 3). However, quantitative differences were noticed among these albumins. As evident from Fig. 3, decrease in the fluorescence intensity at 8:1 PS/albumin molar ratio was more pronounced for GSA (58%), BSA (53%) and HSA (48%) compared to the rest of the albums, showing ~30% quenching (Table 1). Such quenching in the fluorescence intensity of albums in the presence of PS was indicative of ligand–protein interaction [18].

**PS–albumin interaction: Quenching mechanism and binding parameters**

The quenching of protein fluorescence in the presence of a ligand (quencher) is known to involve either collisional or static quenching phenomena [19]. While the former refers to a process where the quencher interacts with the excited state of the fluorophore, the latter describes a mechanism in which the formation of the fluorophore excited state is inhibited by ground state complexation between the ligand and the protein [7].

In order to characterize the mode of quenching involved in the interaction between PS and serum albums, fluorescence quench titration data were analyzed according to the Stern-Volmer equation (Eq. 2). The Stern-Volmer plots, thus obtained with different PS–albumin systems are shown in Fig. 4A, while the values of the corresponding Stern-Volmer constants, *KSV* are listed in Table 2. An upward deviation in the Stern-Volmer plots was observed at higher PS concentrations (not shown), as has been reported in several earlier investigations on ligand–protein interactions [13, 24, 25, 29, 37]. Therefore, only those points falling in the initial linear zone were taken into account for regression analysis and *KSV* value determination. The gradient of the Stern-Volmer plot, the value of *KSV* connotes the extent of quenching observed. Higher values of *KSV* obtained with GSA, BSA and HSA (Table 2) correlated well with the magnitude of quenching observed with these proteins at [PS]:[Albumin] molar ratio of 8:1 (Table 1). Values of *KSV* were used to determine the bimolecular quenching constant, *kq* for different PS–albumin systems according to Eq. 2 and were found to be in the order of $10^{12}$ M$^{-1}$ s$^{-1}$. A collisional quenching process typically produces a value of *kq* in the region of $10^{10}$ M$^{-1}$ s$^{-1}$, whereas the value of *kq* larger than this diffusion-controlled limit usually indicates a binding reaction [19]. In view of the higher values of *kq* ($10^{12}$ M$^{-1}$ s$^{-1}$) obtained with different PS–albumin systems, it appears that the quenching process followed the static quenching mechanism, involving ligand–protein complex formation. We have already confirmed the phenomenon of static quenching in PS–HSA system based on inverse temper-

### Table 1. Fluorescence characteristics of different mammalian albumins in the absence and the presence of pinostrobin (PS)

<table>
<thead>
<tr>
<th>Albumin</th>
<th>Intensity (a.u.)</th>
<th>Emission maximum (nm)</th>
<th>Trp residues$^a$</th>
<th>[PS]:[Albumin] = 8:1 % Quenching$^b$</th>
</tr>
</thead>
<tbody>
<tr>
<td>GSA</td>
<td>729</td>
<td>340</td>
<td>Trp-134, Trp-213</td>
<td>58 ± 3.3</td>
</tr>
<tr>
<td>BSA</td>
<td>773</td>
<td>342</td>
<td>Trp-134, Trp-213</td>
<td>53 ± 2.9</td>
</tr>
<tr>
<td>HSA</td>
<td>407</td>
<td>338</td>
<td>Trp-214</td>
<td>48 ± 2.3</td>
</tr>
<tr>
<td>PSA</td>
<td>810</td>
<td>341</td>
<td>Trp-134, Trp-213</td>
<td>33 ± 2.6</td>
</tr>
<tr>
<td>RbSA</td>
<td>281</td>
<td>335</td>
<td>Trp-214</td>
<td>32 ± 2.8</td>
</tr>
<tr>
<td>SSA</td>
<td>838</td>
<td>341</td>
<td>Trp-134, Trp-213</td>
<td>32 ± 2.9</td>
</tr>
<tr>
<td>DSA</td>
<td>339</td>
<td>333</td>
<td>Trp-214</td>
<td>30 ± 2.7</td>
</tr>
</tbody>
</table>

$^a$ Obtained from the Universal Protein Resource (Uniprot) database at www.uniprot.org.

$^b$Expressed as mean ± SD.
ature-dependence in quenching studies [12].

Binding parameters in terms of the association constant, $K_a$ and the binding stoichiometry, $n$ for PS–albumin interaction were obtained after treating the fluorescence quenching data using Eq. 3. The use of this equation is advantageous over other methods of fluorescence data analysis due to the noninvolvement of any assumption with regard to the free and bound ligand concentrations [6]. The linear double logarithmic plots for different PS–albumin systems are shown in Fig. 4B and the values of $K_a$ and $n$, obtained from these plots are listed in Table 2. Based on the values of the association constant, different serum albumins used in this study can be classified into two distinct groups such as GSA, BSA and HSA with relatively higher PS binding affinity compared to the rest of the proteins, showing relatively lower $K_a$ values. Furthermore, GSA showed highest affinity whereas DSA was found to possess lowest binding affinity (Table 2). Interestingly, the value of $n$ for all PS–albumin complexes was found to be $\sim 1.0$, suggesting a 1:1 binding stoichiometry. The calculation of the free energy change ($\Delta G$) of the binding reaction using Eq. 4 also suggested feasibility of the binding process as $\Delta G$ was found to vary within the range of $\sim -23.8$ to $-27.3$ kJ mol$^{-1}$ (Table 2).

**PS-induced WFN displacement**

Recently we have shown Sudlow’s site I as the preferred binding site of PS on HSA [12]. In the same investigation, we have clearly demonstrated insignificant binding of PS to Sudlow’s site II, the other primary ligand binding site of HSA, even at high PS/HSA molar ratio. In view of the above, we extended our study to
different serum albumins in order to substantiate our
binding results, using WFN as a site I marker ligand.
PS-induced WFN displacement from different WFN–al
bumin complexes was studied using fluorescence
quenching. Figure 5 shows the effect of increasing PS
concentrations on the fluorescence spectra of WFN–GSA
complex. Qualitatively similar spectra were also ob-
gained with other albumin complexes (figures are omitted
for brevity). As shown in the figure, WFN–GSA complex
produced the fluorescence spectrum in the wavelength
range, 360–450 nm with an emission maximum at 383
nm, upon excitation at 335 nm. Addition of increasing
PS concentrations to WFN–GSA complex, however, led
to a progressive decrease in the fluorescence intensity,
which was suggestive of the dissociation of WFN from
its binding locus (site I) on the protein. It is important
to note that both free WFN (spectrum a) and the albumin
(spectrum b) solutions produced weak/insignificant
fluorescence signals within this range.

These fluorescence data were transformed into relative
fluorescence intensity at 383 nm by taking the fluores-
cence intensity value obtained in the absence of PS as
100 and plotted against PS concentration. Figure 6 de-
picts the decrease in the relative fluorescence intensity
at 383 nm of different WFN–albumin complexes with
increasing PS concentrations, while the values of the

Table 2. Binding parameters for the interaction of pinostrobin (PS) with different
mammalian albumins

| Albumin | $K_{37} \, (M^{-1})^a$ | $K_{a} \, (M^{-1})^a$ | n | $\Delta G \, (kJ \, mol^{-1})$
|---------|-----------------|-----------------|---|------------------|
| GSA     | $(5.62 \pm 0.35) \times 10^4$ | $(6.12 \pm 0.38) \times 10^4$ | 1.10 | $-27.3$
| BSA     | $(4.40 \pm 0.31) \times 10^4$ | $(4.82 \pm 0.34) \times 10^4$ | 1.10 | $-26.7$
| HSA     | $(3.36 \pm 0.24) \times 10^4$ | $(3.63 \pm 0.26) \times 10^4$ | 1.09 | $-26.0$
| PSA     | $(1.75 \pm 0.22) \times 10^4$ | $(1.91 \pm 0.24) \times 10^4$ | 1.05 | $-24.4$
| RbSA    | $(1.81 \pm 0.25) \times 10^4$ | $(1.95 \pm 0.27) \times 10^4$ | 1.00 | $-24.5$
| SSA     | $(1.67 \pm 0.20) \times 10^4$ | $(1.86 \pm 0.22) \times 10^4$ | 1.05 | $-24.4$
| DSA     | $(1.38 \pm 0.23) \times 10^4$ | $(1.49 \pm 0.25) \times 10^4$ | 1.04 | $-23.8$

*aExpressed as mean ± SD.

Fig. 5. Displacing effect of pinostrobin (PS) on the fluorescence spectrum of WFN–GSA complex. [GSA] = [WFN] = 3
µM, [PS] = 0–24 µM with 3 µM intervals (1–9), $\lambda_{ex}$ = 335
nm, $T$ = 25 °C. The spectra marked ‘a’ and ‘b’ refer to the
fluorescence spectra of 3 µM WFN and 3 µM GSA, re-
spectively.

Fig. 6. Quenching of WFN–albumin complex fluorescence by
pinostrobin (PS). Plots show the decrease in the relative
fluorescence intensity of different WFN–albumin com-
plexes at 383 nm with increasing PS concentrations. The
symbols used for different albumins are the same as shown
in Fig. 3.
percentage quenching achieved at the highest PS concentration (8:1 [PS]/[Albumin]) are listed in Table 3. While WFN–albumin complexes of GSA, BSA and HSA exhibited marked quenching (64–71%) at 8:1 PS/albumin molar ratio, lesser quenching (33–39%) was observed with other WFN–albumin complexes (Table 3). Interestingly, a striking positive correlation was observed between the affinity of PS towards albumin and its potential to displace WFN from Sudlow’s site I. As shown in Table 2, GSA, BSA, and HSA were also found to possess the necessary facilities.

Acknowledgments

This work was financially supported by the High Impact Research MoE Grant UM.C/625/1/HIR/MoE/SC/02 approved by the Ministry of Education, Government of Malaysia and the University of Malaya. The financial assistance from the University of Malaya to S.R.F. in the form of University of Malaya Postgraduate Research Fund (PG073/2013B) is highly appreciated. We thank the Dean, Faculty of Science and the Head, Institute of Biological Sciences, University of Malaya for providing the necessary facilities.

References


