Integral role of receptor for advanced glycation end products (RAGE) in nondiabetic atherosclerosis

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Abstract
An advanced glycation end products (AGE)/a receptor for AGE (RAGE) axis plays a central role in the pathogenesis of diabetic vascular remodeling. This study was conducted to clarify the role of RAGE in nondiabetic atherosclerosis. We used the aortic and coronary atherosclerotic lesions of Watanabe heritable hyperlipidemic (WHHL) rabbits prone to myocardial infarction (WHHLMI) at 1 to 14 months. Immunohistochemistry demonstrated the significant expression of RAGE as early as at 1 month with the stronger expression at 3 and 7 months, which was remarkably diminished at 14 months. RAGE expression was concordant with AGE accumulation. The major original sources of RAGE expression were macrophages and smooth muscle cells in addition to endothelial cells, and RAGE expression was distributed in the areas of phospholipid products, a component of oxidized LDL and nitrotyrosine. The concentrations of serum AGE did not alter significantly with aging. These findings suggested the expression of RAGE was induced by hyperlipidemia and oxidative stress independent of diabetes in WHHLMI rabbits. Additionally, our in vitro study showed that silencing of RAGE tended to attenuate oxidized-LDL-triggered PAI-1 expression in human cultured macrophages, as well as oxidized-LDL-induced tissue factor expression in peritoneal macrophages, suggesting a possible role of RAGE in prothrombogenic molecular regulation. In conclusion, the present study provides in vivo evidence that RAGE plays an integral role in the initiation and progression of nondiabetic atherosclerosis, suggesting that RAGE may be a novel target for treating not only diabetic but also nondiabetic vascular complications.

Key words: Advanced glycation end products (AGE), RAGE, Atherosclerosis, WHHLMI rabbits, Oxidized LDL
Introduction

Major risk factors such as diabetes, hyperlipidemia and hypertension are recognized to increase atherothrombotic disorders. Especially, diabetes increases the morbidity and mortality of cardiovascular diseases. There is increasing evidence that inflammation, thrombogenicity and proteolysis are associated with the progression and instability of atherosclerotic plaques in diabetes. Several mediators such as monocyte chemoattractant protein-1 (MCP-1), plasminogen activator inhibitor 1 (PAI-1), tissue factor (TF) and matrix metalloproteinase-9 (MMP-9) have been shown to contribute to the process of diabetic vascular complications. Hyperlipidemia is also a well-known risk factor for cardiovascular diseases, and lipid-lowering therapy has been shown to prevent the risk of cardiovascular diseases.

A hyperlipidemia-dependent atherosclerotic animal model, Watanabe heritable hyperlipidemic (WHHL) rabbits prone to myocardial infarction (WHHLMI), has been employed for the research of hyperlipidemic atherosclerosis.

Advanced glycation end products (AGE) are nonenzymatically glycated and oxidized modifiers of proteins and lipids. Activation of a major receptor of AGE (RAGE) enhances inflammatory cell infiltration including macrophages and T cells and oxidant stress, and an AGE/RAGE axis plays a central role in the pathogenesis of diabetic vascular complications. However, it remains to be elucidated whether RAGE mediates vascular responses in non-diabetic atherosclerosis associated with hyperlipidemia.

In this study, we attempted to clarify the role of RAGE in hyperlipidemia-dependent atherosclerosis using WHHLMI rabbits. Furthermore, the effect of the knockdown of RAGE on atherosclerosis-related molecular expression was investigated in human monocyte-derived macrophages and RAGE−/− mouse peritoneal macrophages.

Materials and Methods

Animals

Male and female WHHLMI rabbits with no sex-related differences in atherosclerosis at 1 to 14 months were bred at the Institute for Experimental Animals, Kobe University Graduate School of Medicine. New Zealand White (NZW) rabbits were obtained fromCLEA Japan Inc. (Tokyo, Japan). The animals were kept in rooms equipped with laminar-flow filters at a temperature of ≈22°C and were fed a standard rabbit chow in the Experimental Animal Laboratory of Fukushima Medical University.

The RAGE targeting construct and the generation of RAGE−/− mice were described previously. C57BL/6J wild-type and RAGE−/− mice were produced by mating C57BL/6J RAGE+/− mice, weaned at age 4-5 weeks and maintained in a temperature-controlled (22°C) facility in the Osaka City University Graduate School of Medicine. Procedures in this study were approved by the animal care and use committee at the Osaka City University Graduate School of Medicine. The experiments of RAGE−/− mice were also approved by the Animal Research Committee at Fukushima Medical University.

Study Protocol of WHHLMI Rabbits

One to 14 month-old rabbits were selected on the basis of their serum cholesterol levels, which ranged from 13.0 to 18.1 mmol/l, to make the degrees of atherosclerosis. Rabbits were anesthetized by an intravenous injection of 50 mg/kg sodium pentobarbital. The thoracic aortas and coronary arteries of the rabbits were removed and used for immunohistochemical analysis of RAGE and its cellular origins in atheromatous plaques. These experiments were carried out under the control of the Animal Research Committee in accordance with the Guidelines on Animal Experiments of Kobe University Graduate School of Medicine and Fukushima Medical University, the Japanese Government Animal Protection and Management Law (No. 105), as well as the Guide for the Care and Use of Laboratory Animals (NIH Publication No. 85-23, revised 1996).

Immunohistochemistry

Immunohistochemical staining was performed as previously described. Briefly, rabbits were anesthetized and perfused with lactated Ringer's solution and then Bouin's fixative by use of a perfusion apparatus at a constant pressure of 100 mmHg. After perfusion-fixation, the atherosclerotic lesions of the aortae and coronary arteries were excised and then immersed in Bouin's fixative for at least 24 hours. After immersion-fixation, the atherosclerotic lesions of descending thoracic aortae and coronary arteries were embedded in paraffin and cut into 4-μm-thick section. The sections were used for immunohistochemical analysis. Sections were reacted at 4°C overnight with monoclonal antibodies against rabbit macrophages (RAM11, Dako Cytomation, Carpinteria, CA) diluted 1:100, smooth muscle α-actin (1A4, 1:100 dilution, Dako Cytomation) and
endothelial cells (CD31, 1:100 dilution, Dako Cyto-
mation), phospholipid products, a component of oxi-
dized LDL (ox-LDL) (DLH3)\textsuperscript{10} diluted 1:100, AGE
(6D12, 1:100 dilution, Trans Genic Inc., Kuma-
mo, Japan) and nitrotyrosine (Kamiya Biomedical Co.,
Seattle, WA) diluted 1:100, and with polyclonal an-
tibody against RAGE (Millipore, Billerica, MA) di-
luted 1:400. A streptavidin-biotinylated horserad-
ish peroxidase system (Nichirei, T okyo, Japan) was
used and antibody binding was visualized with 3,
3'-diaminobenzidine and hydrogen peroxide (DAB
SUBSTRATE KIT FOR PEROXIDASE, Vector Lab-
atories, Burlingame, CA).

**RAGE Expression and Cellular Components of Ather-
omatous Plaques**

We defined the atheromatous lesion between
endothelial cells and internal elastic laminae of ar-
teries as plaque area under a light microscope at
magnification ×100 as described previously\textsuperscript{8,12,15}.
The percent area of RAGE expression was defined
as the RAGE positive area to the total plaque
area. In the same way, the percent areas of macro-
phages and smooth muscle cells were defined as the
RAM11-positive and 1A4-positive area to the total
plaque area, respectively. Image analysis was per-
formed to quantify the immunoreactive area using
Image J 1.34 (National Institutes of Health, Bethes-
da, MD).

**Measurement of Serum AGE Levels in WHHLMI Rab-
bbits**

The level of serum AGE was measured using
ELISA as described previously\textsuperscript{17}. Briefly, a 96-well
microtiter plate was coated by overnight incubation
at 4°C with monoclonal antibody, which detects non-
carboxymethyllysine (CML) AGE. Then 100 μl of
rabbit serum was added to each well and incubated
for 2 hours at room temperature with gentle shaking
on a horizontal rotary shaker. Immunoreactivity of
each fraction was read from the calibration curve and
was expressed as AGE unit (U) per ml, with one
unit corresponding to the amount of antibody reactive
material found in AGE-bovine serum albumin at a
protein concentration of 1 μg/ml.

**Isolation of Peripheral Blood Monocytes and Culture of
Monocyte-Derived Macrophages**

Human peripheral blood monocytes were iso-
lated by density centrifugation and adherent method
from normal healthy volunteers and the purity of
isolated monocytes was greater than 95% as deter-
mined by flow cytometry and cytohistochemistry as
described previously\textsuperscript{8,10}. Human monocytes were
plated in collagen type I-coated 6-well plate (BD
Biosciences, Bedford, MA) and incubated in a hu-
midified incubator at 37°C in RPMI 1640 medium
containing 10% fetal bovine serum (FBS) (Invitro-
gen, Carlsbad, CA) for up to 30–72 hours to differen-
tiate into macrophages. Cells were more than 90
% viable as assessed with trypan blue exclusion.
Ethical approval was obtained from Fukushima Med-
ical University for the study.

**Preparation of Oxidized LDL and Lipoprotein Defi-
cient Serum (LPDS)**

Oxidized LDL and LPDS were prepared as de-
scribed previously\textsuperscript{20}. Human LDL (density =
1.019-1.063 g/ml) and LPDS (density > 1.21 g/ml)
were isolated from serum of fasting normolipidemic
volunteers by sequential ultracentrifugation. Oxi-
dized LDL was prepared by incubating native LDL
for 24 hours at 4°C in phosphate-buffered saline (PBS)
containing 5 μmol/l CuSO\textsubscript{4}, then extensively
dialyzed against PBS and sterilized by filtra-
tion. LPDS was dialyzed against saline containing
20 mmol/l CaCl\textsubscript{2} at room temperature and against
saline to remove CaCl\textsubscript{2}. Then, the sample was in-
cubated with silicic acid at 37°C for 4 hours, adjusted
to 50 mg/ml with saline and then sterilized by filtra-
tion for use.

**Fluorescent Immunohistochemistry**

Monocytes were cultured in type I collagen-
coated chamber slides (BD BioCoat\textsuperscript{34}, BD Biosci-
ences) with RPMI 1640 medium containing 10% FBS for 72 hours. Cytospin sample slides of mono-
cytes were also prepared. Fluorescent immunohis-
tochemistry of RAGE in monocytes and monocyte-
derived macrophages was performed as described previously\textsuperscript{21}. The slides were fixed in 4% formal-
dehyde for 60 minutes and were not permeabilized,
and incubated with anti-RAGE antibody (Millipore)
for 20 minutes. After three washes in PBS, Alexa-
Fluor 594-conjugated secondary antibody (Molecular
Probes, Eugene, OR) was reacted. Stained slides
were stored in the dark until they were analyzed by
a confocal microscope (Olympus, Tokyo, Japan) with
tag (488 nm) lasers.

**Knockdown of RAGE**

RAGE expression was silenced by small inter-
ferring RNA (siRNA) 5' - CACUGCAGUCGGAGCU-
AUGG-3' (sense strand)\textsuperscript{22}. Monocyte–derived macrophages were transfected with double-strand
siRNA in serum-free medium mixed with lipo-
fectamine (Invitrogen) according to the manufacturer’s instructions. Four hours after transfection, monocyte-derived macrophages were incubated in a medium containing 2 mg/ml LPDS for 24 hours and subsequently stimulated by 50 μg/ml oxidized LDL. Alternatively, cells were treated with an irrelevant siRNA 5´-GUACCGCACGCUAUUGCACU-3´ (sense strand) as a negative control.

**Western Blotting**

The expression of RAGE, PAI-1, TF and β-actin was determined by Western blotting as described previously. Oxidized LDL (50 μg/ml) was added to nearly confluent human monocyte-derived macrophages in collagen type I-coated 6-well plates and incubated for 18 hours with or without silencing of RAGE by siRNA. The whole cells were collected and solubilized with a hypotonic lysis buffer and the protein concentrations were measured by the Bradford method. Aliquots containing 20 μg of proteins were subjected to SDS/polyacrylamide gel electrophoresis. For Western blotting, we used specific antibodies to RAGE (Santa Cruz Biotechnology Inc., CA) diluted 1 : 200, to PAI-1 (Molecular Innovations, Southfield, MI) diluted 1 : 1,000, to TF (Santa Cruz Biotechnology) diluted 1 : 200, and to β-actin (Santa Cruz Biotechnology) diluted 1 : 2,000. The signals from immunoreactive bands were visualized by an Amersham ECL system (Amersham Pharmacia Biotech UK Ltd., Buckinghamshire, England).

**Isolation of Mouse Peritoneal Macrophages**

We used RAGE−/− mice and wild-type littermate mice at the age of 7 weeks to isolate peritoneal macrophages. RAGE−/− and wild-type mice were injected with 2 ml of thioglycollate medium (Becton, Dickinson and Company, Sparks, MD) into the peritoneal cavities. After 3 days, the mice were sacrificed and injected with 5 ml PBS into peritoneal cavities to collect peritoneal macrophages. Cells were cultured with or without 5 μg/ml oxidized LDL for 18 hours in RPMI 1640 medium containing 2 mg/ml lipoprotein deficient bovine calf serum (Biomedical Technologies Inc., Stoughton, MA).

**Densitometric Analysis**

After scanning blots into a computer (CANON Canoscan 8400F, Canon Inc., Tokyo, Japan), the optical densities of individual immunoblots were analyzed using Image J 1.34 (National Institutes of Health, Bethesda, MD) as described previously.

**Statistical Analysis**

Statistical analyses were performed using ANOVA with Scheffé’s post hoc test and unpaired Student’s t-test. A level of \( P < 0.05 \) was considered significant. Data are represented as means±S.D.

**Results**

**Atheromatous Plaques with Aging**

Atheromatous plaques of the aortae of WHHL-MI rabbits were significantly found as early as at 1 month and accelerated at 3 through 14 months (Figs. 1A and 1B). The data also show the infiltration of cellular components including macrophages and smooth muscle cells (Figs. 1C and 1D). However, at 14 months, the atheromatous plaques consisted of decreased cellular components and increased extracellular fibrous and lipid components compared to 7 months (Figs. 1C and 1D), which was consistent with the previous report.

**RAGE expression**

The expression of RAGE was noted as early as at 1 month in hyperlipidemia-dependent atherosclerotic lesions and progressively increased at 3 and 7 months (Fig. 2A), and the intensive expression of RAGE in the aortae of 3 months of WHHLMI rabbits as determined by Western blotting (Fig. 2B). However, the atherosclerotic plaques at 14 months had the decreased expression of RAGE compared with 7 months (Figs. 2C and 2D).

**RAGE, AGE and Cellular Component Expression**

Fig. 3 shows the cellular components and the expression of RAGE, AGE and phospholipid products at 7 months in the atheromatous plaques by immunohistochemistry. The expression of RAGE appeared to be distributed in the areas of cellular components including endothelial cells, smooth muscle cells and macrophages. In addition, RAGE and macrophage were distributed in phospholipid products expressing area. Interestingly, AGE and RAGE were accumulated in nondiabetic hyperlipidemia-dependent atheromatous plaques.

**RAGE Expression in Coronary Atheromatous Plaques**

The components of coronary atheromatous plaques and the original sources of RAGE were similar to those of aortic plaques at 7 months. The abundant infiltration of macrophages and smooth muscle cells were found in the coronary atheroscle-
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RAGE expression was identified as the cellular sources of macrophages and smooth muscle cells in addition to endothelial cells (Figs. 4A, 4B and 4C). The data also show the quantitative analysis of the area of coronary atheromatous plaques (Fig. 4D), and RAGE expression (Fig. 4E) and its cellular sources with aging (Figs. 4F and 4G). RAGE expression was progressively increased with aging up to 7 months and markedly decreased at the age of 14 months, which was consistent with the extent of cellular infiltration.

Serum AGE Levels

Fig. 5A demonstrated the serum AGE levels at 2, 4, 8 and 15 months of WHHLMI rabbits, indicating no significant difference in the serum AGE levels among the four groups. Fig. 5B shows the serum AGE levels of WHHLMI rabbits and control NZW rabbits at 4 months. There was no significant difference between two groups. The results suggested that the expression of RAGE in hyperlipidemia-dependent atherosclerotic lesions was not related to the serum AGE levels.

AGE Accumulation and Nitrotyrosine Expression with Aging

Fig. 6A shows that the deposition of AGE was increased with aging and that AGE was predominantly recognized at the cellular components infiltrating into atheromatous plaques in WHHLMI rabbit aortae. The deposition of AGE was noted at endothelial cells as early as at 1 month. The whole expression of AGE was increased with aging at 3 and 7 months, and diminished at the age of 14 months. Moreover, immunohistochemistry demonstrated the expression of indirect oxidant marker nitrotyrosine with aging, which was similar to AGE deposition (Fig. 6B). The data suggested that AGE were pro-
duced by oxidative stress and that the profiles of AGE accumulation and nitrotyrosine expression were similar to RAGE expression.

**Role of RAGE in Atherosclerosis-Related Molecular Expression**

Fluorescent immunohistochemistry revealed the expression of RAGE on plasma membrane both of human monocytes and monocyte-derived macrophages (Figs. 7A and 7B). RAGE expression was strongly identified in macrophages cultured for 72 hours rather than for 24 hours by Western blotting (Fig. 7C), suggesting that RAGE expression was increased by the differentiation into macrophages. Fig. 7D shows that oxidized LDL increased the expression of RAGE and PAI-1 in human monocyte-derived macrophages. The silencing of RAGE in the macrophages tented to block PAI-1 expression.

In addition, oxidized LDL promoted RAGE and TF expression significantly in peritoneal macrophages from wild mice. The knockout of RAGE tented to decrease oxidized-induced TF expression in peritoneal macrophages from RAGE−/− mice (Fig. 8).

**Discussion**

We show here that RAGE expression was associated with the initiation and progression of atherosclerosis in WHHLMI rabbits, as well as the RAGE-mediated atherosclerosis-related molecular expression.

The present study clearly shows that RAGE expression is concordant with cellular infiltration of macrophages and smooth muscle cells in the atherosclerotic lesions of nondiabetic WHHLMI rabbits. This is consistent with the previous reports on nondiabetic human and animal atherosclerotic lesions. AGE are shown to be generated by oxidative stress, and Calkin et al. reported that administration of antioxidant attenuated the AGE accumulation in mice. Our data demonstrated that AGE was distributed in the areas of phospholipid products and nitrotyrosine in the atherosclerotic lesions. The present study suggested that hyperlipidemia triggered-oxidative stress may accelerate AGE genera-
tion and RAGE expression in cellular components, especially macrophages in the nondiabetic atherosclerosis.

It is well acknowledged that atheromatous RAGE expression is accelerated with high plasma glucose levels and stronger in diabetics than nondiabetics. Soro-Paavonen et al. have shown that knockout of RAGE attenuates the development of atherosclerosis in diabetic mice. These suggested a central role of RAGE in the atherosclerosis complicated with diabetes. In addition, several studies have suggested that RAGE expression is involved in inflammatory cell infiltration, thrombogenicity and plaque destability through various signaling pathways in diabetic and nondiabetic atherosclerotic lesions. Harja et al. have demonstrated that endothelial RAGE modulates vascular and inflammatory responses independent of diabetes in apoE−/− mice, suggesting the important role of RAGE in the pathogenesis of nondiabetic atherosclerosis.

We demonstrated that the significant expression of RAGE was recognized in endothelial cells as early as at 1 month in the aortic atherosclerotic lesions of WHHLMI rabbits, as well as at 3 and 7 months. This finding concurred with the results reported by Roy et al. that RAGE is expressed in the endothelial cells of the aortic atherosclerotic lesions of diabetic 2-month-WHHL rabbits.

The glucose metabolism in WHHLMI rabbits is a critical issue in this study, which was extensively investigated in adult (10- to 15-month-old) and middle-aged (17- to 21-month-old) WHHLMI rabbits. Briefly, changes in blood sugar levels were similar among the three groups, adult normal and WHHIMI rabbits and middle-aged WHHIMI rabbits. In terms of glucose tolerance, there were two groups in both adult and middle-aged WHHLMI rabbits. The rabbits with high fasting immunoreactive insulin (IRI) levels showed insulin resistance as determined by HOMA-R and Matsuda-insulin sensitivity index (ISI). However, overall, there was no significant difference in glucose tolerance among the three groups (normal adult rabbits, and adult and middle-aged WHHIMI rabbits). Also, we have the
data regarding the glucose metabolism including fasting blood sugar levels, IRI concentrations and HOMA-R in 6-month, 12-month and 18-month-old WHHIMI rabbits, demonstrating that the changes in glucose metabolism are similar among the three groups (unpublished data).

Endothelial dysfunction accelerates the development of atherosclerosis including monocyte chemotaxis, monocytes adhesion to endothelial cells and proteolysis. Moreover, MCP-1, vascular cell adhesion molecule-1 (VCAM-1) and MMP-2 are modulated by RAGE in endothelial cells under non-diabetic conditions. Taken together, our results regarding strong expression of RAGE in endothelial...
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cells suggested that endothelial RAGE may initiate and accelerate nondiabetic atherosclerosis independent of diabetes.

In the present study, we showed that RAGE expression was accelerated with cellular infiltration and that the atherosclerotic lesions with reduced cellular components at 14 months had marked decrease in RAGE expression. However, we found no significant difference in the serum AGE levels with aging, as well as between WHHLMI rabbits and
normal NZW rabbits. These results suggested that serum AGE levels were not a determinant of RAGE expression in atherosclerotic lesions.

We attempted to clarify the issue what regulates RAGE expression in nondiabetic atheromatous plaques. Since Oka et al. reported that the plasma cholesterol and oxidized LDL levels are markedly high as early as 1 month with the persistent high levels in WHHL rabbits, one possibility may be hyperlipidemia which impairs endothelial function and accumulates macrophages in plaques. Although Roy et al. found RAGE expression in atherosclerotic lesions of diabetic WHHL rabbits, we demonstrated the expression of RAGE in endothelial cells and macrophages of pure hyperlipidemia-dependent atherosclerotic animal model independent of diabetes. Our in vitro study demonstrated that oxidized LDL increased RAGE expression in macrophages.
phages. These findings suggest the regulation of RAGE expression by hyperlipidemia and oxidized LDL.

Another possibility of the mechanism of RAGE expression in nondiabetic atherosclerotic lesions may be oxidative stress\textsuperscript{28}, which nonenzymatically generates AGE. It has been shown that statin suppresses RAGE expression by decreasing oxidant stress-dependent AGE generation in human diabetic atherosclerotic plaques\textsuperscript{36}. Statin has been also reported to attenuate RAGE expression probably via decreased oxidant stress in the absence of lipid-lowering effect\textsuperscript{29,41}. In the present study, RAGE expression and AGE accumulation were associated with nitrotyrosine expression with cellular infiltration in the atherosclerotic lesions. This finding suggested that the accumulation of peroxynitrite-mediated protein oxidation reflected hyperlipidemia-dependent oxidative stress, which may induce AGE accumulation and RAGE expression in plaques.

Burke \textit{et al.} have suggested that RAGE expression may be involved in the plaque instability in diabetes since stronger RAGE expression in plaques was found in sudden death diabetic subjects than nondiabetics\textsuperscript{26}. Our study using WHHLMI rabbits suggested the possibility that the intense expression of RAGE in endothelial cells and macrophages of coronary atherosclerotic lesions may contribute to inflammation, thrombogenicity and extracellular matrix degradation in plaques, in turn, plaque destabilization. Further study is required for a better understanding of the mechanism by which RAGE signaling leads to plaque instability.

In conclusion, we show that RAGE may play an integral role in the pathogenesis of atherosclerosis in WHHLMI rabbits independent of diabetes, suggesting that RAGE may be a good target to treat diabetic and nondiabetic vascular complications.

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**Conflict of Interest**

The authors declare that they have no conflict of interest relevant to this article.

**References**

5. Cipollone F, Iezzi A, Fazia M, \textit{et al.} The receptor RAGE as a progression factor amplifying arachido-


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