Carbon Dioxide Release as an Index of Mineralization Rates of Organic Substrates

SAMBO PHEAP*
Royal University of Agriculture, Phnom Penh, Cambodia
Email: psambo@rua.edu.kh, ph.sambo@gmail.com

GINA V. PANGGA
Agricultural Systems Cluster, College of Agriculture, University of the Philippines, Las Baños, Laguna, 4013, Philippines

JOCELYN D. LABIOS
Agricultural Systems Cluster, College of Agriculture, University of the Philippines, Las Baños, Laguna, 4013, Philippines

EVALOUR T. ASPURIA
Crop Science Cluster, College of Agriculture, University of the Philippines, Las Baños, Laguna, 4013, Philippines

Received 15 November 2015  Accepted 11 April 2016  (*Corresponding Author)

Abstract Mineralization of nutrients from organic materials is vital for optimum plant growth and development. Various methods have been used to evaluate the mineralization rate of different organic substrates. Of these, carbon dioxide release is a reliable method to estimate mineralization rate. Four different substrates: dry chicken manure, Azolla, coconut coir dust (CCD), and Gliricidia sepium leaves and the combination of these substrates with SNAP solution on their rates of mineralization was determined. The results were significantly different in the amounts of CO₂ released from the substrates. CO₂ release from the substrates was stimulated by SNAP. Among all treatments, G. sepium with SNAP yielded the highest amount while coconut coir dust gave the lowest. Nitrogen mineralized to about fifty percent (50%) during the first week of all treatments. Using soil organic materials such as G. sepium leaves would quickly provide the soil with more mineralized nutrients which are available for plant growth and development.

Keywords mineralization, CO₂ release, organic substrates, Gliricidia sepium

INTRODUCTION

Mineralization is an important process which regulates the nutrient cycle (Curtin et al., 1997). It is also a continuous process which is controlled by many factors such as temperature (Curtin et al., 1997; Crohn, 2004; Gaskel and Smith, 2007; Li and Li, 2014), moisture (Curtin et al., 1997; Gaskel and Smith, 2007), pH (Curtin et al., 1997; Ouyang et al., 2008), microorganisms (Frankel and Bazylinski, 2003; Allison et al., 2009) or organisms of higher class such as earthworms (Couteaux et al., 1995), herbivores (Schrama et al., 2013), soil properties such as clay types (Deenik, 2006), substrate quality and quantity (Howard and Howard, 1993; Eiland et al., 2001; Gaskell and Smith 2007; Li and Li, 2014), and the cultivation practices.

Understanding the processes involved in mineralization, and the factors that affect mineralization rate, would help estimate the right type and amount of mineralized material to guarantee optimum yield
and minimize the adverse effects of the over application of nitrogen (N) sources (Pereira et al., 2006; Hartz et al., 2000; Crohon 2004; Bordilio et al., 2013; Li and Li, 2014).

The contents of carbon dioxide (CO₂) determine the rate of mineralization by measuring the trapped CO₂ in the alkali solution, such as Ca(OH)₂, NaOH or KOH and titrate it against hydrochloric acid (HCl) with addition of barium chloride (BaCl₂) (Peirera et al., 2006; Hartz and Britton, 2003; Makende and Ayeni, 2013).

**OBJECTIVE**

The objectives of this study were to estimate the mineralization rates of different organic materials using the CO₂ release as an index, and to examine the effects of nutrient additions on the mineralization rates of these materials.

**METHODOLOGY**

Soil samples (Alipit Clay Soil) were collected from an unfertilized upland area of the U.P. Los Baños Central Experiment Station. The soil was air-dried, cleaned from vegetative material and passed through a 2.0 mm sieve. Fifty (50) grams of this sieved soil was placed inside incubation jars and mixed with 0.125 g of each nutrient source (5 t ha⁻¹). Complete Randomized Design (CRD) with three replications was used in this experiment. The treatments were: Control (CT)-soil alone with no additional nutrients, dry chicken manure (CM), coconut coir dust (CCD), *Gliricidia sepium* (GS), *Azolla* (AZ), simple nutrient addition program (SNAP) nutrient solution, CM + SNAP, CCD + SNAP, GS +SNAP and AZ + SNAP. The incubation study was conducted from April to May 2014 in the Soil Fertility Laboratory of the Agricultural Science Cluster (ASC), College of Agriculture, U. P. Los Baños, Philippines.

The Simple Nutrient Addition Program (SNAP) nutrient solution was formulated by the Institute of Plant Breeding, College of Agriculture, U.P. Los Baños. The solution was prepared by combining 20 ml of SNAP A and 20 ml of SNAP B, and adding this to one liter of distilled water. Forty (40) ml of the prepared SNAP solution mixture was added to each SNAP-treated jar.

**Determination of Carbon Dioxide Release and Mineralized Nitrogen**

Mineralization rate was measured by determining the amount of CO₂ released from the different treatments. The incubation jar set-up was prepared prior to the start of the experiment. Prior to application, the substrates were grounded and oven-dried overnight at 40°C. The appropriate amount of residue was added to 50 g soil at a rate of 0.125 g jar⁻¹. Based on the amount of soil, this application rate approximates to 5 t ha⁻¹, the amount of residue applied in many farming systems. For the control treatment, no organic material was added in the same amount of soil. The experiment was conducted for the duration of approximately 8 weeks.

The mixtures of soil and treatment nutrient sources were supplied with moisture at field capacity. A 50 mL beaker containing 30 mL of 0.3 M NaOH was placed in the center of the jar to trap the CO₂ released from each treatment. The trapped CO₂ was then transferred to a 100 mL beaker, and 3 drops of phenolphthalein were added before titration with 0.2 M HCl.

**RESULTS AND DISCUSSION**

The incubation experiment examined the mineralization rate of different organic materials over a 7-day period. Results showed that there were significant differences between treatments. The highest amount
of CO₂ released on the first day was measured from Azolla and AZ + SNAP with 78.3 and 76.6 mg kg⁻¹ soil, respectively. These treatments were closely followed by G. sepium and CM + SNAP with 72.3 and 70.9 mg kg⁻¹ soil respectively. Since the measured CO₂ from these four treatments were ranked closely, differences of the CO₂ released were found to be insignificant.

The measured CO₂ on the second day showed that soft, fresh materials with high source of N continuously exhibited faster rate of mineralization when compared with other organic materials. Treatments with Azolla and G. sepium demonstrated the highest amount of CO₂ with 74.2 and 72.7 mg kg⁻¹ soil, respectively. The addition of nutrients hastened the mineralization rates as shown by Azolla + SNAP and GS+ SNAP. Similar effects were observed from Day 3 to Day 7 as those treatments with SNAP recorded high amount of CO₂ release. The G. sepium + SNAP Treatment dominated other treatments and their differences were found to be significant. Its performance is clearly presented in Fig. 1. Treatments with Azolla and G. sepium alone ranked second and third. It is also interesting to note that coconut coir dust played second on Day 3 to 4. Fontaine et al. (2003) reported that the increase in the amount of CO₂ released in the first week for all treatments may be attributed to the activities of microorganisms attacking the labile and readily decomposable substrates such as sugar, starch, and cellulose. The surge of CO₂ released from the soil in the first day (Fig. 1) might due to rewetting of the soil which allowed the surviving microorganisms to immediately attack SOM and the substrates (Keift et al., 1987). The control, although having a lower total CO₂ release, it had a starting point which was also as high as chicken manure and SNAP solution. The soil had high SOM content (6.3%) could increase mineralization.

![Daily carbon dioxide (mg kg⁻¹) released from different organic substrates over the period of seven days](image-url)

Franzluebbers et al. (1994) concluded that N mineralization can be related to the amount of SOM in the soil. Another study of Chudhury et al. (2014) on the effect of malic acid addition to CO₂ release found that treatments with or without nutrient addition had very high CO₂ production from 20 to 40 hours after incubation. According to Chen et al. (2014), fresh materials have faster mineralization rates and release more CO₂ G. sepium treatment was observed with consistent high amounts of CO₂ released, followed by Azolla and chicken manure.
Haney et al. (2008) also found a strong relationship between organic carbon sources and CO$_2$ release in the first day of incubation. In their study on the effect of drying and rewetting, the greatest CO$_2$ respiration lasted for three days. The reason which led to this highest rate of CO$_2$ release might be due to the active microbial biomass breaking it down when they were exposed to dryness.

The incubation experiment also examined the mineralization rate of different organic materials over an 8-week period. The results showed that there were significant differences between treatments. The highest amount of CO$_2$ released in the first week was measured from *G. sepium* + RR with 375.13 mg kg$^{-1}$ soil. It was followed by Azolla and GS with 312.84 and 312.40 mg kg$^{-1}$ soil, respectively. Notably, their values are almost equal. Azolla + SNAP and CM + SNAP rank next with 299.72 and 288.14 mg kg$^{-1}$ soil, respectively. There is a significant difference of the amount of CO$_2$ released from *G. sepium* compared the succeeding treatments ranked. In a similar study, Pangga et al. (2000) observed that more than 50% of total C has released during the first week of incubation.

A sharp decline in the amounts of CO$_2$ released was observed in Week 2. The highest amount of CO$_2$ released during this week was from CM and GS + RR, both yielding 93.28 mg kg$^{-1}$ soil. This was followed by Azolla and CM + RR with close values of 83.60 and 80.96 mg kg$^{-1}$ soil, respectively. Differences of the CO$_2$ released among treatments were found to be insignificant since the measured CO$_2$ from these four treatments were ranked closely. From Week 2 to Week 6, CM and GS + RR recorded high amounts of CO$_2$ released compared to the other treatments. However, the highest amount of CO$_2$ released during Week 7 and 8 was measured from *G. sepium*.

![Fig. 2 Weekly carbon dioxide (mg kg$^{-1}$) released over the period of eight weeks from different substrates and with the addition of SNAP solution](image)

Throughout the succession of weeks, the amounts of CO$_2$ lowered, although several peaks occurred, yet they were lower after each time. In the third week, there was an increased observation in CM+ SNAP treatment which measured 80.96 to 100.03 mg kg$^{-1}$. Then there was another peak in AZ+ SNAP (60.72 to 65.41 mg kg$^{-1}$). On the seventh week, there were three rises of CO$_2$ in GS, AZ and SNAP treatments (45.17 to 58.66, 38.72 to 55.78 and 41.65 to 45.32 mg kg$^{-1}$, respectively). Makinde and Ayeni, (2013) also observed several peaks of CO$_2$ release in their study. The decomposition of OM can be divided into three stages—(i) active (1-6 weeks), (ii) reduced or slow (7-8 weeks) and (iii) stable or moderately
stable (9-16 weeks) (Ayeni, 2011). In the active stage, the most readily available and easily decomposed substrates and OM will be decomposed. At the same time, microorganism populations also increase quickly. When substrate quality was reduced, microorganism population becomes stable or dead and decomposed to serve as the nutrient source for other microorganisms and plants.

Cumulative CO₂ release also showed that addition of substrates increased the amount of CO₂ as compared with the treatments with substrates only (Figs. 1 and 2). Fresh materials have faster mineralization rates (Chen et al., 2014), especially G. sepium which consistent high amounts of CO₂ released was observed, followed by Azolla and chicken manure. Additional nutrient sources will increase CO₂ and CH₄ production (Gogo and Pearce, 2009). Generally low-quality soil carbon limits microorganism activities as well as carbon mineralization. When additional nutrient sources are added to the soil, microorganisms will attack these sources and rapidly increase their population (Fontaine et al., 2003).

CONCLUSION

Mineralization of organic materials was about fifty percent (50%) in all substrates as compared to the amount of carbon dioxide released during the first week. The highest total CO₂ released was produced during the first week and the peak was between the first two days. The amount of CO₂ released less and less through time as the quality of the substrates eventually decreased. Using additional nutrient source (SNAP) to all organic materials resulted in faster rates of mineralization. Overall, the GS + SNAP treatment has the fastest rate of CO₂ release.

ACKNOWLEDGEMENTS

The authors extend the gratitude and appreciation to the Southeast Asian Regional Center for Graduate Study and Research in Agriculture (SEARCA), Philippines, for granting scholarship in this study; and Royal University of Agriculture (RUA) for the opportunity and providing valuable assistance in the completion of the study.

REFERENCES


Hartz, T.K. and Brittan, K. 2003. Nitrogen mineralization rate of biosolids and biosolids compost. Department of Vegetable Crops, University of California, Davis, USA.


