Successful Treatment of Refractory Thrombotic Thrombocytopenic Purpura with Cyclosporine and Corticosteroids in a Patient with Systemic Lupus Erythematosus and Antibodies to ADAMTS13

Terukazu Enami, Takeshi Suzuki, Satoshi Ito, Ai Yoshimi, Makoto Sugihara, Mizuko Mamura, Taichi Hayashi, Daisuke Goto, Isao Matsumoto, Akito Tsutsumi and Takayuki Sumida

Abstract

A 46-year-old woman with systemic lupus erythematosus was hospitalized for purpura, hematochezia and hematuria. One week after admission, she developed grand mal seizures and coma and was diagnosed with thrombotic thrombocytopenic purpura (TTP) when fragmented red cells were found on the peripheral blood smear. Laboratory findings showed severe ADAMTS13 (a disintegrin-like and metalloprotease with thrombospondin type 1 repeats) deficiency and anti-ADAMTS13 antibodies, which in recent reports have indicated a poor prognosis. She was refractory to methylprednisolone pulse therapy and plasma exchange, but administration of cyclosporine induced remission without adverse effects. We propose that cyclosporine may be an effective treatment for cases of refractory TTP.

Key words: thrombotic thrombocytopenic purpura (TTP), systemic lupus erythematosus (SLE), cyclosporine

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Introduction

Thrombotic thrombocytopenic purpura (TTP) is a rare but life-threatening disease. It is associated with thrombi, composed primarily of platelets, in affected organs. Recently, a mechanism for platelet consumption in TTP has been elucidated. Von Willebrand factor (vWF) is synthesized in endothelial cells and assembles in large multimers that are present in normal plasma. These multimers, called unusually large von Willebrand factor (ULVWF), are rapidly degraded in the circulation to normal size range VWF multimers by a specific von Willebrand factor-cleaving protease (ADAMTS-13; a disintegrin-like and metalloprotease with thrombospondin type 1 repeats) (1-4). A deficiency of ADAMTS-13 or an autoantibody directed against it is responsible for some cases of TTP, leading sequentially to accumulation of ULVWF multimers, platelet aggregation, and the thrombosis that is characteristic of the disease (5).

Based on clinical studies, plasma exchange (PE) with fresh frozen plasma is effective and is performed as an initial treatment for TTP. It is considered that PE can remove some acquired autoantibodies (such as anti-ADAMTS13) and ULVWF multimers, and plasma infusion presumably supplies the missing enzyme. However, some patients become either refractory to or dependent upon PE. Recent studies have demonstrated that adult-onset TTP patients with detectable anti-ADAMTS13 antibodies tend to have a worse prognosis than those without the antibody, with a refractory and often relapsing course (6-8). Here we report a case of refractory TTP with anti-ADAMTS13 antibodies in a patient with systemic lupus erythematosus (SLE) who was successfully treated with cyclosporine and achieved disease remission.

Case

In August 2004, a 46-year-old Japanese woman with SLE...
myasthenia gravis (MG) was admitted to our hospital because of purpura, hematochezia and hematuria. In 1990, she had been diagnosed with idiopathic thrombocytopenic purpura (ITP) and started on 30 mg/day of oral prednisolone (PSL). In 1993, she developed arthralgia with severe thrombocytopenia (platelet count 0.4×10^4/μl). She was positive for antinuclear antibody (ANA) (homogeneous 80) and anti-double stranded DNA antibody (16 U/ml), and was diagnosed with SLE. She had no significant organ involvement. Oral PSL was increased to 60 mg/day then gradually tapered. In January 1999, she began to suffer from muscle weakness of the extremities, diplopia and dysphagia. She was diagnosed with myasthenia gravis after a positive tensilon test and a positive repetitive nerve stimulation study, and athymectomy was performed. Following this she had no symptoms of SLE or MG. Her titer of anti-DNA antibody was 5 to 10 IU/ml, her lymphocytes count stayed at about 1,000/μl and her complement level was about 7 U/ml. There was no proteinuria or hematuria.

In July 2004, while on 15 mg/day of oral PSL, she developed purpura, hematochezia and hematuria, and her platelet count was 0.7×10^4/μl. Physical examination on admission showed the patient to be lucid and afebrile. Her blood pressure was 118/72 mmHg and her pulse rate 86 bpm and regular. There was mild tenderness in the upper abdomen without rebound, guarding or hepatosplenomegaly. Purpura was present in the extremities. Laboratory findings on admission revealed severe thrombocytopenia and a hemolytic anemia, with lowered haptoglobin and elevated bilirubin and lactate dehydrogenase. A coagulation screen was normal. ANA and double-stranded DNA antibodies (by RIA and ELISA methods) were slightly elevated (Table 1). Bone marrow aspirate showed increased numbers of megakaryocytes. She was considered to have an exacerbation of thrombocytopenia caused by SLE and the dose of PSL was raised from 15 mg/day to 60 mg/day, but the platelet count did not increase.

On day 7 after admission, she developed transient syncope with fever. Computed tomography scan and magnetic resonance imaging revealed no evidence of cerebral hemorrhage or infarction. Fragmented red blood cells (RBC) were observed on the peripheral blood smear. Because of fragmented red blood cells, worsening anemia and syncope, she was regarded as having TTP and was transfused with 6 units of fresh frozen plasma (FFP). Her diagnosis was confirmed with the finding of von Willebrand factor-cleaving protease (ADAMTS13; a disintegrin-like and metalloprotease with thrombospondin type 1 repeats) activity at less than 3% of normal, and an inhibitory autoantibody to ADAMTS13 was found on a blood test on day 7.

On day 8, the platelet count decreased to 0.5×10^4/μl, renal function deteriorated, creatinine was 1.2 mg/dl, proteinuria appeared, grand mal seizures occurred and she became comatose. She was ventilated and started on daily plasma exchange (PE), which improved the level of consciousness, and the platelet count increased to 1.2×10^4/μl on day 11. She was extubated in a good respiratory condition, and her proteinuria and renal function improved. However, she continued to require daily PE to maintain the platelet count.

To reduce the need for PE, she was treated with 1,000 mg/day of methylprednisolone as pulse therapy on days 18-21. On day 25 the platelet count improved to 6.0×10^4/μl. PE was stopped, but two days later the platelet count had fallen to 1.3×10^4/μl. Daily PE was resumed and administration of oral cyclosporine was started at 2.5 mg/kg/day. She was treated with methylprednisolone pulse therapy again on days 37-40 because of the slow recovery of the thrombocytopenia (Fig. 1). The platelet count was around 6.0×10^4/μl while we

<table>
<thead>
<tr>
<th>Urine</th>
<th>Blood chemistry</th>
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<tr>
<td>Protein (1+)</td>
<td>AST 32 IU/l</td>
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<tr>
<td>Sugar (-)</td>
<td>Na 141 mEq/l</td>
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<td>Occult blood (-)</td>
<td>ALT 15 IU/l</td>
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<tr>
<td>Bilirubin (-)</td>
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<td>Lym 3.8%</td>
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<td>Mo 10.5%</td>
<td>BUN 16.7 mg/dl</td>
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<tr>
<td>RBC 3.13×10⁴ /mm³</td>
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<td>haptoglobin 3 μg/dl</td>
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Table 1. Laboratory Findings on Admission

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reduced the frequency of PE. She was finally withdrawn from PE on day 67 after admission (total number of PE was 39). On day 68, the dose of cyclosporine was increased to 3.3 mg/kg/day because the trough level was low (55.8 ng/ml). Platelet count was more than 12.0×10^4/μl during tapering of oral PSL (Fig. 1).

Her proteinuria disappeared after the second series of PE and serum creatinine kinase normalized at the same time. Her titer of anti-DNA antibody (RIA) decreased to 2 U/ml on day 36 and her hematuria resolved on day 44. Her complement levels were low during hospitalization, and she had no proteinuria or hematuria. She was discharged without any clinical symptoms on the 88th day after admission. In November 2006, she is taking PSL at a dose of 12 mg/day and cyclosporine at a dose of 4.2 mg/kg/day.

Discussion

TTP was first described by Moschowitz in 1925 as a new disease characterized by the unique pathological findings of hyaline thrombi in many organs (9), and he defined the classic pentad of clinical features: thrombocytopenia, microangiopathic hemolytic anemia, neurologic symptoms and signs, renal function abnormalities and fever (10). These symptoms may, however, also be present in relapses of SLE. TTP is a rare complication of SLE, but it is important to distinguish between the two diseases because of the therapeutic implications. Detection of the fragmented peripheral RBCs helps in the early diagnosis of TTP (11). Severe ADAMTS13 deficiency and the presence of an inhibitor to ADAMTS13 may be highly specific for TTP (12, 13).

PE was initiated for a diagnosis of TTP with thrombocytopenia, microangiopathic hemolytic anemia and neurologic symptoms. Laboratory findings showed ADAMTS13 activity of less than 3%, detectable inhibitory anti-ADAMTS13 autoantibody and a normal coagulation screen. Recent reports suggest that detectable inhibitory anti-ADAMTS13 antibodies indicate a poor prognosis, a delayed response to PE, a higher plasma volume requirement to achieve complete remission, and a trend to more frequent flare-ups (6, 7). In addition, a high titer of anti-ADAMTS13 antibodies may be associated with a more advanced stage of the disease or refractory disease (8).

Our patient was treated with 1,000 mg of methylprednisolone in addition to PE because of delayed platelet count recovery. Although a transient increase in the platelet count occurred after pulse methylprednisolone therapy, she continued to be dependent on PE. A number of anecdotal reports have indicated that patients with poorly responsive disease following continued PE plus glucocorticoids may benefit from the use of more intensive immunosuppressive agents, including cyclosporine (14, 15), cyclophosphamide (16) and rituximab (17). Among these agents, cyclophosphamide, an alkylating agent that prevents cell division by cross-linking DNA strands and decreasing DNA synthesis, induces a dose-dependent injury to the hemopoietic bone marrow (18, 19). The most common adverse effects of monoclonal antibodies, including rituximab, are myelosuppression and infusion-related and hypersensitivity reactions (20, 21). In contrast, cyclosporine, which lacks clinically significant myelosuppressive activity, has an immunosuppressive effect by blocking production and release of interleukin 2 and inhibiting interleukin 2-induced activation of resting T-lymphocytes (22). Thus use of cyclosporine may represent a lower risk for patients with severe cytopenia who need immunosuppressive therapy. However, cyclosporine has not been generally used in TTP because it can itself cause TTP. Recently, anecdotal reports have indicated success following the use of cyclosporine in patients with poorly responsive disease. At relapse, reinstitution of cyclosporine treatment
can lead to a lasting response (23-26). There are no reports of the effectiveness of cyclosporine in TTP induced by infection or the hemolytic uremic syndrome. The mechanism of action of cyclosporine in TTP is unknown, but it may be mediated through one or both of two separate pathways (27, 28). It may work directly as a T cell inactivator. T cell inhibition by cyclosporine might suppress anti-ADAMTS13 antibodies and improve ADAMTS13 activity. Secondly, an inhibition by cyclosporine might suppress an ADAMTS13 an-

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