Varicella-Zoster Virus-associated Fulminant Hepatitis Following Allogeneic Hematopoietic Stem Cell Transplantation for Multiple Myeloma

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Abstract

Disseminated visceral varicella-zoster virus (VZV) infection rarely occurs in recipients of allogeneic hematopoietic stem cell transplantation (allo-HSCT). To date, only a few cases of isolated VZV-induced fulminant hepatitis following allo-HSCT have been reported. We herein describe the case of a 47-year-old Japanese man with multiple myeloma who developed fulminant hepatitis 17 months after undergoing allo-HSCT. Despite receiving fresh frozen plasma and platelet transfusions, he developed a bleeding tendency (systemic purpura, petechiae and oral bleeding), slipped into a coma and eventually died. He was retrospectively diagnosed with viscerally disseminated VZV infection based on a postmortem examination and multiplex polymerase chain reaction (PCR) amplification.

Key words: varicella-zoster virus, fulminant hepatitis, allogeneic hematopoietic stem cell transplantation


Introduction

Varicella-zoster virus (VZV) reactivation is a frequently observed opportunistic infection that develops after allogeneic hematopoietic stem cell transplantation (allo-HSCT), with a high incidence of 30-50% (1, 2). However, disseminated visceral VZV infection, presenting with ileus, abdominal pain, hepatitis or meningoencephalitis, rarely occurs in recipients of allo-HSCT (3-14). To the best of our knowledge, only a few cases of isolated VZV-induced fulminant hepatitis following allo-HSCT have been reported to date (15-17). We herein report the clinical course of an allo-HSCT patient with multiple myeloma who developed fulminant hepatitis 17 months after undergoing allo-HSCT and was diagnosed with viscerally disseminated VZV infection on a postmortem examination.

Case Report

A 47-year-old Japanese man was diagnosed with Durie/Salmon stage IIIA IgD-λ multiple myeloma. No chromosomal abnormalities were detected on a conventional cytogenetic analysis of the patient’s bone marrow. Fluorescent in situ hybridization was not performed. The patient received three courses of vincristine, doxorubicin and dexamethasone therapy, resulting in a partial response. Subsequently, he received high-dose melphalan followed by an infusion of autologous peripheral blood stem cells mobilized with high-dose cyclophosphamide and granulocyte colony-stimulating factor. The patient’s history of exposure to varicella and zoster was unknown.

Seven months after the autologous transplantation, the patient underwent reduced-intensity stem cell transplantation (RIST) as an institutional trial approved by the Akita University Research Ethics Board. Informed consent was ob-
tained from the patient before performing RIST, according to institutional guidelines. A peripheral blood stem cell graft was obtained from an HLA-matched sibling donor. The conditioning regimen included 30 mg/m² of fludarabine administered for five days and 4 mg/kg of busulfan administered for two days. The graft-versus-host disease (GVHD) prophylaxis consisted of cyclosporine and a short course of methotrexate. The patient was administered 1,000 mg/day of acyclovir starting on day -12 before RIST to day 54 after RIST for prophylaxis against herpes simplex virus infection. Engraftment was prompt without the development of acute GVHD, and the dose of cyclosporine was slowly tapered.

As one of the treatments, the laboratory data continued to indicate severe liver dysfunction, with AST, ALT and fibrinogen levels of 9,600 IU/L, 4,200 IU/L and 41.8 mg/dL, respectively. The patient developed a bleeding tendency (systemic purpura, petechiae and oral bleeding), slipped into a coma with hyperammonemia (145 mM) on day 524 and died on day 525. Neither brain CT nor lumbar puncture were performed. Multiplex polymerase chain reaction (PCR) amplification for human herpes viruses in a blood sample obtained on day 524 retrospectively revealed the presence of VZV DNA. With the consent of the patient’s family, an autopsy of the liver was performed. The liver exhibited extensive foci of necrosis throughout the parenchyma, and an immunohistopathological examination revealed most hepatic cells to be positive for VZV.

**Discussion**

In the present case, VZV infection was retrospectively confirmed on a pathological examination and multiplex PCR analysis. To our knowledge, only a few cases of isolated VZV fulminant hepatitis developing after allo-HSCT have been reported to date (15-17), and disseminated visceral VZV infection presenting with ileus, abdominal pain, hepatitis and/or meningoencephalitis rarely occurs in recipients of allo-HSCT (3-14). The occurrence of GVHD and the duration of the initial antiviral prophylaxis are known to be significant and independent risk factors for the development of VZV after allo-HSCT (1, 18), with a pretransplant diagnosis of a lymphoproliferative disorder being another relevant risk factor (2). In the present case, the relapsed myeloma that developed after allo-HSCT led to profound immunodeficiency due to the presence of chronic GVHD and repeated chemotherapy as well as the myeloma itself; thus, the patient was at a high risk for VZV and other opportunistic infections.

Although acyclovir is commonly used during the first year after HSCT for prophylaxis against herpes simplex virus and VZV infection with high efficacy, the appropriate dose and duration of prophylaxis and the point of termination of acyclovir treatment for patients at a high risk for VZV reactivation remain unclear. Prophylaxis has been shown to reduce the incidence of VZV infection only while the administration of such therapy continues, and the incidence of VZV infection at four years post-transplantation is
not modified by a 1-year administration of prophylaxis (19). In allo-HSCT recipients with risk factors for VZV infection, long-term prophylactic acyclovir treatment may be required to prevent a fatal VZV infection until the patient’s immune system is successfully reconstituted. Thomson et al. attempted to resolve the issue of the appropriate duration of continued VZV prophylaxis after allo-HSCT (20). In their study, HSCT recipients received low-dose acyclovir (400 mg/day) until all immunosuppressants were discontinued and the CD4 T-cell count exceeded 200/mm$^3$. Although acyclovir prophylaxis prevented early VZV reactivation, the long-term incidence of the disease did not appear to be affected, as infection occurred once prophylaxis was discontinued, even after the CD4 T-cell count exceeded 200/mm$^3$.

In contrast, Asano-Mori et al. reported that long-term prophylaxis consisting of ultra-low-dose acyclovir at 200 mg/day continued until the end of immunosuppressive therapy and for at least one year after transplantation resulted in successful prevention of severe VZV-related symptoms and death, with a significantly decreased overall incidence of VZV reactivation (21). Moreover, with this prophylaxis, visceral dissemination was completely eliminated (21). If our patient had received ultra-low-dose acyclovir prophylaxis until the end of immunosuppressive therapy, it is possible that the fulminant hepatitis could have been prevented.

Recently, the efficacy of 1-year low-dose valacyclovir prophylaxis therapy against VZV infection was prospectively evaluated, wherein visceral involvement and serious complications were completely eliminated for two years after HSCT (22). The inactivated varicella vaccine reduces the risk of VZV reactivation in autologous HSCT patients with lymphoma (23). Although there is controversy surrounding the use of the live attenuated varicella vaccine in allo-HSCT recipients, it is possible that reconstructing the host immunity with the VZV vaccine would reduce the frequency of clinical VZV infection in the period after allo-HSCT. The development of a new prophylactic strategy for treating high-risk patients, including a protocol for VZV vaccination, is needed to prevent fatal VZV infections following allo-HSCT.

When VZV disseminates into organs without any cutaneous manifestations, the patient is frequently not treated in a timely manner due to the difficulty in diagnosing the infection. Therefore, disseminated visceral VZV infection, particularly isolated fulminant hepatitis, has an extremely poor prognosis (15-17). Recently, Okamoto et al. reported the successful treatment of severe hepatitis associated with varicella zoster virus infection in a patient with diffuse large B-cell lymphoma treated with rituximab plus combination chemotherapy (24). They initiated treatment with high-dose acyclovir, immunoglobulin and thrombomodulin-alpha and successfully rescued the patient. VZV infection should be considered in patients with unexplained liver dysfunction under severe immunosuppressive conditions, even in the absence of viral exposure and/or skin involvement, and appropriate treatment against VZV infection should be started immediately as empiric therapy due to the high mortality of the disease.

In summary, we herein reported the clinical course of an allo-HSCT patient with multiple myeloma who developed fulminant hepatitis 17 months after undergoing allo-HSCT and who was diagnosed with visceral VZV infection on a postmortem examination. Unfortunately, the patient was not treated in a timely manner due to the difficulty in diagnosing the infection, and he died from VZV infection-induced hepatic failure. In order to successfully treat VZV infections in immunocompromised patients, it is essential to start antiviral therapy with acyclovir empirically when unexplained liver dysfunction occurs after conducting a meticulous physical examination and prompt clinical laboratory tests. The extended use of acyclovir prophylaxis, based on an evaluation of risk factors for VZV, until the end of immunosuppressive therapy should be carefully considered in allo-HSCT patients.

The authors state that they have no Conflict of Interest (COI).

References

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**Figure.** A. The left panel indicates extensive foci of hepatocyte necrosis throughout the parenchyma. Hematoxylin and Eosin staining; magnification, ×100. B. The right panel indicates the reaction product of staining with anti-varicella-zoster virus (VZV) antibodies (MAB8612, IgG2b, Chemicon, Millipore corporation, Billerica, MA, USA). Immunohistochemical staining; magnification, ×100.


