Histological Transformation to Large Cell Neuroendocrine Carcinoma from Lung Adenocarcinoma Harboring an *EGFR* Mutation: An Autopsy Case Report

Rika Moriya¹, Satoshi Hokari¹, Satoshi Shibata¹, Takeshi Koizumi¹, Takafumi Tetsuka²,
Kazuhiko Ito², Hideki Hashidate³ and Hiroki Tsukada²

Abstract

We herein report a 58-year-old Japanese woman who survived 14 years after surgery for lung adenocarcinoma harboring an epidermal growth factor receptor (*EGFR*) exon 19 deletion. She developed recurrence, for which she underwent multimodal therapy, including *EGFR*-tyrosine kinase inhibitor (TKI) administration. She ultimately died from a rapidly progressive right lung tumor that was resistant to *EGFR*-TKI. According to the autopsy findings, she had combined large-cell neuroendocrine carcinoma (LCNEC) and adenocarcinoma in the right lung, which retained an *EGFR* exon 19 deletion in both components. Therefore, the histological transformation to LCNEC can be a mechanism of acquired *EGFR*-TKI resistance.

Key words: large-cell neuroendocrine carcinoma, adenocarcinoma, epidermal growth factor receptor tyrosine kinase inhibitor, histological transformation

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Introduction

Epidermal growth factor receptor (*EGFR*) mutations are present in from 10-20% of all non-small cell lung cancers (NSCLCs) and predict a response to *EGFR*-tyrosine kinase inhibitors (TKIs) (1). Although *EGFR*-TKIs are an effective treatment for these carcinomas, most patients ultimately acquire resistance (2). Recent studies have elucidated some of the mechanisms underlying *EGFR*-TKI resistance, including pathological transformation into small cell lung cancer (SCLC) (3). We herein report a rare case of a patient with adenocarcinoma that transformed into large-cell neuroendocrine carcinoma (LCNEC) at the time that *EGFR*-TKI resistance developed.

Case Report

A 58-year-old Japanese woman presented to our institution with a chest radiograph abnormality in 1999. She was a

life-long non-smoker with no previous medical problems. A physical examination and laboratory data were unremarkable. A computed tomography (CT) scan revealed a 25-mm nodule in the left S8 of the lung. Based on the subsequent workup, this was classified as cT1bN0M0, stage IB (TNM classification 7th edition) lung adenocarcinoma. She underwent a left lower lobe lobectomy and was diagnosed with well-differentiated lung adenocarcinoma. However, mediastinal lymph node metastases were pathologically detected. She was ultimately diagnosed with pT1bN2M0, stage IIIA (TNM classification 7th edition) lung adenocarcinoma. In the retrospective examination, the surgical specimen showed an *EGFR* exon 19 deletion. She was administered adjuvant chemotherapy with cisplatin, mitomycin, and vindesine, and then was changed to an oral tegafur-uracil regimen because of the side effects associated with the initial regimen.

In March 2002, she developed bilateral multiple intrapulmonary metastases and was given carboplatin and paclitaxel. Gefitinib was then administered in November 2002. During a follow-up examination in 2004, brain metastases were

¹Department of Respiratory Medicine and Infectious Diseases, Niigata University Graduate School of Medical and Dental Sciences, Japan, ²Department of Respiratory Medicine, Niigata City General Hospital, Japan and ³Department of Pathology, Niigata City General Hospital, Japan
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Correspondence to Dr. Satoshi Hokari, g1jockey@nifty.com

Figure 1. Computed tomography (CT) scans obtained in 2010. (A) A chest CT scan revealed intrapulmonary metastases in both lungs. (B, C) An abdominal CT scan revealed a huge right adrenal metastasis (arrowheads).

identified by magnetic resonance imaging, and the metastases were treated with brain gamma knife radiosurgery that was repeated each time new lesions emerged (seven times in total). In 2010, a CT scan revealed that her intrapulmonary metastases had progressed (Fig. 1A). Although she was administered docetaxel while receiving gefitinib treatment, a right adrenal metastasis was detected (Fig. 1B and C). She was therefore administered erlotinib in July 2010, and the disease stabilized. Our biggest concern was that the massive adrenal metastasis might rupture or cause symptoms associated with increased pressure, and we therefore performed right adrenal resection in January 2011. The immunohistochemical results revealed that it was a metastasis from the lung (Fig. 2), and a fragment analysis detected an *EGFR* exon 19 deletion (4).

Liver metastasis was detected in December 2011, and the patient was administered chemotherapy with pemetrexed, gefitinib, gemcitabine, and vinorelbine, but these regimens all proved to be ineffective. Notably, the metastases in the right lower lobe of lung and liver progressed rapidly in comparison to other metastases (Fig. 3). In January 2013, she was admitted to the hospital due to bacterial pneumonia and

Figure 2. Microscopic findings of the resected right adrenal metastasis. (A) Hematoxylin and Eosin staining of the adrenal specimen showed that the tumor was poorly to moderately differentiated adenocarcinoma. (B) Immunohistochemical staining revealed that the specimen was positive for thyroid transcription factor-1, suggesting that these were metastases from the primary lung adenocarcinoma.

Figure 3. Computed tomography (CT) scans obtained in 2013. The largest mass in the right lower lobe of the lung (A) and the liver metastases (B) grew more rapidly than the other metastases, which was consistent with the emergence of a tumor with higher-grade morphology.

Table 1. The Patient's History of Anticancer Treatments.

Date	Treatment
September 1999	Right upper lobe lobectomy followed by adjuvant chemotherapy
March 2002	Carboplatin-paclitaxel chemotherapy
November 2002	Gefitinib
April 2004	Gamma knife radiosurgery for brain metastases (repeated seven times) under continuation of gefitinib
January 2010	Docetaxel added on gefitinib
July 2010	Erlotinib
January 2011	Resection of right adrenal metastasis under continuation of erlotinib
January 2012	Pemetrexed
May 2012	Gefitinib re-challenge
August 2012	Gemcitabine
December 2012	Vinorelbine

Figure 4. Autopsy specimen of the largest mass in the right lower lobe of the lung. (A) Hematoxylin and Eosin (H&E) staining of the largest mass in the right lower lobe of the lung showed a transitional zone of well-to-moderately differentiated adenocarcinoma and neuroendocrine morphology. (B) H&E staining of the neuroendocrine tumor portion revealed that the tumor grew in sheets and rosette-like structures and exhibited necrosis. The tumor cells were large and had abundant cytoplasm and prominent nucleoli. The neuroendocrine tumor portion was positive for neural cell adhesion molecule (C) and synaptophysin (D), supporting a diagnosis of large-cell neuroendocrine carcinoma (LCNEC). Both the LCNEC (E) and adenocarcinoma portions (F) of the lesion expressed an *EGFR* mutation with an exon 19 deletion.

eventually experienced fatal cardiac arrest in April of that year. The patient's history of anticancer treatments is shown in Table 1.

Autopsy findings revealed that the largest mass in the right lower lobe of the lung had directly invaded the epicardium and the liver. Using microscopy, phenotypic heterogeneity was observed among the different metastatic disease sites. Adenocarcinoma was detected in the intrapulmonary metastases, neuroendocrine tumors had disseminated through the peritoneum, and metastases were present in the liver and the epicardium. Surprisingly, these two morphologies and the transitional zone were both found in the largest lung

mass (Fig. 4A). The neuroendocrine tumor grew in sheets and rosette-like structures and exhibited necrosis. The tumor cells were large and had abundant cytoplasm and prominent nucleoli (Fig. 4B). An immunohistochemical analysis revealed that the neuroendocrine component was positive for neural cell adhesion molecule (NCAM), synaptophysin, and Ki-67 (Fig. 4C and D). In contrast, the adenocarcinoma portion was negative for NCAM and synaptophysin. Although LCNEC is often difficult to distinguish from SCLC, this neuroendocrine component was consistent with LCNEC. Notably, the *EGFR* mutation from the antecedent adenocarcinoma was retained in both components (Fig. 4E and F).

Table 2. Histological Diagnosis and *EGFR* mutation Status.

Specimen	Organ	Histological diagnosis	<i>EGFR</i> mutation
Surgery in 1999	Lung (left lower lobe; primary tumor)	Adenocarcinoma	exon 19 del., T790M (-)*
	Mediastinum lymph node	Adenocarcinoma	N/E
Surgery in 2011	Right adrenal gland	Adenocarcinoma	exon 19 del., T790M (-)*
Autopsy in 2013	Lung (multiple intrapulmonary metastases)	Adenocarcinoma	exon 19 del.
	Mediastinum lymph nodes	Adenocarcinoma	exon 19 del., T790M (-)*
	Pleural dissemination (left)	Adenocarcinoma	exon 19 del.
	Lung (right lower lobe; metastatic tumor)	Combined LCNEC and adenocarcinoma	exon 19 del. (both components)
	Pleural dissemination (right)	LCNEC	exon 19 del.
	Pericardium (invasive lesion)	LCNEC	exon 19 del.
	Liver (right lobe)	LCNEC	exon 19 del.
	Peritoneum dissemination	LCNEC	exon 19 del., T790M (-)*
	Para-aortic lymph nodes	LCNEC	exon 19 del.

EGFR: epidermal growth factor receptor, exon 19 del.: exon 19 deletion, LCNEC: large-cell neuroendocrine carcinoma, N/E: not examined

**EGFR* T790M mutation was examined using the Scorpion amplification refractory mutation system method.

The histological diagnosis and *EGFR* mutation status are summarized in Table 2.

Discussion

We herein report a case of lung adenocarcinoma that transformed to LCNEC and became resistant to EGFR-TKIs. LCNEC of the lung is a subtype of large-cell carcinoma and generally has a poor prognosis compared to other NSCLCs (5). Although a case of combined LCNEC and adenocarcinoma with an *EGFR* mutation has been reported (6), it is unlikely that the present patient originally had a combined LCNEC because a transitional zone between the two components was detected in the autopsy findings, despite no LCNEC portion existing in the lobectomy specimen. Furthermore, it is unlikely that a patient with recurrent LCNEC would have survived without optimal treatment for 10 years.

The development of EGFR-TKI resistance is known to be concordant with a phenotypic switch to SCLC from NSCLC (7-9). Because LCNEC is very similar to SCLC (10), we believe that histological transformation to LCNEC can be a mechanism of acquired EGFR-TKI resistance. Recently, Kogo et al. reported a case of *EGFR* mutant adenocarcinoma that transformed to LCNEC, which was diagnosed by bronchoscopic tumor resection (11). In their case, the patient had received long-term treatment for lung adenocarcinoma with many chemotherapy regimens including EGFR-TKIs, as performed in the present case. A single tumor that expresses an *EGFR* mutation in cells of many different morphologies may reflect the existence of cancer stem cells. In our case, we suspect that either minor clones with stem cell-like properties were selected for when the major clones were eliminated by EGFR-TKI treatment or that the EGFR-TKI treatment induced stem cell-like properties within cells (12).

Besides histological transformation, several mechanisms of acquired resistance have been reported, such as secondary

EGFR mutations and the activation of collateral *EGFR* signaling pathways (3). With respect to secondary mutations, *EGFR* T790M mutation is well-known and develops in 50% of lung adenocarcinomas that acquire resistance to EGFR-TKIs. Recently, inter-tumor heterogeneity was reported as a mechanism of acquired resistance to EGFR-TKIs. Furugen et al. reported a case in which the patient developed EGFR-TKI resistance via SCLC transformation and an *EGFR* T790M mutation in separate metastatic organs (13). In addition, several reports have suggested a reciprocal relationship between a T790M mutation and histologic transformation (14, 15). Although an *EGFR* T790M mutation was not detected in our case, it is possible that another resistance mechanism was present.

This is a rare case report of transformed LCNEC after EGFR-TKI exposure, although histological transformation in this setting is not uncommon. Sequist et al. reported that 14% of patients who underwent tumor biopsies experienced a fundamental histological transformation into SCLC after acquiring EGFR-TKI resistance (3). In the present case, we emphasize the value of repeatedly assessing cancers throughout the course of the disease and the importance of extending our understanding of drug resistance, which in turn would allow for the determination of the best treatment option for each patient. However, our case involved multiple organ metastases, making it difficult to select an appropriate biopsy site. Yanagisawa et al. reported a case of a patient with lung adenocarcinoma that expressed an *EGFR* mutation who had a lung mass that gradually enlarged despite treatment with erlotinib, although the other metastatic lesions remained the same size. The patient underwent a second lung biopsy via video-assisted thoracoscopic surgery and was ultimately diagnosed with LCNEC, which might have resulted from transformation of adenocarcinoma, as with our case (16). When EGFR-TKI resistance is detected, a second histopathological examination should be considered, especially with a rapidly progressing or highly invasive lesion.

Although cases of small cell transformation have been

previously reported to be responsive to regimens used to treat SCLC (3, 8), there are few reports about the treatment of LCNEC transformed from adenocarcinoma. However, LCNEC responds well to SCLC regimens, compared with traditional NSCLC treatments (17). Additionally, Kogo et al. reported a case of LCNEC transformation in which the patient was treated with SCLC regimens and obtained a partial response (11). We believe the present LCNEC could have been treated with effective regimens, such as amrubicin or irinotecan, if the patient had been diagnosed with LCNEC earlier.

In summary, we herein reported an autopsy case of LCNEC that transformed from adenocarcinoma harboring an *EGFR* mutation. The findings from this case showed the clinical importance of pathological assessment at the time of progression in patients with tumors that express an *EGFR* mutation. Further investigation is necessary to clarify the *EGFR*-TKI resistance mechanisms involved in transformation to LCNEC.

The authors state that they have no Conflict of Interest (COI).

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