Rituximab is a highly effective agent that is used in the treatment of B-cell lymphoma. Rituximab-induced acute thrombocytopenia (RIAT) is a rare side effect that has previously been reported in a small number of patients with malignant lymphoma; its mechanism is still unknown. We herein report the case of a 74-year-old man who was diagnosed with follicular lymphoma and who developed severe acute thrombocytopenia the day after the administration of rituximab. Coagulation abnormality, which mimicked disseminated intravascular coagulation, also appeared. When physicians use rituximab to treat high-risk patients, the platelet count should be closely monitored to avoid possible adverse events.

**Key words:** rituximab, acute thrombocytopenia, follicular lymphoma

**Case Report**

A 74-year-old man with inguinal lymphadenopathy was referred to our hospital. His medical history revealed chronic atrial fibrillation, which was treated with warfarin. A physical examination showed bilateral cervical, subclavian, axillary, and left inguinal lymphadenopathy. Laboratory tests revealed pancytopenia: white blood cell (WBC) count, 2,900/μl, with 2% atypical lymphocytes; hemoglobin (Hb), 12.0 g/dl; and platelet (PLT) count, 61,000/μl. Computed tomography revealed systemic lymphadenopathy and massive splenomegaly. The histological examination of a left axillary lymph node biopsy specimen revealed grade 2 follicular lymphoma. Immunohistochemistry revealed that the lymphoma cells were positive for CD20 and Bcl-2. A chromosome analysis revealed 46, XY, add(1)(p36,3), t(14;18)(q32;q21.3) in the tumorous lymph nodes. A polymerase chain reaction detected immunoglobulin heavy chain gene rearrangement. Bone marrow biopsy revealed the infiltration of the bone marrow by lymphoma cells (abnormal lymphoid cells that were positive for CD20 and Bcl-2).
The patient was scheduled to receive R-CVP (rituximab, cyclophosphamide, vincristine, prednisolone) chemotherapy. Doxorubicin was eliminated due to the patient’s low cardiac function. The patient was pretreated with prednisolone to avoid the risk of tumor lysis syndrome; cyclophosphamide and vincristine were then administered. On day 14 of CVP therapy, he underwent rituximab infusion. He developed fever and shivering during the infusion. The infusion was stopped for a few hours and then restarted at a slow rate without any complications. On day 2 of the 2nd cycle of R-CVP, he underwent a 3-hour infusion of rituximab without any reactions to the infusion. The next day, a blood test showed a drop in the PLT count from 75,000/μl to 36,000/μl with a slight elevation in the PT-INR from 2.31 to 2.62. He did not experience any major adverse events until the end of the 2nd cycle of R-CVP therapy.

On day 2 of the 3rd cycle of the R-CVP therapy, he received a 3-hour rituximab infusion. A blood test before the administration of rituximab revealed the following findings: WBC count, 3,400/μl; Hb, 10.5 g/dl; PLT count, 63,000/μl; d-dimer, 6.3 μg/ml; and PT-INR, 1.90. The next day, a blood test showed a drastic drop in the PLT count to 14,000/μl, the elevation of the d-dimer level to 33.3 μg/ml and the elevation of the PT-INR to 2.97. Transfusions were performed twice a day in a row with 20 units of platelets, 8 units of fresh frozen plasma, and 19,200 units of recombinant thrombomodulin alpha. The post-transfusion PLT count was 63,000/μl. The serum levels of creatinine, uric acid, lactate dehydrogenase and electrolytes were not affected by the administration of rituximab, which ruled out an association with tumor lysis syndrome. Contrast-enhanced computed tomography detected no intravenous thrombosis and the patient’s enlarged spleen was found to have shrunk from the 1st cycle of R-CVP therapy. His platelet count spontaneously recovered to 110,000/μl within one week. The fourth administration of rituximab also caused acute thrombocytopenia. The following day, the PLT count decreased from 61,000/μl to 9,000/μl, the d-dimer level increased from 4.6 μg/ml to 64.2 μg/ml, and the PT-INR increased from 2.56 to 3.10 without a decrease in either the WBC count or the Hb level. The thrombin-antithrombin complex (TAT) level was 4.9 ng/ml and the plasmin-α2 plasmin inhibitor complex (PIC) level was 6.1 μg/ml, which indicated coagulopathy. The extent of thrombocytopenia in the 4th cycle of rituximab therapy was much greater than that observed during the 3rd cycle. The serum biochemistry and electrolytes values showed almost no change after the administration of rituximab. His platelet counts spontaneously recovered again within one week. Figure shows the platelet count in the 4th course of rituximab infusion. Platelet associated immunoglobulin G (PA-IgG) was not detected by an enzyme-linked immunosorbent assay (ELISA) of blood samples obtained on day 1 after the administration of rituximab. Furthermore, human anti-chimeric antibodies (HACA) to the murine fragment of rituximab were not detected by an ELISA (HACA to rituximab was measured by Falco Biosystems, Kyoto, Japan). Rituximab was omitted from the subsequent cycles of treatment to avoid the risk of life-threatening thrombocytopenia.

**Discussion**

Rituximab is commonly used in the treatment of B-cell lymphoma and its use is generally well-tolerated. RITAT has been reported as a rare side effect and its mechanism remains unclear. We searched the PubMed database using the terms “rituximab” and “acute thrombocytopenia”. The search yielded a total of 21 cases, the diagnoses of the cases included mantle cell lymphoma (n=13) (4, 6, 8-12, 14, 15), hairy cell leukemia (n=3) (6, 7, 13), prolymphocytic leukemia (n=1) (5), lymphoplasmacytic lymphoma (n=1) (3), follicular lymphoma (n=1) (16), pre-B acute lymphoblastic leukemia (n=1) (17), and autoimmune hemolytic anemia (n=1) (18). Table shows the reports on RITAT. As described above, in most cases, RITAT occurred in patients with mantle cell lymphoma. The incidence of RITAT in this lymphoma subtype differs from the incidence of the histological lymphoma subtype (the frequency of mantle cell lymphoma is low in Japan and throughout the rest of the world). This means that development of RITAT is strongly associated with the lymphoma subtype. We herein report the 2nd case of a patient with follicular lymphoma who developed severe acute thrombocytopenia as a result of the administration of rituximab. It is important that RITAT also occurs in follicular lymphoma, which is a more frequent subtype of lymphoma. Giezen et al. reported on the incidence of rituximab-induced thrombocytopenia (platelet count, <100,000/μl within 30 days after the administration of rituximab) in a cohort study (19). Among 90 rituximab-treated patients, 27 patients developed thrombocytopenia (30%); the diagnoses of these patients included Burkitt lymphoma (n=3), diffuse large B-cell lymphoma (n=6), NHL (n=11), CLL (n=2) and other (n=5).

Our case shared some common clinical features with the cases of previous reports, specifically, massive splenomegaly, and bone marrow infiltration (Table). On the
other hand, our case was distinguished from previous cases in that the patient developed RIAT from the 3rd cycle of rituximab onwards and the lymphoma subtype was follicular lymphoma. Although the tumor burden and the degree of splenomegaly were still severe when RIAT first developed, the tumor volume was lower and the size of the spleen was smaller in comparison to the 1st chemotherapy cycle. This means that the tumor burden itself may not independently explain the pathogenesis of RIAT. Furthermore, our case had coagulopathy. Some of the other RIAT cases showed coagulopathy while others did not (Table). Although our case developed an infusion reaction on the first administration of rituximab, he did not develop an infusion reaction during the subsequent courses of rituximab therapy. However, in many of the RIAT cases, the infusion reaction occurred simultaneously; thus, this may be a risk factor.

Our hypothesized mechanisms of RIAT are as follows. First, in some RIAT cases, including ours, the patient simultaneously developed coagulopathy (7, 13, 16, 17). Thus, infused rituximab interacts with lymphoma cells, which might activate immune components and cause consumption coagulopathy, which expends platelets (17). Second, platelets have an Fc receptor (20); thus, rituximab antibodies, which bind to lymphoma cells, also bind the platelets through their Fc receptors, leading to platelet degradation. Previous reports have suggested that the presence of soluble CD20 antigen in the circulation may cause an antigen-antibody reaction and immune-mediated cell lysis (21).

There have been several reports on the re-infusion of rituximab in patients with RIAT (Table). In some cases (including our own), RIAT recurred (4, 8, 11, 12, 17). In other cases, there was no recurrence of RIAT after rituximab re-infusion (3, 14, 15). Furthermore, in several cases (including our own), RIAT occurred first after several rituximab infusions (8, 10, 14). At present, the reason for these differences is unknown.

The incidence of RIAT may be underestimated because platelet counts are not always monitored in the first days after rituximab therapy, especially when patients are treated in the outpatient setting. Moreover, it is possible that a patient’s thrombocytopenia could be so mild that RIAT goes unnoticed. In our case, mild thrombocytopenia occurred after the second infusion of rituximab and might have also been RIAT.

We herein reported the case of a patient with follicular lymphoma who developed RIAT in the 3rd and 4th cycles of rituximab chemotherapy. Routine blood count monitoring should be considered for a few days after the administration of rituximab, especially for high-risk lymphoma patients.

The authors state that they have no Conflict of Interest (COI).

References

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