Effects of Soaking and High-Pressure Treatment on the Qualities of Cooked Rice

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Abstract: In this study, changes in rice qualities from High-Pressure (HP) treatment were investigated. Milled rice grains were presoaked in water at 25 and 55°C for 30 min, subjected to HP treatment at 400 MPa for 10 min, and soaked in water overnight. The effects of presoaking temperature and HP treatment on the physical and chemical properties of rice were evaluated. The viscosities of milled rice grains increased with the soaking process. The total sugar content increased and change in the internal structure of the rice grains occurred after HP treatment. The structural change seemed to promote the water penetration and brought about a higher degree of swelling in rice granules, which might result in a higher degree of gelatinization and higher digestibility of HP-treated cooked rice. HP treatment also brought about denaturation of water soluble proteins and an increase in some free amino acids. HP processing was thus proved to be one of the advantageous processing methods for cooked rice with better palatability.

Key words: high pressure, rice, gelatinization, NMR imaging, digestibility

Recently high pressure (HP) technology has been recognized to be very useful in the food industry.1 Numerous studies were carried out about effects of HP treatment on microbial physiology and control of enzymatic reaction related to sterilization and food processing.1,2 There were, however, only a few reports on HP-treated rice in spite of its importance as an energy and protein source. The application of HP treatment to indica rice grains changed their cooking properties as well as enzymatic activities, and protein solubility using low-amylose, medium-amylose and high-amylose rice. The internal structural change of grains by HP treatment was discussed in relation to degree of gelatinization and digestibility of the cooked rice.

MATERIALS AND METHODS

Rice samples. Mature seeds of non-glutinous rice (Oryza sativa L.) cultivars, Milky Queen (Japonica, low-amylose rice), Koshihikari (Japonica, high-quality rice) and Hamasari (Indica, high-amylose rice), were used. Milky Queen was grown in Ibaraki Prefecture, Japan in 2001, Koshihikari in Niigata Prefecture, Japan in 2002, and Hamasari in Saitama Prefecture, Japan in 2002. The samples were stored at low temperature (1–3°C) before use. Brown rice grains were milled to the milling yield of
90% with a TM-05 Grain Testing Mill (SATAKE, Hiroshima, Japan).

**High-Pressure (HP) treatment.** After removal of broken grains, 20 g of whole grains were put into a polyethylene film bag together with 30 mL distilled water. The sample was vacuum-packaged and kept at 25 or 55°C for 30 min. The sample was then treated at 400 MPa for 10 min in a HP machine (Ishikawajima-Harima Heavy Industries Co., Ltd., Tokyo, Japan). On the other hand, a “non-HP sample” was kept for at 25 or 55°C for 15 min and stored in a refrigerator at 1–3°C overnight.

**Preparation of cooked rice sample.** The soaked rice grains with water were put into an aluminum cup and kept at room temperature for 30 min. The sample was cooked in an electric rice cooker (SR-ULH18, National Co., Ltd., Osaka, Japan) and kept warm in the cooker for 15 min. Ten grains of cooked rice were frozen rapidly with liquid nitrogen and then lyophilized.

**Preparation of rice flour sample.** The soaked rice grains were lyophilized and ground into flour with a Cyclone sample mill (UDY Corp., Ft. Collins, Colorado, USA). Dried cooked rice grains and intact rice grains were ground into flour, too.

**Chemical analysis.** The amylase content of the samples was measured by the Juliano method. The total sugar of cooked rice samples was measured by the phenol-sulphuric acid method. The reducing sugar of cooked rice was measured by the method of Somogyi-Nelson using glucose as a standard. Free amino acid contents were measured by the method of Saikusa et al. Free amino acids were extracted from 0.5 g of flour from intact rice grains or treated cooked rice with 5.0 mL of sulfosalicylic acid (2%) and then centrifuged. The supernatant was filtered through a 0.45 μm filter (GL chromato-disk 25A, GL Science Co., Ltd. Japan). The prepared sample solution (10 μL) was analyzed by an amino acid analyzer (Hitachi 8500, Tokyo, Japan). Nitrogen content of the samples was measured by the combustion method using a LECO System (LECO FP-528, LECO Corporation, Michigan, USA) using EDTA (LECO Corporation) as a standard.

**Enzyme activity assay.** Protease activity was measured by the method of Palmiano and Juliano with minor modifications. The rice grains were separated from soaking water by centrifugation after the HP treatment. The soaked rice grains were homogenized with a homogenizer for 1 min in a HP machine (Ishikawajima-Harima Heavy Industries Co., Ltd., Tokyo, Japan) and kept warm in the cooker for 15 min. Ten grains of cooked rice were frozen rapidly with liquid nitrogen and then lyophilized. One unit of protease activity was expressed as the amount of enzyme that increase absorbance of the solution as 0.1 under the conditions of assay. α-Amylase and β-amylase activities were determined using the kits of Megazyme International Ireland, Ltd. (Wicklow, Ireland). α-Glucosidase activity was measured according to the method reported by Imai et al. and Iwata et al.

**Pasting properties.** Pasting properties of soaked and intact rice flours were measured using a RVA (Rapid-Visco-Analayzer, New-port Science Ltd., Sydney, Australia) by the method of Toyoshima et al.

**Starch digestibility of cooked rice.** Starch digestibility was measured by the method of Niihara and Miyoshi with minor modifications. The cooked rice was cooled down to 40°C at room temperature. Ten grams of cooked rice were homogenized with a homogenizer for 1 min in 90 mL water, and the pH was adjusted to 2.0. The sample was incubated at 37°C for 30 min with the addition of 2.5 mL of pepsin (Sigma-Aldrich Co., St. Louis, USA) solution (100 mg/mL). The pH of the solution was then adjusted to 7.1 at 37°C, and 0.1 mL aliquots were analysed. The residue was then incubated at 37°C for 35 min with the addition of 2.5 mL of pancreatin (360 mg/12 mL), and 1.0 mL aliquots were analysed after incubation for 5, 15, 30 and 60 min.

**Degree of starch gelatinization of cooked rice.** Degree of starch gelatinization was measured by the method of Kainuma et al. using dried cooked rice flour as samples.

**Nuclear Magnetic Resonance (NMR) micro imaging.** NMR micro imaging was performed by the method of Horigane et al. Soaked raw rice grains and cooked rice grains were used as samples.

**Preparation of gel electrophoresis sample.** For water soluble protein analysis, the HP treated sample was homogenized for 1 min and centrifuged at 5000 rpm at 4°C for 20 min. The supernatant was filtered and freeze-dried. The sample was dissolved in water, and centrifuged. The supernatant was lyophilized again, and then dissolved in 0.5 mL of water. The solutions (0.05 mL) were mixed with Lysis buffer (0.1 mL) and subjected to 2D-PAGE, as described by Hirano.

**Electrophoresis.** Isoelectric focusing (IEF) gels were prepared as described by O’Farrell. Water soluble protein samples were used, 0.05 mL per glass tube. The sodium dodecyl sulphate (SDS)-PAGE was performed by the method of Laemmli with 17% separation gels and 5% stacking gels. A molecular weight marker kit (Perfect Protein Markers 10–225 kDa, Novagon Co., Ltd.) was used as standard proteins. After electrophoresis, a part of the gels were stained with Coomassie Brilliant Blue R-250 (CBB).

### RESULTS AND DISCUSSION

In this study, we used low-amylase (Milky Queen), medium-amylase (Koshihikari) and high-amylase (Hamasari) rices as samples. Table 1 summarizes the amylase content and pasting property of the rices. The maximum viscosity, minimum viscosity and final viscosity increased significantly with soaking, and differed (p<0.05) from
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Table 1. Amylose content, particle size and pasting properties of intact and processed rice flours.

<table>
<thead>
<tr>
<th></th>
<th>Presoaked temp. (C)</th>
<th>Amylose (%)</th>
<th>Particle size</th>
<th>Pasting properties</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Average (μm)</td>
<td>Under 200 μm (%)</td>
<td>Max. Visc. (RVU)</td>
</tr>
<tr>
<td>Intact milled rice</td>
<td>—</td>
<td>8.3 ± 0.07</td>
<td>175.5</td>
<td>327.0</td>
</tr>
<tr>
<td>Milky Queen</td>
<td>25</td>
<td>8.4 ± 0.07</td>
<td>116.3</td>
<td>431.0</td>
</tr>
<tr>
<td>non-HP</td>
<td>55</td>
<td>8.4 ± 0.14</td>
<td>119.6</td>
<td>450.8</td>
</tr>
<tr>
<td>HP</td>
<td>25</td>
<td>9.0 ± 0.21</td>
<td>146.7</td>
<td>405.9</td>
</tr>
<tr>
<td>HP</td>
<td>55</td>
<td>8.2 ± 0.00</td>
<td>169.9</td>
<td>397.3</td>
</tr>
<tr>
<td>Koshihikari</td>
<td>non-HP</td>
<td>44.0 ± 0.12</td>
<td>120.0</td>
<td>407.3</td>
</tr>
<tr>
<td>HP</td>
<td>25</td>
<td>17.8 ± 0.07</td>
<td>131.9</td>
<td>384.3</td>
</tr>
<tr>
<td>HP</td>
<td>55</td>
<td>17.6 ± 0.28</td>
<td>145.1</td>
<td>379.3</td>
</tr>
<tr>
<td>Hamasari</td>
<td>non-HP</td>
<td>44.0 ± 0.12</td>
<td>120.0</td>
<td>407.3</td>
</tr>
<tr>
<td>HP</td>
<td>25</td>
<td>28.2 ± 0.21</td>
<td>119.0</td>
<td>284.0</td>
</tr>
<tr>
<td>HP</td>
<td>55</td>
<td>28.4 ± 0.36</td>
<td>137.4</td>
<td>234.4</td>
</tr>
</tbody>
</table>

Mean value ±SD (n = 3). Different letters in the same column indicate significant difference (p < 0.05).

Table 1 summarizes the particle size of the flour sample. The difference between pre-soaked samples and the intact ones was marked in each cultivar, and was significantly different (p < 0.05) from those of the intact rice, except for in HP-Milky Queen at 55°C. In Milky Queen and Koshihikari, the particle size diameter 200 μm in intact milled rice was lower than in pre-soaked samples except for in HP-Milky Queen at 55°C. High amylose rice particles were smaller than for other cultivars,27 and same result in our study. It was reported that maximum viscosity of the large size particle was lower than for the small size one in amylogram.26 Maximum viscosity in RVA (Table 1) and particle size of flour changed with soaking. However, no correlation (r = −0.074) between maximum viscosity in RVA and particle size of flour.

Total sugar increased with the soaking at 55°C and HP treatment, and significantly differed (p < 0.05) from those of the non-HP at 25°C (Fig. 1A). Milky Queen had the largest amount of total sugar and decreased in the order of Hamasari and Koshihikari. Reducing sugar increased with HP treatment at 25°C, the difference of which was small with that of the non-HP treated sample (Fig. 1B). Hamasari had the largest amount of reducing sugar, followed by Milky Queen and Koshihikari. The result suggested that total and reducing sugar contents were not related to amylose content, but to HP treatment (p < 0.01). The results suggested that change in the enzyme activity in the rice grains was caused by the HP-treatment.

α-Amylase activity was highest in the intact milled rice for all cultivars, but decreased with soaking and HP-treatment (Fig. 2A). On the other hand, β-amylase activity was detected after HP treatment and soaking in Milky
Queen and Koshihikari (Fig. 2B) and showed a little high activity even after soaking or HP treatment at 55°C. This may be due to one of the α-amylases having a characteristic of high temperature resistance.

Amylase in high amylose rice (Hamasari) showed high activity as well as that of β-amylase. As shown in Fig. 3, α-glucosidase activity of Milky Queen and Koshihikari was higher than that of Hamasari. β-Glucosidase activity of intact milled rice was highest for each cultivar. The activity decreased a little with soaking but maintained almost the same value even after HP treatment. It was reported that β-glucosidase activity was detected even during the cooking process, and it had activity at a high optimum temperature. The tolerance of β-glucosidase activity might lead to an increase in the low-molecular weight sugars, such as glucose and maltose, during presoaking and HP treatment. Masui reported that the β-glucosidase activity increased with HP treatment when milled rice flour was used. However, β-glucosidase activity after HP treatment was smaller than that of intact milled rice in our experiment.

Figure 4 shows the results of the starch digestibility test of cooked rice. HP treatment increased starch digestibility for all cultivars. In Milky Queen and Koshihikari, reducing sugar content after 5 min digestion reached about 80–90% for 60 min digestion. With HP treatment, more reducing sugars were generated by pancreatin digestion. In Hamasari, reducing sugar content after 5 min digestion was about 70–80% for 60 min digestion. The lowest di-

![Fig. 2. Amylase activity in the rice grains and soaking solution.](image)

![Fig. 3. α-Glucosidase activity in the rice grains and soaking solution.](image)

![Fig. 4. Starch digestibility of cooked rice.](image)
Table 2. Degree of gelatinization of starch and protein contents of samples.

<table>
<thead>
<tr>
<th>Presoaked temp. (°C)</th>
<th>Degree of gelatinization of starch (%)</th>
<th>Protein content</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Total protein</td>
<td>Insoluble protein</td>
</tr>
<tr>
<td>Intact milled rice</td>
<td>—</td>
<td>7.59±0.08</td>
<td>—</td>
</tr>
<tr>
<td>Milky Queen</td>
<td></td>
<td>7.49±0.07</td>
<td>5.89±0.04</td>
</tr>
<tr>
<td>non-HP</td>
<td>25</td>
<td>85.6</td>
<td>7.57±0.04</td>
</tr>
<tr>
<td>HP</td>
<td>25</td>
<td>83.6</td>
<td>7.52±0.13</td>
</tr>
<tr>
<td>HP</td>
<td>55</td>
<td>90.7</td>
<td>7.53±0.02</td>
</tr>
<tr>
<td>Koshihikari</td>
<td></td>
<td>5.94±0.02</td>
<td>—</td>
</tr>
<tr>
<td>non-HP</td>
<td>25</td>
<td>85.5</td>
<td>5.88±0.04</td>
</tr>
<tr>
<td>HP</td>
<td>25</td>
<td>84.6</td>
<td>5.97±0.03</td>
</tr>
<tr>
<td>HP</td>
<td>55</td>
<td>91.4</td>
<td>5.82±0.00</td>
</tr>
<tr>
<td>Hamasari</td>
<td></td>
<td>7.92±0.02</td>
<td>—</td>
</tr>
<tr>
<td>non-HP</td>
<td>25</td>
<td>73.2</td>
<td>7.95±0.26</td>
</tr>
<tr>
<td>HP</td>
<td>25</td>
<td>74.7</td>
<td>7.83±0.04</td>
</tr>
<tr>
<td>HP</td>
<td>55</td>
<td>81.5</td>
<td>7.83±0.13</td>
</tr>
<tr>
<td></td>
<td></td>
<td>8.01±0.10</td>
<td>6.42±0.16</td>
</tr>
</tbody>
</table>

Mean value±SD (n = 3). Ratio: Insoluble protein/Total protein.

Fig. 5. NMR micro images of raw and cooked rice grains of cv. Koshihikari.

a, raw rice grains (non-HP); b, raw rice grains (HP); c, cooked rice grains (non-HP); d, cooked rice grains (HP). Arrows refer to the following structures: (cr), crack; (ho), hollow. The bar indicates 1 mm.
gestibility for high-amylose rice (Hamasari) was observed. For HP-treated samples, reducing sugar content after 60 min digestion was 1.12–1.27 times higher than that of non-HP sample. The result indicated that HP-treated rice was more easily digested. It was reported that water penetrates to the deep part of the rice grain after HP treatment. This suggested that HP treatment injures the microstructure of the rice grains to some extent, which allows water to penetrate to the inner part and makes starch easier to digest.

The degree of gelatinization of starch in cooked rice is listed in Table 2. The degree of starch gelatinization of just-cooked rice is usually 94–96%. Though our cooked rice was lyophilized, Milky Queen and Koshihikari showed a high degree of gelatinization. HP treatment resulted in a higher degree of gelatinization than for non-HP samples, probably due to the high penetration of water.
Table 3. Free amino acid contents of high pressure treated raw rice samples (mg/100 g flour sample, dry basis).

<table>
<thead>
<tr>
<th>Components</th>
<th>Milky Queen</th>
<th>Koshihikari</th>
<th>Hamasari</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>25°C</td>
<td>55°C</td>
<td>25°C</td>
</tr>
<tr>
<td></td>
<td>non-HP</td>
<td>HP</td>
<td>non-HP</td>
</tr>
<tr>
<td>Asp</td>
<td>9.37</td>
<td>9.84</td>
<td>10.00</td>
</tr>
<tr>
<td>Thr</td>
<td>0.60</td>
<td>0.79</td>
<td>0.69</td>
</tr>
<tr>
<td>Ser</td>
<td>2.85</td>
<td>3.20</td>
<td>3.09</td>
</tr>
<tr>
<td>Glu</td>
<td>7.56</td>
<td>7.31</td>
<td>7.14</td>
</tr>
<tr>
<td>Glu NH₃</td>
<td>10.74</td>
<td>11.93</td>
<td>11.93</td>
</tr>
<tr>
<td>Gly</td>
<td>1.03</td>
<td>1.77</td>
<td>1.24</td>
</tr>
<tr>
<td>Ala</td>
<td>0.06</td>
<td>0.06</td>
<td>0.06</td>
</tr>
<tr>
<td>GABA</td>
<td>7.82</td>
<td>9.02</td>
<td>9.25</td>
</tr>
<tr>
<td>Total amino acids</td>
<td>40.04</td>
<td>43.92</td>
<td>43.40</td>
</tr>
</tbody>
</table>

Different letters in the same column indicate significant difference (p < 0.05), (n = 3).

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into the rice grains brought about by HP treatment.

The effect of HP treatment on the structure of the inner part of the rice grains was investigated by NMR micro imaging. Figure 5 shows the MR slice images at the central part of raw and cooked grains of Koshihikari. Soaked raw rice grains had cracks filled with penetrated water, but no difference was observed between non-HP (Fig. 5a) and HP-treated samples (Fig. 5b). As for cooked grains, internal hollows were found in non-HP grains (Fig. 5c) but not in HP-treated samples (Fig. 5d). It was reported that internal hollows were observed more in cooked non-waxy rice grains than in waxy rice, which contained only small rounder hollows. Cultivars with a higher final hollow ratio in the cooked grains generally showed a higher peak in RVA, which was reflected in rigidity to starch granules. In this study, HP-treated rice had a lower peak in RVA than non-HP rice (Table 1). Yamazaki et al. demonstrated that water penetrated to the deep part of the grain after HP-treatment, which suggested that the HP treatment allows water to penetrate to the inner part of the rice grain and makes the starch granules more swollen and fragile than the normal soaking process under ordinary pressure. This suggestion is supported by the results that HP-treated starch granules were gelatinized at a lower peak viscosity than those of normally soaked samples (Table 1), and HP-treated samples showed a higher degree of gelatinization (Table 2). The starch granules in the inner part of the HP-treated grains may be more expanded than those in the normally soaked grains and invade most of the hollows at the end of cooking like in waxy rice.

Figure 6 shows the change in protease activities with the soaking and/or HP treatment. The intact milled rice grains showed the highest enzyme activity, but the activity decreased with the soaking. When the soaking temperature was high, the activity decreased. HP treatment also made the protease activity decrease. It has been reported that protease was located in the outer layer of the rice grain. In our study, protease activity decreased with soaking at high temperature and/or HP treatment.

Table 2 shows the total protein and insoluble protein contents. Total protein content was 7.49–7.59% in Milky Queen, 5.82–5.97% in Koshihikari and 7.83–8.01% in Hamasari, and no definite difference was observed between non-HP and HP. The ratio of insoluble proteins to total protein increased in the order of intact rice, non-HP rice and HP treated rice. Allergic proteins of rice were found in a globulin and albumin fraction. In our study, HP-treatment denatured soluble proteins and made them water insoluble.

Figure 7 shows 2D-PAGE patterns of water soluble protein. The protein spots corresponding to non-HP and HP were clearly observed in the stained gel. In HP, several protein spots disappeared and some other protein spots markedly decreased their staining intensity compared with non-HP. Rice proteins are classified by their solubility. The greater part of water soluble proteins are estimated to be albumin, which is located in aleurone cells. Thus, it was estimated that HP treatment would be effective to denature the water soluble protein, albumin, because it was located in the outer layer of the milled rice.

Table 3 summarizes the free amino acid contents. Although free amino acid content was higher in Hamasari compared to the other two cultivars, no definite difference was observed between non-HP and HP treated samples. In the case of Milky Queen and Koshihikari, some free amino acids increased with the soaking at 55°C or HP treatment. This result was similar to that reported by Kinefuchi et al. Hamasari had the highest amounts of free amino acids, followed by Milky Queen and Koshihikari. The result suggested that the amount of total and reducing sugar were not related to amylose content. From the results, it was shown that some amino acids increased in HP treatment, though protease activity decreased in presoaking and HP treatment. The preferential proteinolysis by HP treatment was reported. The rate of protein hydrolysis increased as the pressure increased. This suggested that protein was denatured by the HP treatment, and therefore proteolysis increased. From the results, it was concluded as follows: (1) The hydrolytic enzymes, optimum temperature of which were high, functioned during the presoaking. (2) Rice proteins were decomposed to increase the free amino acids during the HP treatment. And one after another, some proteases were damaged. (3) Some free amino acid such as serine, glycine and GABA increased with soaking temperature and/or HP treatment. This means that high temperature presoaking and HP treatment can be effective for the improvement of taste, nutrition and bio-functionality such as prevention of hypertension, which was reported for GABA of cooked rice.
REFERENCES

超高圧処理と浸漬が炊飯米の品質に及ぼす影響
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超高圧処理は、食品産業における新しい技術であり、
米飯にも応用されている。日本では通常、炊飯前に浸漬
を行う。浸漬もまた炊飯米の性状に影響を及ぼす要因
である。夜に洗米・浸漬を行い、翌朝に炊飯を開始する
ということもしばしば行われる。本報告では、25℃ある
いは55℃で30分間の予備浸漬を行い、その後400 MPa
で10分間の超高圧(HP)処理を行った後で一晩の浸漬を続
け、予備浸漬温度とHP処理の有無が米に与える物理的、
化学的影響を調べた。その結果、糊化特性では、浸漬操
作により最高粘度が増加し、食味が向上する可能性が示
唆された。また、HP処理により、糖含量の増加が認めら
れ、また炊飯米粒内部の構造の変化も認められた。また、
炊飯米の糊化度が増し、消化性が向上した。また、HP処
理区において、純水可溶性タンパク質の変性がみられた。 
呈味成分である遊離アミノ酸は、予備浸漬温度、HP処理
に伴って増加していた。以上のことから、超高圧処理は 
米加工に利用でき、新たな食味改善技術の一つとしての
可能性が期待できると考えられる。