Starch and Alcohol Fermentation of Viviparous Buds of Mangroves

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The viviparous buds of mangroves were found to contain starch in amounts up to 28% on a dry basis. The starch granules existed separately, being inserted between cellulosic fibers of the tissue. The starch granules were ellipsoidal or oval of ca. 10-8 x 5-3 μm, depending on the kind of mangroves.

The viviparous buds were examined for alcohol fermentation after steeping in hot water and crushing mechanically. Glucoamylase and pectic enzymes were added to the mash for saccharification of starch and maceration of the crushed tissue, respectively. The starch in the viviparous buds were fermented to alcohol, but a satisfactory fermentation efficiency was not obtained. From the facts that the tissue of viviparous buds is tough to treat and contains tannic substance, the processing for alcohol fermentation is discussed.

Mangroves, an embryonic plant constituting the major forest of the plant community in tidal swamp area of South East Asia, bear a great number of viviparous buds annually. The size of the buds varies from 25 to 60 cm in length and, from 15 to 100 g in weight depending on the kind of mangroves. Their tissues are generally tough and consist of cellulosic, hemicellulosic and pectic substances, and contain sodium and calcium salts as the major inorganic substance. They also contain starch and tannic substance. Application of this tannic substance has been established to some extent. However, most of the viviparous buds lose their lives in vein by being carried away with sea water or by falling down in so unfortunate way on the ground as not to be able to root for growth. It happens in some circumstances that the beach of mangrove flora is covered with dead viviparous buds. Therefore, it may be necessary to manage the viviparous buds of mangrove trees to keep them as a sound forest to be able to play the important role of protecting the sea shore from erosion and to maintain various living organisms in the surrounding sea water.

The present study was thus attempted to find a clue for development of utilization of viviparous buds of mangroves. As a preliminary report, this paper describes isolation and some properties of starch in mangrove viviparous buds and feasibility of application of the buds for alcohol fermentation.

MATERIALS AND METHODS

1. Viviparous buds of mangroves examined. Ceriops tagal, Rhizophora apicurata and Rhizophora mucronata were collected from marsh lands in Phuket, Southern Thailand in March, 1982. The viviparous buds of R. mucronata obtained in Iriomote Island, Okinawa Pref. of Japan in October were also examined.

2. Detection and quantitative determination of starch. The starch in tissue of viviparous bud was detected by staining the tissue section with iodine-potassium iodide solution. For
quantitative determination of starch, viviparous
buds were grated using a grater at kitchen.
The mash obtained was suspended in 50 or 100
weights of water and the suspension was boiled
for 30 min. The suspension was cooled to 50°C,
the pH was adjusted at 5.0, and a commercial
preparation of glucoamylase was added to the
suspension in a proportion of 2.0 mg per 1 g of
the buds on dry basis. After incubation for 16 hr
under few drops of toluene, the suspension was
centrifuged and the supernatant was subjected
to determination of reducing sugar as glucose
by the method of Shaffer-Somogyi. The control
run was the suspension incubated with the
enzyme previously inactivated by heating in
boiling water for 20 min, and the reducing sugar
as glucose estimated with this suspension was
subtracted from the reducing sugars observed
for the incubation mixture with active gluco-
amylase. The starch content was estimated by
multiplying the factor, 162/180, to the glucose
value thus obtained.

The reducing sugar produced by hydrolyzing
the grated viviparous buds with 100 volumes of
1.0 N hydrochloric acid at 100°C for 2 hr was
also estimated. The difference between the
value estimated as glucose by this method and
that of glucose determined by using glucoamy-
lase was tentatively expressed as hemicellulosic
polysaccharides.

3. Isolation of starch. The isolation of
starch from the viviparous buds was done as
follows: Five hundred grams each of the buds
were sliced and suspended in 5 weights of
water. The suspension was subjected to homog-
enization by a Waring blender at a high speed
for 1 min. The homogenate obtained was
transferred to a big glass cylinder and tap water
was allowed to flow through a glass tube to the
bottom to wash out light specific gravity mate-
rials leaving starch as precipitate.

4. Fermentability for alcohol of viviparous
buds. The buds were cut down into rough
pieces and kept in water heated at 70°C for
30 min to inactivate polyphenol oxidase. Then,
the bud pieces were taken out and cooled to
room temperature. An equal weight of water
was added to the pieces and homogenized in a
Waring blender at a high speed for 2 min.
Commercial preparations of pectic enzymes
(0.25 g), glucoamylase (0.25 g) and dry yeast
(2.5 g) were added to 500 g of the homogenate
and incubated in a modified Meissel bottle at
28°C. The degree of production of alcohol was
tentatively expressed by weight loss of the
incubation mixture due to evolution of carbon
dioxide.
5. Enzyme preparations used. Glucoamylase was a preparation from Rhizopus niveus. The activity was 6,000 units per g preparation (One unit of the enzyme produced 10 mg of glucose in 30 min at 40°C and at pH 5.0). A pectic enzyme preparation obtained from Aspergillus niger had the activity of 6,000 units as pectin depolymerase per g preparation (One unit of pectin depolymerase activity was defined as the enzyme amount that reduced the viscosity of 1.0% citrus pectin of total 10 ml to a half in 10 min at pH 4.0 and at 40°C). Both enzyme preparations were purchased from Hankyu Kyoei Co. Ltd., Osaka, Japan. The dry yeast used was for bread making which was manufactured by Oriental Yeast Co. Ltd., Tokyo, Japan.

Fig. 2. Photomicrographs of starch granules isolated from mangrove viviparous buds.
Left, without staining; right, after staining with iodine; magnif., ca. 800.
A, Rhizophora mucronata; B, Rhizophora apiculata; C, Ceriops tagal.
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Table 1. Starch granules in viviparous buds of several mangroves.

<table>
<thead>
<tr>
<th>Mangrove</th>
<th>Shape</th>
<th>Size (μm)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Rhizophora mucronata</em></td>
<td>Thailand</td>
<td>ellipsoidal</td>
</tr>
<tr>
<td></td>
<td>Okinawa, Japan</td>
<td></td>
</tr>
<tr>
<td><em>Rhizophora apiculata</em></td>
<td>round-oval</td>
<td></td>
</tr>
<tr>
<td><em>Ceriops tagal</em></td>
<td>round-oval</td>
<td>8-5 x 5-3</td>
</tr>
</tbody>
</table>

Table 2. Contents of starch and hemicellulosic polysaccharides of viviparous buds of mangroves.

<table>
<thead>
<tr>
<th>Species</th>
<th>Rhizophora mucronata</th>
<th>Rhizophora apiculata</th>
<th>Ceriops tagal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Moisture* (%)</td>
<td>50.5</td>
<td>47.1</td>
<td>62.9</td>
</tr>
<tr>
<td>Starch* (%)</td>
<td>13.6</td>
<td>13.9</td>
<td>6.87</td>
</tr>
<tr>
<td>Hemicellulosic polysaccharides* (%)</td>
<td>6.37</td>
<td>9.07</td>
<td>5.77</td>
</tr>
</tbody>
</table>

a Determined with a Kett infrared ray moisture analyzer.
b Starch content = (glucose produced by incubation with glucoamylase) x \( \frac{162}{180} \).
c Hemicellulosic polysaccharides = (total sugar content after hydrolysis with 1 N HCl at 100°C for 2 hr) - (glucose produced by incubation with glucoamylase x \( \frac{162}{180} \)).

RESULTS

1. Starch in viviparous buds

The results of staining of the viviparous bud section with iodine-potassium iodide are shown in Fig. 1, where the starch are seen as black spots. Figure 1 indicates that no special loci for starch granules to assemble are there. However, the starch granules mainly existed as a scrambled state like a bunch of grape and relatively uniformly distributed among fibrovascular bundles in viviparous bud.

The photomicrographs of starch granules isolated from the viviparous buds are shown in Fig. 2. The shape and size of the starch granules are summarized in Table 1. They were generally regular or irregular oval or ellipsoidal form resembling arrowroot starch. No cracks nor stripes as those found on corn starch or potato starch granules were observed. Their size varied to a considerable extent, depending on the kind of mangrove.

Upon gelatinization in boiling water, the starch granules were found to be hydrolyzed by glucoamylase with nearly the theoretical yield of glucose.

2. Starch content of viviparous buds

As shown in Table 2, the starch content of fresh viviparous buds amounted to about 14% for *R. mucronata* and *R. apiculata*. Therefore, the starch content of the former bud is estimated to be 28% on a dry basis of the sample. As to the content of hemicellulosic polysaccharides, *R. apiculata* was greater than other two mangroves. The other main solid materials were cellulose and inorganic substance.

3. Alcohol fermentation of viviparous buds

The results of alcohol fermentation performed by above method are shown in Fig. 3. The amount of carbon dioxide evolved was greatest with the fermented mash of *R. mucronata* viviparous buds. It is interesting that although the starch content of the buds of *C. tagal* was nearly a half of those of other two viviparous buds, the degree of carbon dioxide evolution was almost similar to that of *R. mucronata*. Nevertheless, the fermentation efficiency of *C. tagal* buds was only 60%.
DISCUSSION

When gelatinized, the granules isolated as starch from the viviparous buds were hydrolyzed by glucoamylase into glucose almost completely. Therefore, the granules were considered to be true starch. The viviparous buds of mangroves examined, however, seem to be a little too small in the starch content to be worth for calling the bud as a starchy organ.

The viviparous bud of *R. mucronata* collected in Okinawa, Japan, was fairly small as compared with that produced in Thailand. But no significant difference in the starch content and fermentability under the conditions were observed between the two viviparous buds. More research will of course be necessary to find the mangroves bearing viviparous bud of higher starch contents than those described above. Also, there may be the possibility to increase the starch content of the bud by breeding.

For alcohol fermentation, the viviparous buds were heated at 70°C. This was to inactivate polyphenol oxidase, otherwise the mash was colored in deep dark which decreased the activity of glucoamylase. Heating the buds at temperatures than the above value did not necessarily result in more favorable alcohol fermentation.

However, the fermentation efficiency of the viviparous buds was unreasonably low. This result may be partly due to the mode of existence of starch granule in the tissue of viviparous buds. The existence of tannic substance may be another reason. When the bud slices were boiled with 0.1 N sodium hydroxide for 10 min and then, neutralized, the alcohol fermentation of the mixture was extremely delayed and the fermentation efficiency was far less than those in the present paper.

The content of cellulosic substance of the viviparous buds used was about 15% when examined according to the method of Ost and Wilkening. It was interesting that cellulosic substance was readily separated by pressing the fermented mash and that the beer thus obtained from 500 g mash (250 g fresh bud and 250 g water) amounted 300–330 ml. The alcohol concentration of the beer from *R. mucronata* viviparous buds after fermentation for 5 days was 2.0% (v/v). If all the starch existed was completely fermented, the alcohol concentration would be increased two or three fold. For improving the alcohol fermentation of viviparous buds, further experiments under various condi-
tions will be necessary. The experiment to make clear the reason of difference in the alcohol fermentation efficiency between the viviparous buds of *R. apiculata* and *C. tagal* may lead to finding a clue for improvement of fermentation efficiency of the bud.

The cellulosic fiber isolated after alcoholic fermentation of the viviparous buds is under study to apply for methane fermentation which will be published elsewhere.

REFERENCES

1) P. A. SHAFFER and M. SOMOGYI: *J. Biol. Chem.*, 100, 695 (1933).