Antioxidant effects of antioxidant biofactor on reactive oxygen species in human gingival fibroblasts

Satoshi Matsui,1,2,* Yasuhsia Tsujimoto,1,2 Toshihiko Ozawa3 and Kiyoshi Matsushima1,2

1Department of Endodontics, Nihon University School of Dentistry at Matsudo, 870-1, Sakaecho, Nishi-2, Matsudo, Chiba 271-8587, Japan
2Research Institute of Oral Science, Nihon University School of Dentistry at Matsudo, Matsudo, Chiba 271-8587, Japan
3Department of Health Pharmacy, Yokohama College of Pharmacy, 601 Manataro, Totsuka-ku, Yokohama 245-0066, Kanagawa 245-0066, Japan

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The purpose of this study was to investigate the effects of antioxidant biofactor (AOB) on reactive oxygen species (ROS). Generation of superoxide radical (O₂⁻) and hydroxyl radical (‘OH) was determined using an electron spin resonance (ESR) spin-trapping method. AOB was added at different concentrations to these free radical generating systems. The generation of both O₂⁻ and ‘OH was scavenged by the addition of AOB in a dose-dependent manner. These results indicate that AOB has strong antioxidant properties against these radicals. We further investigated the anti-oxidative effect of AOB on human gingival fibroblasts (HGFs). HGFs were treated for 3 h with α-MEM containing a combination of AOB and H₂O₂ (AOB + H₂O₂ group), containing H₂O₂ (H₂O₂ group), or containing AOB alone (AOB group). Non-stimulated HGFs were used as a control group. The number of surviving cells in the order of the AOB group > control group > AOB + H₂O₂ group. The level of expression of type I collagen mRNA and production of collagen were also in the order of the AOB group > control group > AOB + H₂O₂ group. In conclusion, our results suggest that AOB may protect HGFs against oxidative stress by reducing stress-induced ROS.

Key Words: free radical, antioxidant biofactor, electron spin resonance spin-trapping method, periodontal disease, collagen

Reactive oxygen species (ROS) include superoxide anion (O₂⁻), hydrogen peroxide (H₂O₂) and hydroxyl radicals (‘OH) that are generated from an oxidative burst by enzymatic pathways of inflammatory cells.1,2 One well-known process of free radical production in the body is the transition from O₂⁻ to H₂O₂, which generates ‘OH.3 Excess production of ROS is considered to be the cause of various diseases. It has been reported that ROS is involved in apoptosis, inflammation, DNA damage and aging.4,5 ROS is also considered to be an inflammation factor in periodontal disease.6 ROS generated by bacteria and/or metal ions in the oral cavity may attack gingival tissue, periodontal ligaments and osteoblasts.7,8,9 Moreover, periodontal disease is a life style disease.10 Therefore, development of a technique to eliminate ROS in the oral cavity is necessary for the prevention of periodontal disease, and antioxidant treatment is beneficial for the treatment of periodontal disease.

Recently, foods have been developed with the aim of ROS elimination. One such food is a processed grain food termed antioxidant biofactor (AOB), which has strong antioxidant properties and inhibits cellular oxidant stress.11,12 AOB is a unique processed grain food. In addition, AOB is known to contain a variety of substances that have antioxidant activity, including flavonoids, α-tocopherol, vitamins, tannins, superoxide dismutase and catechins. Minamiyama et al.12 reported that AOB strongly inhibited the auto-oxidation of rat brain homogenates in vitro in a dose-dependent manner. They suggested that AOB has strong anti-oxidant properties and further additional biological effects, which might be of importance in the context of the prevention of degenerative diseases. Therefore, it is considered that AOB have a powerful mixture of scavenging activity on ROS of various kinds and improve immunity. However, there have been few studies of the anti-oxidative effects of the various components of AOB in periodontal disease. The present study was conducted to investigate the ability of AOB to scavenge both O₂⁻ and ‘OH radicals by using the electron spin resonance (ESR) spin-trapping method with 5,5-dimethyl-1-pyrroline-N-oxide (DMPO) in vitro. We further investigated cell viability, Type I collagen mRNA expression and the production of collagen following oxidative stress in the presence or absence of AOB in human gingival fibroblasts (HGFs).

Materials and Methods

Chemicals. AOB was obtained from AOJA (Kobe, Japan). H₂O₂ and TiO₂ were obtained from Wako Pure Chemicals (Osaka, Japan). DMPO (Dojin Chemicals, Kumamoto, Japan) was used as a spin-trapping agent for free radicals.

Adjustment of AOB. AOB (3 g) was mixed with pure water (PW: 27 ml) by vortexing and by exposure to a super sonic wave for 10 min. The solution was then allowed to stand, after which the supernatant fluid was collected and filtered as a 10% AOB solution. AOB was then diluted to the following concentrations: 0.16, 0.31, 0.63, 1.25, 2.50, 5.00 or 10.00 w/v%.

Materials. The visible light irradiation device used was the xenon light device (APOLLO 95E, Dental/Medical Diagnostic Systems Inc., Woodland Hills, CA), which was used at 440–500 nm wavelength.

O₂⁻ scavenging effects of AOB. O₂⁻ was generated for this experiment by reacting H₂O₂ with TiO₂ under visible light irradiation.12 The experimental protocol was as follows: 100 μl of a 6% H₂O₂ solution was mixed with 8 mg-TiO₂, 30 μl of PW, 50 μl of a 0–10% AOB solution and 20 μl of an 890 mM-DMPO solution in 96-well plates. After 15 s incubation, the mixture was irradiated with visible light for 6 s. After 1 min, the ESR spectrum was recorded using a JES FA200 (JEOL, Tokyo, Japan). The conditions for ESR were as follows: microwave power, 8 mW;
with the α-MEM growth medium supplemented with a combination of AOB (final concentration 0.001%) and H:O2 (final concentration 1 mM), and were then cultured for 3 h (AOB + H:O2 group). Other HGF groups were stimulated with 1 mM H:O2 for 3 h (H:O2 group) or with 0.001% AOB for 3 h (AOB group).

Non-stimulated cells were used as the control group. All groups were then washed with fresh α-MEM to remove the treatment medium.

**Assay of cell viability.** For assay of cell viability, HGFs were seeded into 96-well plates at a density of 4 × 10^4 cells/well in 0.1 ml of the appropriate medium containing α-MEM with 1% FBS and were cultured at 37°C for 24 h. The cells were then stimulated with AOB and/or H:O2.

Cell viability was measured 24, 48 or 72 h later by using the Cell Counting Kit-8 (CCK-8) assay (Dojindo Laboratories, Kumamoto, Japan). This assay is based on the conversion of the water-soluble tetrazolium salt, WST-8 [2-(2-methoxy-4-nitrophenyl)-5-(2,4-disulfophenyl)-2H-tetrazolium, monosodium salt] to a water-soluble formazan dye upon reduction in the presence of an electron carrier by dehydrogenases. The medium (100 μl) was incubated with 10 μl of the WST-8 solution for 2 h at 37°C. Absorbance was recorded at 540 nm on a microplate reader. Cell viability was expressed as a percent of the control culture value.

**RNA extraction and RT-PCR technique.** HGFs (1 × 10^4) were cultured in tissue culture dishes (10 cm) containing 10 ml of α-MEM supplemented with H:O2, AOB or AOB + H:O2 for 1 or 3 days. Total cellular RNA was extracted from the cells using an RNAeasy mini kit (QIAGEN, Hilden, Germany). The RNA isolation procedure was in accordance with the protocol provided with the RNAeasy mini kit. cDNA synthesis and amplification by RT-PCR were conducted using a One-Step RT-PCR kit. For the PCR mixture, RNA (200 ng) and oligonucleotide primers (500 nM) were used. The PCR primers used for PCR of Type I collagen and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) were designed with reference to the reported sequences, these primers were as follows: GAPDH (forward) 5'-ATG GTG CTA CT-3', (reverse) 5'-TTAG CAC CAG TGT CTC CAT GAG-3'; Type I collagen (forward) 5'-GTG GAA ATG ATC ACC ATC TTC CAG GAG-3', (reverse) 5'-GTG GTA GAA ATG GTG CTA CT-3'; Type II collagen (forward) 5'-GTG GTA GAA ATG GTG CTA CT-3'; and Type III collagen (forward) 5'-TTAG CAC CAG TGT CTC CT-3'.

The GeneAmp PCR system 9600 (PerkinElmer, Waltham, USA) was programmed for cDNA synthesis, and the procedure involved pre-denaturation for 30 min at 95°C, followed by 30 thermal cycles of denaturation for 30 s at 94°C, primer annealing for 30 s at 55°C, chain elongation for 30 s at 72°C, and final extension for 10 min at 72°C. The PCR fragments were electrophoresed on 1.5% agarose gels and were subsequently stained with ethidium bromide.

**Collagen production.** The collagen content of samples was estimated on day 3 to day 9 following treatment using the method described by Walsh et al. Briefly, type I collagen solution was diluted in either 50 mM HCl, 100 mM NaCl, or 10 mM CaCl2, together with 0.92% NaCl and 0.9% Na 3 (pH 7.5), or in culture medium. The standards and cell culture supernatants were dispensed into individual wells of microtiter plates and the plates were incubated at 37°C for 16 h (humidified) and then at 37°C (dry) for 24 h. After the plates were rinsed in distilled water, the wells were filled with 0.21% Sirius Red F3BA (Chromagellschaft Schmid GMBH Co., Stuttgart, Germany) in saturated picric acid (wt/vol), and the samples were stained for 1 h at room temperature. The plates were washed with 100 mM/L of NaOH. The eluted stain was then drawn up and down several times into a multichannel pipette and was then placed into a second plate. Absorbance was read at 540 nm using a model 2550 microplate reader (Bio-Rad Labs, Sunnyvale, CA), and the collagen content of the samples was estimated from the absorbance of the standards.
**Statistical analysis.** All values are presented as means ± SD, and the significance of differences was determined using the Bonferroni (p<0.05).

**Results**

**O·· scavenging activity of AOB.** To determine the antioxidant properties of AOB, we first tested if AOB can scavenge O·· species generated from the reaction of H$_2$O$_2$ with TiO$_2$ when irradiated by visible light. Fig. 1 shows representative ESR spectra of DMPO-spin adducts of O·· (DMPO-OH) generated in this way in the presence of different concentrations of AOB. A plot of the DMPO-OH signal intensity as a function of the concentration of AOB indicated that AOB can scavenge DMPO-OH in a dose-dependent manner.

**OH scavenging activity of AOB.** We next analyzed the effect of different concentrations of AOB on the ESR spectra of DMPO-spin adducts of ·OH (DMPO-OH) generated from the photolysis of H$_2$O$_2$ when irradiated by visible light. A plot of the DMPO-OH signal intensity as a function of the concentration of AOB indicated that AOB can also scavenge DMPO-OH in a dose-dependent manner (Fig. 2).

**Cell viability.** To determine if the elimination of H$_2$O$_2$ by AOB enhances HGF cell viability, the viability of cells exposed to H$_2$O$_2$ and/or AOB for 3 h was assayed using a commercial kit. Non stimulated cells were used as the control group. The cell viability of these HGFs was in the order of the AOB group > control group > AOB + H$_2$O$_2$ group > H$_2$O$_2$ group. The cell viability of the AOB group was significantly increased compared with that of the control group after 24, 48 or 72 h of treatment. The cell viability of the AOB + H$_2$O$_2$ group was significantly greater than that of the H$_2$O$_2$ group after 24, 48 or 72 h of treatment (Table 1) (b: p<0.05) (n = 6).

**mRNA expression of type I collagen.** We then determined if AOB enhancement of cell viability in the presence of H$_2$O$_2$ involved an effect on the expression of Type I collagen mRNA. Collagen is the main component of gingival tissue and its production is inhibited following periodontal disease. The effect of exposure of HGFs to H$_2$O$_2$ and/or AOB for 1 or 3 days on the mRNA expression of Type I collagen was analyzed using RT-PCR. Type I collagen mRNA expression was increased in cells treated only with AOB for 1 or 3 days compared to the control group. The level of expression of Type I collagen mRNA was in the order of the AOB group > control group > AOB + H$_2$O$_2$ group > H$_2$O$_2$ group (Fig. 3).

**Collagen production.** To confirm that the production of collagen correlated with collagen protein levels, the collagen content of the treated cells was assayed from day 3 to day 9 after treatment with AOB and/or H$_2$O$_2$ using a colorimetric assay. The production of collagen increased in all groups over this time period and peaked on day 9. The production of collagen was significantly greater in the AOB group than in the control group on days 6 and 9, and was also significantly higher in the AOB + H$_2$O$_2$ group than in the H$_2$O$_2$ group on days 6 and 9 (Fig. 4) (p<0.05) (n = 7).

**Discussion**

Excess production of ROS is considered to be the cause of various diseases. Further, it has been reported that free radicals

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**Table 1. Effect of ROS on cell viability in the presence or absence of AOB**

<table>
<thead>
<tr>
<th></th>
<th>24 h</th>
<th>48 h</th>
<th>72 h</th>
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<tbody>
<tr>
<td>control</td>
<td>100.00 ± 2.45</td>
<td>100.00 ± 4.16</td>
<td>100.00 ± 3.02</td>
</tr>
<tr>
<td>H$_2$O$_2$</td>
<td>86.53 ± 3.68$^b$</td>
<td>79.98 ± 2.12$^b$</td>
<td>73.68 ± 1.94$^b$</td>
</tr>
<tr>
<td>AOB + H$_2$O$_2$</td>
<td>93.53 ± 2.74$^b$</td>
<td>85.65 ± 4.69$^b$</td>
<td>83.66 ± 2.74$^b$</td>
</tr>
<tr>
<td>AOB</td>
<td>110.43 ± 3.19$^a$</td>
<td>108.75 ± 1.98$^a$</td>
<td>109.65 ± 1.62$^a$</td>
</tr>
</tbody>
</table>

*a: Significantly (p<0.05) control vs H$_2$O$_2$ or AOB.

*b: Significantly (p<0.05) H$_2$O$_2$ vs H$_2$O$_2$ + AOB.

$n = 6$.

HGFs were incubated with AOB, H$_2$O$_2$, AOB + H$_2$O$_2$ or with buffer control for the indicated times.

Cell viability was then assessed using cell counting kit-8. The viability of the AOB + H$_2$O$_2$ group was significantly greater than that of the H$_2$O$_2$ group, and the viability of the AOB group was significantly higher than the control group, at 24, 48 or 72 h after treatment (b: p<0.05).
are involved in tumorigenesis, apoptosis, inflammation, DNA damage, and aging. Therefore, health food is taken into consideration for deletion of ROS. Since AOB has strong antioxidant properties and biological effectiveness against ROS, AOB has recently received a lot of attention as a health food for the elimination of ROS. However, there have been few studies of the anti-oxidation ability of the various ingredients contained in AOB. Furthermore, there have been few reports concerning the free radical scavenging ability of AOB. Therefore, an analysis of the ability of AOB to scavenge O$_2^-$ and 'OH is required.

In this study, we demonstrated the ability of AOB to scavenge O$_2^-$ and 'OH radicals by using an in vitro ESR spin-trapping method and we further showed that AOB can protect cells against H$_2$O$_2$ inhibition of cell viability and collagen production. Using this ESR spin-trapping method, and in vitro O$_2^-$ and 'OH generating systems, we showed that AOB can strongly suppress the generation of these free radicals by visible light irradiation in vitro in a dose-dependent manner. It is well known that the components of AOB include substances such as flavonoids, α-tocopherol, tannins, catechins, and vitamins. Kashima has reported that the generation of O$_2^-$ and 'OH from the reaction of H$_2$O$_2$ with metal ions, which is similar to our in vitro method for generation of these radicals, was suppressed by catechin. Furthermore, it has been reported that ROS is eliminated by flavonoid and α-tocopherol. These data suggest that the ability of AOB to scavenge O$_2^-$ and 'OH that we observed in our study was most likely mediated by more than one component of AOB.

Periodontal disease is a bacterially induced inflammatory disease that is accompanied by gingival inflammation and alveolar bone loss. The tissue damage induced by periodontal disease is directly caused by bacteria and is indirectly caused by inflammation and by an immune response against these bacteria. During the inflammatory process, activated phagocytes such as neutrophils and macrophages produce large quantities of ROS via NADPH oxidase. Moreover, patients with periodontal disease display increased neutrophil numbers and activity. In this respect it is of interest that our results showed that AOB can protect HGFs against damage as a result of oxidant stress. Thus, cells treated with AOB had a higher viability than control cells, and cells exposed to H$_2$O$_2$ had a significantly higher viability if AOB was present. Moreover, not only did AOB increase cell viability, but we showed that AOB also significantly enhances the production of both type I collagen mRNA and collagen protein, and protects HGFs against the inhibition of collagen production induced by H$_2$O$_2$. Since collagen is the main component of gingival tissue, and the progress of periodontal disease is associated with decreased collagen production, our data further suggest that AOB may be useful for combating periodontal disease. Our data further suggest that ROS may play a role in the breakdown of the collagen network in gingivitis. Tanaka et al. reported that ROS is an important inflammatory factor for HGFs. Our data are also consistent with other studies that have shown that AOB can decrease cell damage and the destruction of collagen due to ROS in HGFs. The combined data suggest that AOB may collaborate with signals such as growth factors that modulate the synthesis of collagen in HGFs. However, in our study, we did not describe the relationship between expression of matrix metalloproteinases (MMPs) and ROS in HGFs. MMPs are main factor in destruction of the collagen in gingival tissue. Therefore, we are investigating the inhibitory effect of AOB on expression of MMPs caused by ROS in HGFs. Further investigations into this are currently under way in our laboratory.

Acknowledgment

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Abbreviations

AOB antioxidant biofactor
ESR electron spin resonance
O$_2^-$ superoxide anion
'OH hydroxyl radical
MMPs matrix metalloproteinases

Fig. 3. mRNA expression of type I collagen after treatment with H$_2$O$_2$ in the presence or absence of AOB. HGFs were incubated with AOB, H$_2$O$_2$, AOB + H$_2$O$_2$ or with buffer control for 1 or 3 days. Type I collagen mRNA expression was then assessed using RT-PCR. Type I collagen mRNA expression was much greatest in the AOB group than in other group at 1 or 3 days after treatment, and greater in the AOB + H$_2$O$_2$ group than in H$_2$O$_2$ group at 1 or 3 days after treatment.

Fig. 4. Production of collagen after treatment with H$_2$O$_2$ in the presence or absence of AOB. HGFs were incubated with AOB, H$_2$O$_2$, AOB + H$_2$O$_2$, with control for 3, 6, or 9 days. Collagen production was then assessed using a colorimetric assay. Collagen production was significantly greater in the AOB + H$_2$O$_2$ group than in the H$_2$O$_2$ group on 6 or 9 days after treatment and was significantly greater in the AOB group than in the control group on 6 or 9 days. However, collagen production was not significantly greater in the AOB + H$_2$O$_2$ group than in the control group on 3, 6 or 9 days.
References


