Alcohol is an oxidative stressor for gastric epithelial cells: detection of superoxide in living cells

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Alcohol/ethanol has been reported to derived necrosis and apoptosis with an oxidative stress in gastric mucosal cells. However the clear evidence for reactive oxygen species (ROS) generation by alcohol in gastric cells in vitro is none. In this study, we elucidated ethanol is an oxidative stress inducer on rat gastric epithelial cells by electron paramagnetic resonance measurement in living cells. We also confirmed whether ethanol-induced cellular ROS was derived from mitochondria or not. The results of cellular ROS determination showed that an increment of cellular ROS was shown for 15 min from exposing 1% (v/v) ethanol. Lipid peroxidation in cellular membrane also induced by 1% ethanol and the tendency is same in the results of cellular ROS determination. JC-1 stained showed the decrement of mitochondrial membrane potential. Additionally the localization of cellular ROS coincided with mitochondria. These results indicated that ethanol is not merely a necrotizing factor for gastric epithelial cells, but also an oxidative stress inducer via injured mitochondria. Key Words: alcohol, reactive oxygen species, mitochondria, stomach cells, Electron Paramagnetic Resonance

A lcohol/ethanol is an aggressive factor for gastrointestinal tract. The favorite alcohol like a beer or wine is contained 4–20% ethanol. World Health Organization (WHO) reported that alcohol is a casual factor in 60 types of both diseases and injuries and component causes in 200 others. Almost 4% of all deaths worldwide attributed to alcohol, which was about 2.25 million in 2004. Global distribution of deaths (%) related in gastrointestinal tract were 7.0% of esophagus cancer, 3.4% of mouth/oropharynx cancers, and 0.8% of colon/rectum cancers in 2004.(1) Commonly, alcohol consumers are suffering from a bout of heartburn and nausea. Reflux of caustic gastric contents, reactive oxygen species (ROS) such as superoxide radical and hydroxyl radical and release of lysosomal enzymes, is known to directly or indirectly cause symptoms such as heartburn and nausea.(2,3) These symptoms suggested that alcohol is probably an oxidative stress. In liver, toxic effects of alcohol have long been studying because excessive consumption of alcohol worldwide attributed to alcohol, which was about 22.5 million in 2004. Globally, 8% of all deaths related to alcohol in 2004.(4) Alcohol-induced liver disease, such as alcoholic liver disease and alcoholic hepatitis, are major causes of death and disability worldwide. Alcoholic liver disease is the single most important cause of death of liver disease. Alcohol/ethanol has been reported to derived necrosis and apoptosis with an oxidative stress in gastric mucosal cells. However the clear evidence for reactive oxygen species (ROS) generation by alcohol in gastric cells in vitro is none. In this study, we elucidated ethanol is an oxidative stress inducer on rat gastric epithelial cells by electron paramagnetic resonance measurement in living cells. We also confirmed whether ethanol-induced cellular ROS was derived from mitochondria or not. The results of cellular ROS determination showed that an increment of cellular ROS was shown for 15 min from exposing 1% (v/v) ethanol. Lipid peroxidation in cellular membrane also induced by 1% ethanol and the tendency is same in the results of cellular ROS determination. JC-1 stained showed the decrement of mitochondrial membrane potential. Additionally the localization of cellular ROS coincided with mitochondria. These results indicated that ethanol is not merely a necrotizing factor for gastric epithelial cells, but also an oxidative stress inducer via injured mitochondria.

Key Words: alcohol, reactive oxygen species, mitochondria, stomach cells, Electron Paramagnetic Resonance

Materials and Methods

Materials. Aminophenyl Fluorescein (APF) (SEKISUI MEDICAL CO., LTD., Tokyo, Japan), 2-[5,5-Dimethyl-2-oxo-2H-[1,3,2]dioxaphosphinan-2-yl]-2-methyl-3,4-dihydro-2H-pyrrole-1-oxide (CYP-MOPO) (Radical Research Inc., Tokyo, Japan), β-Nicotinamide adenine dinucleotide (NADH) (SIGMA), D-Glutamic acid (SIGMA), Malic Acid (Wako Pure Chem. Ind. Ltd., Osaka, Japan), Succinic acid (SIGMA-ALDRICH), Diphenyl-1-prenylphosphine (DPPP) (DOJINDO, Kumamoto, Japan), Cell counting kit-8 (DOJINDO), MitoRed (DOJINDO) and Ethanol (Wako) were purchased. Alcohol-contained culture medium was prepared by mixing alcohol, and the culture medium was used after filter-sterilized (Millex 0.22 μm, Millipore Co., Billerica, State).

Cell culture. RGM-1 was cultured in DMEM/F12 (Gibco). This culture medium contained 10% inactivated FBS and 1% penicillin/streptomycin. Cells were cultured in 5% CO2 cell culture incubator at 37°C.

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**Cell viability test by WST assay.** Cell viability test was examined with cell counting kit-8 according to the manufacturer’s instructions. RGM-1 was dispersed in the 96-well dish at 10,000 cells/well and it was incubated for overnight. The medium was replaced to the alcohol-contained culture medium which contained ethanol of 0, 0.1, 0.2, 0.5, 1, 2, 5, 10, 15, 20% (v/v) and it was incubated for 0, 3, and 24 h. After incubation, medium was replaced to the medium contained 10% cell counting kit-8 of 100 μL and cells were incubated for 1 h. The absorbance of 450 nm was measured by Varioskan plate reader (Thermo Fisher Scientific K. K., Kanagawa, Japan).

**Lipid peroxide determination by DPPP.** The lipid peroxidation was measured by DPPP as follows; Cells were dispersed at the concentration of 31,250 cells/cm², and cells were incubated for 24 h. The culture medium was thereafter replaced to the culture medium contained 10 μM DPPP. After incubated for 15 min, cells were washed twice with cold PBS. The fluorescence intensities at Ex. 352 nm and Em. 380 nm of DPPP were measured by the plate reader.

**Intracellular ROS determination by APF.** Free radicals (hydroxyl radical and peroxynitrite) were detected by APF as follows; APF was diluted with PBS and it exposed to the cells at the concentration of 1 μM for 30 min. After incubation, cells were washed using a cold PBS twice. The intensities of APF-fluorescent were measured by Varioskan at Ex. 490 nm and Em. 515 nm. Cellular fluorescent images were observed with a chilled CCD camera (AxioCam color, ZEISS, Germany)-mounted epifluorescence microscope (Axiovert 135M, ZEISS) connected to an image analyzing system (AxioVision, ZEISS).

**The determination of the area at ROS-production in the cell.** Cells were cultured on the 35 mm² glass-bottom dish at the density of 15,000 cells/cm². Cells were exposed to 0, 1 and 5% ethanol/medium for 1 h. Medium was replaced to PBS with 10 μM APF and 200 nM MitoRed. Cells were incubated for 30 min. After washed with cold PBS twice, cells were taken pictures using confocal microscopy (LSM700, ZEISS).

**Electron paramagnetic resonance (EPR) measurement.** The methods of EPR measurement were consulted previous reports and so on. Free radicals were detected by EPR measurement using spin-trapping agents (CYPMPO). Fig. 2 showed the results of cellular ROS determination by APF. Cellular ROS was significantly increased under the condition of 1–5% (v/v) ethanol after incubation for 1 h. 15–20% ethanol-exposed cells were completely death. Fig. 3 showed EPR signals obtained under the following conditions: 10 mW incident microwave power, 0.1 mT modulation width, 8 min sweep time, 7.5 mT sweep width, 0.1 s time contrast, 333.5 mT center field, and 15 mT scan range. Spectral computer simulation was performed using a Win-Rad Radical Analyzer System (Radical Research).

**Static analysis.** Significant static value (p value) was calculated using ANOVA followed by Turkey HSD.

**Results**

**Ethanol induced the cell death.** The cell viability after exposing ethanol was determined by cell viability test in comparison with the normal rat gastric mucosa cells (RGM-1). Fig. 1 showed 1% ethanol had cytotoxicity for 24 h. RGM-1 died completely in the medium contained one hour exposure of more 15% (v/v) ethanol, and we decided that necrosis was involved on these cells. On the other hand, the cells survived environments under less than 10% ethanol for several hours suggested that another kind of death was derived on these cells.

**Ethanol induced cellular ROS.** The ROS concentration from the cells was determined by both the AFP study and the EPR measurement using spin-trapping agents (CYPMPO). Fig. 2 showed the results of cellular ROS determination by APF. Cellular ROS was increased with ethanol concentration. The amounts of cellular ROS in 1–5% ethanol exposing cells was significantly higher than that in the control cells. Fig. 3 showed EPR signals...
intensities in RGM-1 exposed different concentrations of ethanol. These results showed the 15 min exposure of 1% ethanol induced ROS from the RGM-1.

**Ethanol induced lipid peroxidation.** Fig. 4 showed the amounts of lipid peroxidation in cellular membrane after one hour exposure with ethanol. The graph shows the intensity of DPPP fluorescence. These results showed same tendency of the results in APF measurements (Fig. 2).

**Ethanol injured mitochondrion.** We confirmed that ethanol injured mitochondria. Fluorescence characteristics of JC-1 were changed in accordance with mitochondrial membrane potential dependence. Green fluorescence and Red fluorescence of JC-1 means injured mitochondria (decreasing membrane potential) and healthy mitochondria (normal membrane potential), respectively. Fig. 5 shows the results of JC-1 stained. Injured mitochondria were showed in 5% ethanol exposed cells (Fig. 5A and C). These results indicated that ethanol injured mitochondrion.

**Mitochondria produced ROS by exposing ethanol.** We confirmed that the localization of ROS and mitochondria due to clarify the ROS production site. Fig. 6 is pictures of stained-cells with APF and MitoRed. The cells were exposed with 0, 1 and 5% ethanol for 1 h. The MitoRed fluorescence coincided with the APF fluorescence. This result indicated that mitochondria were the ROS production organelle after the ethanol exposure.

**Discussion**

In this study, we demonstrated for the first time that ethanol treatments involved reactive ROS production, in particular superoxide anion, in gastric epithelial cells.

We performed this study under the condition from 0 to 20%, which is popular alcohol’s ethanol concentration. More than 15% ethanol exposure derived immediate cell death within 1 h, while cells were survived for a few hours under less than 10% ethanol condition although cellular ROS production was involved. We thus proposed that high concentration of ethanol more than 15% was a necrotizing factor, while moderate concentration of ethanol was an oxidative stress. Suzuki et al.\(^\text{(23)}\) have demonstrated that the effects of ethanol may be associated with a disturbance in the balance between gastric mucosal protective and aggressive factors. In fact, administration of a low dose ethanol have been reported to protect the gastric mucosa from gastric lesions;\(^\text{24–26}\) however, it lead to apoptotic cell death *in vitro*.\(^\text{27}\) Gastrointestinal tracts including the stomach are called the first-pass metabolism of alcohol. In the metabolism, microsomal ethanol oxidizing
system (MEOS) requires CYP2E1 (cytochrome P450 family) for generating oxidized NADPH, which used to localize in cytoplasm. CYP2E1 accelerates the expression of cyclooxygenase-2 (COX-2) in liver. COX-2 produces prostaglandins, and it should protect gastric lesions in vivo. However, there are few reports investigating the relations between ethanol-induced ROS and mitochondrion. We have demonstrated that NSAIDs and bisphosphonate involved superoxide anion production by EPR measurement using separated mitochondria. EPR measurement with living cells also proved that gastric acid is a mitochondrial superoxide anion inducer, whereas it has been generally accepted as a necrotizing factor. The pictures of microscopic observation for ethanol-exposing cells showed the decrement of mitochondrial membrane potential (Fig. 5) and co-localization of mitochondria and APF-stained cellular ROS (Fig. 6). Thus we concluded that ethanol inhibited a mitochondrial electron transfer system to involve superoxide anion production. Mitochondrial ROS was likely to play a role to derive the cellular injury. Mitochondrial ROS has been reported to be related with many diseases. As one example, several reports indicates that mitochondrial ROS production enhances tumor specific properties. Mitochondrial ROS also indicates the relation with the expression of oncogene expression.

In our life, alcohol is diluted with the contents in the stomach. Additionally, almost alcohol contains some antioxidant like polyphenols, resveratrol and flavonoid. These antioxidants were expected that it has health effects for healthy and affected individual. They should play a part of defense against

Fig. 5. Mitochondrial injury by exposing ethanol. Mitochondrial injury was measured by JC-1 after exposed-ethanol for 1 h. Green fluorescence (Ex. 485 nm, Em. 535 nm) and red fluorescence (Ex. 560 nm, Em. 595 nm) show mitochondrial injury and healthy mitochondria, respectively. Muted mitochondria were significantly increased under the condition of 5% (v/v) ethanol. (A) Confocal microscopic fluorescent images, (B) Red fluorescence intensity of JC-1, (C) Green fluorescence intensity of JC-1. The fluorescence intensity was measured by plate reader. n = 4, Error bar; SD.
alcohol-toxicity. In fact, resveratrol in red wine suppressed ethanol-induced cytotoxicity comparison with white wine and beer.\(^{(27)}\) Recently, new chemicals with the capacity to scavenge ROS were synthesized.\(^{(39–41)}\) We expected to contribute to develop the prevention of alcoholic harmful effects.

In conclusion, ethanol is not merely a necrotizing factor for gastric epithelial cells, but also an oxidative stress inducer. ROS after ethanol treatment were involved from mitochondria. Now we are undergoing the study to clear the relations between ethanol-induced ROS and carcinogenesis in gastric epithelial cells.

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**Conflict of Interest**

No potential conflicts of interest were disclosed.

**References**


