Boysenberry Polyphenols Suppressed Elevation of Plasma Triglyceride Levels in Rats

Shigeru Mineo1, Akane Noguchi1, Yuta Nagakura1, Kinji Kobori1, Tatsuo Ohta2, Ei Sakaguchi1 and Takashi Ichiyanagi3,4

1Bourbon Institutes of Health, Bourbon Corporation, 1–3–1 Ekimae, Kashiwazaki, Niigata 945–8611, Japan
2Department of Applied Life Sciences, Niigata University of Pharmacy and Applied Life Sciences, Akiha-ku, Niigata 956–8603, Japan
3Graduate School of Natural Science and Technology, Okayama University, 1–1–1 Tsushima-naka, Okayama, Okayama 700–8530, Japan
4Department of Environmental Science, Niigata Institute of Technology, 1719 Fujihashi, Kashiwazaki, Niigata 945–1195, Japan

(Received November 5, 2014)

Summary Boysenberry, a hybrid Rubus berry, is mainly cultivated in New Zealand. We previously reported that consumption of boysenberry juice (BBJ) exhibited anti-obesity effects in high-fat feeding rats. In this study, we focused on the suppressive effect of BBJ and its fraction on triglyceride absorption from the gastrointestinal tract. BBJ effectively inhibited pancreatic lipase activity in vitro, and was separated into four fractions (Fr1, Fr2, Fr3 and Fr4) by HP-20 column chromatography. Among all the fractions, Fr3, the ellagic acid-rich fraction, showed the most potent inhibition against pancreatic lipase in vitro with Fr2, the anthocyanin-rich fraction, second. Authentic ellagic acid equivalent in Fr3 showed poor activity against pancreatic lipase. Then, each fraction was orally administered with corn oil to rats fitted with a jugular catheter to examine the effects of each fraction on plasma triglyceride levels. Both Fr2 and Fr3 effectively suppressed the plasma triglyceride level elevation at a dose of 1,000 mg/kg body weight. These findings demonstrated that BBJ contains chemical components which inhibit triglyceride absorption from the gastrointestinal tract.

Key Words boysenberry, anti-obesity effect, pancreatic lipase, polyphenol, lipid absorption

MATERIALS AND METHODS

Reagents. Ellagic acid and cholic acid were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan). 4-Methylumbelliferyl oleate (4MUO) and lipase from porcine pancreas were purchased from Sigma-Aldrich (Tokyo, Japan). A corn oil manufactured by Ajinomoto (Tokyo, Japan) was used for in vivo experiments. Cyanidin-3-O-β-D-glucoside chloride was purchased from Tokiwa Phytochemical Co., Ltd. (Chiba, Japan). Other reagents were purchased from Kanto Chemical Co., Inc. (Tokyo, Japan) without further purification. BBJ was obtained from Berryfruit Export NZ, Ltd. (Nelson, New Zealand) (Bx 65.1–65.2) together with other berry juices (blueberry, blackcurrant, red rasp-
Suppression of Triglyceride Levels by Boysenberry

Boysenberry (Rubus idaeus L., Rubus fruticosus agg., and grape) (Bx 65.1–65.2˚) for comparison. Each juice was stored at −18˚C until examination.

Fractionation of BBJ. BBJ was diluted with distilled water and loaded onto a column packed with Diaion® HP-20 resin (Mitsubishi Chemical Corporation, Tokyo, Japan). The column was washed with distilled water, and components of BBJ were subsequently recovered with ethanol. The water fraction (Fr1) was freeze-dried. The ethanol fraction was concentrated to remove the organic solvent in vacuo at 40˚C; then the remaining water fraction was removed and powdered by freeze-drying. The ethanol fraction was resolved in distilled water and loaded onto the same column to achieve further separation. After washing the column with distilled water, two fractions (Fr2 and Fr3) were successively recovered with 50% aqueous methanol and 100% methanol. The remaining components in the column were recovered with acetone (Fr4). The organic solvent in each fraction was removed with the same procedures described above and was powdered by freeze-drying.

Inhibitory effects on pancreatic lipase. The lipase-inhibitory effects of BBJ were evaluated by the method of Kurihara et al. (14), with slight modifications. Briefly, 20-μL aliquots of various juices were diluted with sodium carbonate adjusted to pH 7.4, and solutions of the above fractions (0–8 mg/mL) were individually combined with 130 μL of pancreatic lipase solution in 0.2 μL phosphate buffer (0.6 mg/mL) and incubated for 20 min at 37˚C. Next, 5 μL of 4MUO solution (1 mM) was added to the mixtures. After incubation for 30 min at 37˚C, 50 μL of 5 M acetic acid was added to stop the reaction. The fluorescence generated by 4MUO was measured using a fluorescence analyzer (Infinite M200; Tecan Japan Ltd., Kanagawa, Japan) with an excitation wavelength of 327 nm and an emission wavelength of 449 nm. The inhibition of pancreatic lipase activity was calculated as a percentage against the control reaction. The 50% inhibitory concentration (IC50) value of each fraction was calculated from a mathematical formula obtained from linear approximation of curves as the logarithm of the concentration of each fraction versus the inhibitory activity.

Animal treatment. Male Wistar rats (7 wk of age) purchased from Japan SLC, Inc. (Shizuoka, Japan) were housed in an air-conditioned room (22 ± 2˚C) with a 12-h/12-h light/dark cycle. The rats were allowed free access to tap water and a commercial diet (Rabo-MR Stock, Nihon Nosan Kogyo, Tokyo, Japan) for 6 d before the experiment.

After the 6-d conditioning period, 18 rats were cannulated with a polyethylene tube (PE-50) in the jugular vein under anesthesia with diethyl ether. Diethyl ether was inhaled for quick recovery of the rats from anesthesia before the administration study. The right jugular vein was isolated, and a small hole was made using scissors to insert the polyethylene tube. The tip of the tube was set on exactly the correct position of entrance of the right auricle of the rats. After insertion of the tube, the vein was occluded, and the tube penetrated the skin and was guided out from the back of the rat. After fast-

![Fig. 1. Anthocyanins and ellagic acid in boysenberry.](image)
ing for 20 h, the rats were divided into three groups to make the mean body weight equal in each group. Emulsions were prepared by ultrasonic mixing of 5 mL of corn oil containing 200 mg of cholic acid and 5 mL of each (none, Fr2 and Fr3) fraction dissolved in distilled water to adjust to an administration dose of 1,000 mg/kg. Then, each emulsion was quickly administered to a different group of rats by direct stomach intubation (10 mL/kg body weight).

Blood samples (0.2 mL) were taken from each rat via the jugular catheter using a heparinized syringe at hourly intervals over a period of 10 h, and centrifuged at 15,500 g for 10 min to obtain blood plasma. Then, concentrations of triglyceride (TG) in the blood plasma were quantified with a triglyceride test kit (Triglyceride E-test Wako; Wako Pure Chemical Industries). The area under the plasma concentration–time curve (AUC) was calculated from the time-dependent plasma TG profile over the 10-h period following administration. The animals were maintained according to the Guidelines for Animal Experimentation of Okayama University, and the protocol was approved by the Ethical Committee at Okayama University.

Analysis of components in BBJ fraction. Qualitative and quantitative analyses of anthocyanins and ellagic acid were performed using a Shimadzu Prominence system (Shimadzu Corp., Kyoto, Japan) equipped with L-column 2 ODS (2.1×100 mm, 2 μm) (Chemicals Evaluation and Research Institute, Tokyo, Japan) at 40°C. Compounds were eluted with a gradient system using 1% aqueous formic acid (A) and 1% formic acid in acetonitrile (B) as the elution solvent. The gradient system was as follows: 8% B at 0 min and a linear gradient of 18% B at 30 min at a flow rate of 0.2 mL/min. The injection volume of samples was 5 μL. The eluted components were monitored with a UV-Vis detector at 520 nm for anthocyanins and 350 nm for ellagic tannin. Anthocyanins and ellagic acid were quantified with commercially available authentic cyanidin 3-O-β-D-glucoside equivalent and ellagic acid. The eluent was further monitored with a triple quadrupole mass spectrometer, API 3200 (AB SCIEX, Framingham, MA), to obtain molecular and product ions. Total ion chromatograms were recorded over a mass range of \textit{m/z} 250 to \textit{m/z} 800 using a scan duration of 1 amu. Peaks giving \textit{m/z} values corresponding to possible anthocyanins in total ion chromatograms were further investigated by the product ion scan. Individual precursors and product ions were ionized via electrospray ionization (ESI) operated in the positive ion mode. Curtain gas and collision gas were applied as \textit{N}_2. The ion spray voltage and temperature were set at 5.5 kV and 400°C.

Statistics. The results of the in vitro assays are expressed as the mean±standard deviation (SD), while those of the in vivo assays are expressed as the mean±standard error (SE). The data were analyzed using a single factorial design for each sampling time, and examined for statistically significant differences by Tukey’s multiple range test (Statcel 2 software; OMS Publishing, Saitama, Japan). A probability value of less than 0.05 was considered statistically significant.

RESULTS

Inhibitory effects of various berry juices on pancreatic lipase

BBJ showed the most potent inhibition among the examined juices at a dilution range from 1.25×10⁻³ to 1×10⁻². Furthermore, BBJ, blueberry and blackcurrant juices showed similarly high inhibitory effects of more than 80% at a dilution of 2×10⁻², when the activity of the control was assumed to be 100% (Fig. 2).

Polyphenol components in BBJ fraction

BBJ, which showed the highest inhibition against pancreatic lipase, was separated into four fractions by HP-20 column chromatography. The recovery of each fraction was 55.3 g for Fr2, 20.1 g for Fr3 and 1.1 g for Fr4, when 1,500 g of BBJ was treated. Qualitative
and quantitative analyses demonstrated both Fr2 and Fr4 contained high amounts of anthocyanins. HPLC-UV-MS/MS analysis showed that peaks 1, 2 and 3 had a molecular ion (M*) at m/z 611, 449 and 757, respectively, and a product ion (aglycone*) at m/z 287. Peak 4 had a molecular ion (M*) at m/z at 595. The total anthocyanin contents in the fractions were 111 mg/g for Fr2, 2.03 mg/g for Fr3 and 54.8 mg/g for Fr4 (Table 1). Similarly, contents of ellagic acid in each fraction were determined as 1.21, 108 and 0.21 mg/g for Fr2, Fr3 and Fr4, respectively.

**Inhibitory effects of BBJ fractions on pancreatic lipase**

Pancreatic lipase-inhibitory activity of BBJ fractions was evaluated in the same manner using the assay comparing the different berry juices. Fr1 did not inhibit pancreatic lipase (IC₅₀ > 0.8 mg/mL), while the other fractions inhibited pancreatic lipase in a concentration-dependent manner. The inhibition of lipase activity was as follows: Fr3 > Fr2 = Fr4 (IC₅₀ = 0.17, 0.33 and 0.36 mg/mL, respectively) (Table 1). Because Fr3 contained the highest amount of ellagic acid, lipase-inhibitory activity also was examined for authentic ellagic acid. However, the authentic ellagic acid equivalent in Fr3 did not contribute to the inhibition of pancreatic lipase activity observed for Fr3 (data not shown).

**Effects of BBJ fractions on plasma TG levels in rats**

The plasma TG levels in the control group reached a maximum of 129.9±21.6 mg/dL 4 h after oral administration of the emulsion. The Fr2 and Fr3 groups both showed significant (p<0.05) suppression of the elevation of plasma TG levels 3 h after the administration relative to the control group. Furthermore, both groups again showed significant (p<0.05) suppression of plasma TG levels relative to the control group 6 to 7 h after oral administration (Fig. 3A). The AUC levels of TG after administration of the emulsion containing Fr2 and Fr3 also were suppressed significantly (p<0.05 and p<0.01, respectively) compared with that of the control group (Fig. 3B).

**DISCUSSION**

Recent studies have demonstrated that several polyphenols such as flavanols (4), procyanidins (15) and hesperidin (16) showed potent lipase inhibition in vitro. Thus, consumption of food containing various polyphenols has attracted much attention to prevent obesity caused by a high-fat diet. We have previously reported BBJ significantly lowered plasma TG levels together with active EAS and cholesterol levels in the liver, leading to the suppression of elevations of total body weight and body fat in an obesity rat model induced by a high-fat diet including 30% lard (13). These findings indicate that BBJ contains beneficial components associated with anti-obesity effects. BBJ contains high amounts of various polyphenolic components; therefore, in the present study, we focused on polyphenolic components in BBJ and obtained several fractions by column chromatography to evaluate lipase inhibition in vitro and suppression of plasma TG levels after a single oral administration of corn oil with BBJ fractions in rats. BBJ exerted the most potent inhibitory effects among all examined juices at a dilution range of 1.25×10⁻³ to 1×10⁻². Therefore, BBJ was separated into several fractions by column chromatography for the prediction of active components. Both Fr2 and Fr3 showed strong lipase inhibition, while Fr1 recovered with distilled water including carbohydrates and organic acids showed poor activity.

Tandem mass spectrometry results revealed that peaks 1, 2 and 3 had a molecular ion (M*) at m/z=611, 449 and 757, respectively, and a product ion (aglycone*) at m/z=287, which indicates carrying cyanidin as aglycone. The molecular ion (M*) of peak 4 was obtained at m/z=595, although the product ion could not be observed. The anthocyanins in boysenberry were previously reported elsewhere (8). Taking the present results and previous reports into consideration, four anthocyanins in the BBJ fractions obtained in this study were determined as cyanidin-3-[2-(glucosyl)glucoside] for peak 1, cyanidin-3-glucoside for peak 2, cyanidin-3-[6-(rhamnosyl)glucoside] for peak 3, and cyanidin-3-[2-(glucosyl)-6-(rhamnosyl)glucoside] for peak 4.
3-[(2-glucosyl)-6-(rhamnosyl)glucoside] for peak 3 and cyanidin-3-[(6-rhamnosyl)glucoside] for peak 4.

Fr3, the ellagic acid-rich fraction, expressed the most potent inhibitory activity among all examined fractions, although the authentic ellagic acid equivalent to Fr3 showed poor inhibitory effects on pancreatic lipase. This indicates the presence of other potent inhibitors of pancreatic lipase in Fr3. Furuuchi et al. (8) reported boysenberry contains quercetin glycosides and propelargonidin. Quercetins showed anti-obesity effects (17) based on inhibition of adipogenesis and apoptosis induction in 3T3-L1 cells (18), while inhibition by these polyphenols of lipid absorption from the gastrointestinal tract remains unclear. Therefore, these polyphenols may contribute to the lipase inhibitory activity of Fr3, although further studies are required to clarify the presence of these components in the fraction.

BBJ fractions (Fr2 and Fr3) were administered to jugular cannulated rats together with corn oil to confirm whether the suppression of plasma TG levels actually was expressed in vivo. Fr3 exhibited significant suppression of plasma TG levels after 3 h and 6 to 7 h after administration of corn oil in rats. The AUC level 10 h after administration of Fr3 also was suppressed compared with control levels. The suppressive effect of Fr3 on the plasma TG elevation was stronger than that observed for Fr2. These results were coincident with those obtained in vitro, which was thought to be the strong inhibition of pancreatic lipase activity.

On the other hand, Fr2 also exhibited significant suppression of plasma TG levels after 3 h and 6 to 7 h after administration, although Fr2 showed weak lipase inhibition compared to that of Fr3. These results suggest Fr2 suppressed elevation of plasma TG levels by different pathways from pancreatic lipase inhibition. Inhibition of the constitution of the emulsion is another candidate mechanism for the Fr2-mediated suppression of the plasma TG levels. Numerous studies have demonstrated natural polyphenols inhibit the elevation of serum cholesterol levels in mammals (19–21). We have previously demonstrated that BBJ significantly decreased cholesterol levels in rat livers after long-term feeding of a high-fat diet, similar to other polyphenols (13). Ikeda et al. (22) reported that tea catechins reduced the solubility of cholesterol in mixed micelles, leading to lower serum cholesterol levels. Thus, Fr2 could have the potential to
inhibit lipid absorption from the gastrointestinal tract by inhibition of the constitution of the emulsion through binding of BBJ polyphenols to cholic acid, similar to the case of tea catechins. Further studies on the binding abilities of BBJ polyphenols to cholic acid should be examined.

Certain polyphenol components were thought to express their anti-obesity effect by modulating various factors related to lipid accumulation after being absorbed into adipose tissues. Tea catechins have been shown to increase acyl-CoA oxidase (5, 23) and (−)-epigallocatechin-3-gallate-inhibited lipid accumulation in 3T3-L1 cells (24). Anthocyanins in blueberry also ameliorated serum leptin levels (2,5) and authentic cyanidin 3-O-glucoside suppressed lipogenesis components at the mRNA level (26). In our previous work, BBJ significantly decreased active FAS activity in the liver (13) after long-term feeding of a high-fat diet containing BBJ in rats, although the same results were also obtained in rats consuming a normal diet containing BBJ. Therefore, we inferred that the polyphenols such as the anthocyanins contained in Fr2 may modulate adipocytes and enzymes promoting lipogenesis after absorption of these polyphenols, although further studies are required to clarify these points.

In conclusion, the anti-obesity effects of BBJ were, in part, the result of various polyphenolic components in BBJ through the inhibition of pancreatic lipase activity in the small intestine leading to the result of suppression of triglyceride absorption from the gastrointestinal tract.

Acknowledgments
The authors gratefully acknowledge Mr. Taichi Kazama for his advice with the HPLC-UV-MS/MS analysis.

REFERENCES
21) Prior RL, Wu X, Gu L, Hager T, Hager A, Wilkes S, Howard L. 2009. Purified berry anthocyanins but not whole berries normalize lipid parameters in mice fed an obeso-


