

## Nutrients and Circadian Rhythms in Mammals

Tao WU, Cencen YAO, Liangfeng HUANG, Youxiang MAO, Wanjing ZHANG,  
Jianguo JIANG, Zhengwei FU\*

College of Biological and Environmental Engineering, Zhejiang University of Technology,  
No.6 District, Zhaohui, Hangzhou, Zhejiang, 310032, China

**Summary** The circadian rhythm is generally existed in mammalian behavior and metabolic processes, which results from the self-sustained circadian clocks. The mammalian circadian clocks are composed of a master clock located in the hypothalamic suprachiasmatic nucleus (SCN), and of many peripheral clocks in tissues and extra-SCN brain regions. It is indicated that feeding could take over part of the SCN signaling, and affect internal synchrony between the master clock and the peripheral clocks. Thus, recent studies focus more on the relationship between the nutrients and circadian rhythms. Various nutrient components (glucose, amino acid, alcohol) are found to be able to directly affect the circadian rhythm of clock genes. Moreover, the feeding schedule of nutrients is as important as the nutrient components in maintaining a healthy circadian rhythm. Therefore, the circadian homeostasis needs not only balanced nutrient components but also regular timed nutrients.

**Key Words** circadian rhythm, clock, nutrient components, feeding schedule

### The Biological Clock and Circadian Rhythm

In mammals, the approximate 24-h rhythm in behavior and physiology is generally existed, such as wake/sleep cycle, blood pressure, body temperature and concentration of melatonin. The circadian rhythms result from the cell-autonomous and self-sustained oscillators (circadian clocks), which rely on interlocking transcription/translational feedback loops involving a series of clock genes (*Clock*, *Bmal1*, *Per*, *Cry*, *Dec* etc.) and their proteins (1). The mammalian circadian clocks are composed of a master clock located in the hypothalamic suprachiasmatic nucleus (SCN), and of many peripheral clocks in tissues and extra-SCN brain regions (2). The master clock is mainly entrained by the light cue, while the peripheral clocks are not only affected by the SCN clock but also entrained by the food cue (3). Moreover, feeding could take over part of the SCN signaling, and affect internal synchrony between the master clock and the peripheral clocks.

### Nutrient Components and Circadian Rhythms

Nutrient components have been identified as important factors to entrain the peripheral circadian clocks (4).

Glucose is a particularly potent entraining factor for peripheral clocks (5). Min-Dian Li et al. found that high concentration of glucose increased the transcription levels of *Bmal1* and *Cry1* genes without the alteration of the phase of *Bmal1* cycling, and low concentration of glucose delayed the phase of BMAL1 protein accumulation via western blot (6). How could glucose influence the circadian clock and further affect the following metabolic process aroused extensive attention. Some

researchers revealed that cellular nutrient sensors such as nuclear receptors were proposed as candidates for the circadian clock entrainment by nutrient components (7, 8). A recent study showed that the O-GlcNAc signaling entrained the circadian clock by inhibiting BMAL1/CLOCK ubiquitination, which may be the molecular mechanism underlying the glucose entrainment of the circadian clock (6).

Intraperitoneal injection of amino acids combined with glucose delayed the phase of the liver clock as similar as the effect of delayed feeding (9). L-carnosine is a dipeptide of the amino acids  $\beta$ -alanine and L-histidine, which is identified to be related with various physiological alterations (including blood glucose, blood pressure etc.) through the autonomic nerves. Bilateral lesions of the SCN in rat results in destructive effects on the carnosine induced physiological alterations mentioned above (10). In our study, we found that L-carnosine administration could accelerate the resetting rate of peripheral clock genes in the hearts, which is regulated by the autonomic nervous system (11).

Alcohol is another entraining factor for circadian clocks. Chronic alcohol administration induced the hepatic steatosis and disturbed the circadian clocks in the liver (12). Studies in human indicated that the expression level of clock genes in leukocytes of male alcoholic patients were lower than the healthy men (13).

### Timed Nutrients and Circadian Rhythms

Timed nutrients are as important as nutrient components themselves for maintaining normal circadian rhythms. More and more evidences show that feeding schedules play entraining roles in peripheral clocks and behavior rhythms, which is independent of the SCN signaling (14). Arble DM et al. focused on the role of the timed nutrients in weight gain, and found that mice fed

\*Corresponding author.  
E-mail: azwfu@zjut.edu.cn

a high-fat diet only during the 12-h light phase gained significantly more weight than those fed only during the 12-h dark phase (15). Recent research showed that feeding time controlled the diurnal rhythm of relative abundance of intestinal microbiota (16), which was crucial for the life span (17). More studies paid attention to why timed nutrients could significantly affect the circadian system and physiological function. Some studies showed that changes in behavior and physiological functions, such as locomotor activity, body temperature, blood hormones occurred before the anticipatory timed nutrients, were established by restricted daily feeding schedules (18). Olivo D et al. found that the anticipatory metabolic activity entrained by feeding schedules existed in piriform cortex and olfactory tubercle, but not in suprachiasmatic nucleus (19). Timed nutrients shifted the circadian phases of clock genes at the mRNA and protein levels in the gastrointestinal tract but not in the central SCN clock (20). Therefore, irregular timed nutrients might induce the alteration of circadian rhythms in the peripheral tissues independent of the SCN, resulting in the uncoupling between the SCN and peripheral clocks, which may further affect the downstream metabolic processes.

Moreover, three meals a day is a well-established human feeding habit, and the breakfast is always regarded as one of the most important meals of the day. In our study, we found that the first meal of the daily cycle activated the transcription of peripheral clock genes and determined their phases, while the daily last meal was tightly related to lipid metabolism and adipose tissue accumulation (21).

### The Connection between Nutrients and Circadian Rhythms

Studies mentioned above review the relationship between the nutrients and circadian rhythms. Then, how are the circadian rhythms entrained by various nutrients? Studies on the nutrient sensors identified several crucial metabolic factors being able to integrate the nutrient signals with the circadian clock. SIRT1 is a member of the sirtuin family, which plays an important role in regulating many proteins involved in response to nutrients (22). In the liver, the rhythmic activity of SIRT1 lies on the level of  $NAD^+$ , which is controlled by the BMAL1/CLOCK heterodimer (23). In return, SIRT1 regulates the circadian clock via the deacetylation of BMAL1 and PER2 proteins then further influencing the clock function (24). Therefore,  $NAD^+$ -dependent SIRT1 functions as a sensor linking the nutrients and the circadian clocks. Moreover, REV-ERB $\alpha$  is a transcription factor regulating adipogenesis, which can inhibit the transcription of *Bmal1* (25). ROR $\alpha$  is a nuclear receptor involved in both lipogenesis and lipid storage, which can activate *Bmal1* transcription (26). Glucocorticoids (GC) and its receptor (GR) have also been identified as an important link between circadian clocks and nutrients (27). Studies have shown that liver-specific GR mutation display accelerated phase shifting in response to restricted feeding schedule in daytime (28), while injec-

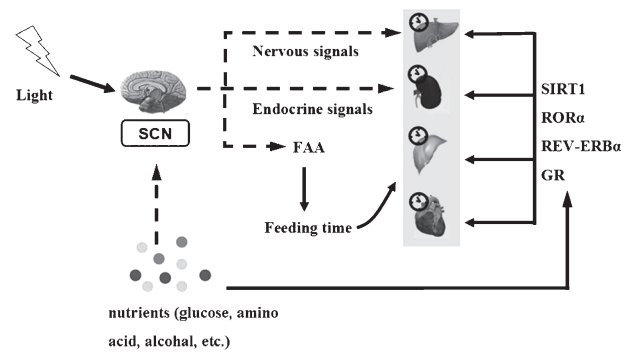


Fig. 1. Peripheral clocks are entrained by nutrients and SCN-derived resetting cues. SCN clock transmits messages to peripheral clocks via nervous and endocrine signals. Nutrients could take over part of the SCN signaling to affect the peripheral clocks via some nutrient sensors such as SIRT1, REV-ERB $\alpha$ , ROR $\alpha$ , GR. Nutrients information also goes to the hypothalamus, then produces FAA before feeding time.

tion of corticosterone do not cause the phase shifting induced by restricted feeding (29).

### Concluding Remarks

Various nutrient components, timed nutrients, the time interval between meals, and nutrients amount all can entrain the circadian clocks, which may further affect the downstream metabolic processes. Life-long calorie-restricted diet could extend the life span in mice (17). Eating at the active phase reduces weight gain compared to that at the rest phase (15). Time-restricted feeding (TRF) can prevent nutritional challenges and has the protective effect on maintaining relative physiological homeostasis, which is hard to be interrupted by occasional irregular eating (30). Therefore, the circadian homeostasis needs not only balanced nutrient components but also regular timed nutrients.

### Acknowledgments

This work was funded by the Natural Science Foundation of Zhejiang Province, China (No. Y3090563), the National Natural Science Foundation of China (No. 31200890), and Cerebos Pacific Ltd, Singapore.

### REFERENCES

- 1) Reppert SM, Weaver DR. 2002. Coordination of circadian timing in mammals. *Nature* **418**: 935–941.
- 2) Schibler U, Sassone-Corsi P. 2002. A web of circadian pacemakers. *Cell* **111**: 919–922.
- 3) Mendoza J, Graff C, Dardente H, Pevet P, Challet E. 2005. Feeding cues alter clock gene oscillations and photic responses in the suprachiasmatic nuclei of mice exposed to a light/dark cycle. *J Neurosci* **25**: 1514–1522.
- 4) Froy O. 2007. The relationship between nutrition and circadian rhythms in mammals. *Front Neuroendocrinol* **28**: 61–71.
- 5) Stephan FK, Davidson AJ. 1998. Glucose, but not fat, phase shifts the feeding-entrained circadian clock. *Physiol Behav* **65**: 277–288.
- 6) Li MD, Ruan HB, Hughes ME, Lee JS, Singh JP, Jones

- SP, Nitabach MN, Yang X. 2013. O-glcnaC signaling entrains the circadian clock by inhibiting bmal1/clock ubiquitination. *Cell Metab* **17**: 303–310.
- 7) Yang X, Downes M, Yu RT, Bookout AL, He W, Straume M, Mangelsdorf DJ, Evans RM. 2006. Nuclear receptor expression links the circadian clock to metabolism. *Cell* **126**: 801–810.
- 8) Yang X. 2010. A wheel of time: The circadian clock, nuclear receptors, and physiology. *Genes Dev* **24**: 741–747.
- 9) Oike H, Nagai K, Fukushima T, Ishida N, Kobori M. 2011. Feeding cues and injected nutrients induce acute expression of multiple clock genes in the mouse liver. *PLoS One* **6**: e23709.
- 10) Nagai K, Tanida M, Nijima A, Tsuruoka N, Kiso Y, Horii Y, Shen J, Okumura N. 2012. Role of l-carnosine in the control of blood glucose, blood pressure, thermogenesis, and lipolysis by autonomic nerves in rats: Involvement of the circadian clock and histamine. *Amino Acids* **43**: 97–109.
- 11) Wu T, Tao Y, Tsang F, Abe K, Xu L, Jiang Q, Xu L, Fu H, Fu Z. 2014. The effect of L-carnosine on the circadian resetting of clock genes in the heart of rats. *Mol Biol Rep* DOI: 10.1007/s11033-014-3745-x.2013.
- 12) Zhou P, Ross RA, Pywell CM, Liangpunsakul S, Duffield GE. 2014. Disturbances in the murine hepatic circadian clock in alcohol-induced hepatic steatosis. *Sci Rep* **4**: 3725.
- 13) Huang MC, Ho CW, Chen C-H, Liu SC, Chen CC, Leu SJ. 2010. Reduced expression of circadian clock genes in male alcoholic patients. *Alco-Clin Exp Res* **34**: 1899–1904.
- 14) Martinez TM, Parra-Gamez LG, Aguilar RR, Escobar CB. 2001. Leptin rhythmicity is entrained to feeding schedules in rats. *Soc Neurosci Abstr* **27**: 486–486.
- 15) Arble DM, Bass J, Laposky AD, Vitaterna MH, Turek FW. 2009. Circadian timing of food intake contributes to weight gain. *Obesity* **17**: 2100–2102.
- 16) Thaiss CA, Zeevi D, Levy M, Zilberman-Schapira G, Suez J, Tengeler AC, Abramson L, Katz MN, Korem T, Zmora N, Kuperman Y, Biton I, Gilad S, Harmelin A, Shapiro H, Halpern Z, Segal E, Elinav E. 2014. Transkingdom control of microbiota diurnal oscillations promotes metabolic homeostasis. *Cell* **159**: 514–529.
- 17) Zhang C, Li S, Yang L, Huang P, Li W, Wang S, Zhao G, Zhang M, Pang X, Yan Z, Liu Y, Zhao L. 2013. Structural modulation of gut microbiota in life-long calorie-restricted mice. *Nat Commun* **4**: 2163.
- 18) Feillet CA, Ripperger JA, Magnone MC, Dulloo A, Albrecht U, Challet E. 2006. Lack of food anticipation in *per2* mutant mice. *Curr Biol* **16**: 2016–2022.
- 19) Olivo D, Caba M, Gonzalez-Lima F, Vazquez A, Corona-Morales A. 2014. Circadian feeding entrains anticipatory metabolic activity in piriform cortex and olfactory tubercle, but not in suprachiasmatic nucleus. *Brain Res* **1592**: 11–21.
- 20) Hoogerwerf WA, Hellmich HL, Cornelissen G, Halberg F, Shahinian VB, Bostwick J, Savidge TC, Cassone VM. 2007. Clock gene expression in the murine gastrointestinal tract: Endogenous rhythmicity and effects of a feeding regimen. *Gastroenterology* **133**: 1250–1260.
- 21) Wu T, Sun L, ZhuGe F, Guo X, Zhao Z, Tang R, Chen Q, Chen L, Kato H, Fu Z. 2011. Differential roles of breakfast and supper in rats of a daily three-meal schedule upon circadian regulation and physiology. *Chronobiol Int* **28**: 890–903.
- 22) Rodgers JT, Lerin C, Haas W, Gygi SP, Spiegelman BM, Puigserver P. 2005. Nutrient control of glucose homeostasis through a complex of *pgc-1*  $\alpha$  and *sirt1*. *Nature* **434**: 113–118.
- 23) Nakahata Y, Sahar S, Astarita G, Kaluzova M, Sassone-Corsi P. 2009. Circadian control of the nad(+) salvage pathway by clock-sirt1. *Science* (80-) **324**: 654–657.
- 24) Asher G, Gattfield D, Stratmann M, Reinke H, Dibner C, Kreppel F, Mostoslavsky R, Alt FW, Schibler U. 2008. Sirt1 regulates circadian clock gene expression through *per2* deacetylation. *Cell* **134**: 317–328.
- 25) Preitner N, Damiola F, Molina LL, Zakany J, Duboule D, Albrecht U, Schibler U. 2002. The orphan nuclear receptor *rev-erb*  $\alpha$  controls circadian transcription within the positive limb of the mammalian circadian oscillator. *Cell* **110**: 251–260.
- 26) Sato TK, Panda S, Miraglia LJ, Reyes TM, Rudic RD, McNamara P, Naik KA, Fitzgerald GA, Kay SA, Hogenesch JB. 2004. A functional genomics strategy reveals *rora* as a component of the mammalian circadian clock. *Neuron* **43**: 527–537.
- 27) Dickmeis T, Foulkes NS. 2011. Glucocorticoids and circadian clock control of cell proliferation: At the interface between three dynamic systems. *Mol Cell Endocrinol* **331**: 11–22.
- 28) Le Minh N, Damiola F, Tronche F, Schutz G, Schibler U. 2001. Glucocorticoid hormones inhibit food-induced phase-shifting of peripheral circadian oscillators. *Embo Journal* **20**: 7128–7136.
- 29) Stokkan KA, Yamazaki S, Tei H, Sakaki Y, Menaker M. 2001. Entrainment of the circadian clock in the liver by feeding. *Science* (80-) **291**: 490–493.
- 30) Chaix A, Zarrinpar A, Miu P, Panda S. 2014. Time-restricted feeding is a preventative and therapeutic intervention against diverse nutritional challenges. *Cell Metab* **20**: 991–1005.