Impact of Purslane (*Portulaca oleracea* L.) Leaves Extract to Enhance the Anti-oxidant Potential of Edible Oils during Heating

Niharika Shanker and Sukumar Debnath*

Department of Technology Scale-up CSIR-Central Food Technological Research Institute, Mysore-570 020, INDIA

Abstract: The present work deals/ reveals with the effect of purslane leaves extract on the stability of soybean oil during heating and the acceptability of oil after preparation of *poori* (an Indian fried bread) by frying and its sensory evaluation. The ethanolic purslane leaves extract was blended with soybean oil at three different concentrations such as, 500, 1000 and 1500 ppm (T1, T2 and T3) and compared with control. The sample added with 100 ppm TBHQ was used as a positive control. Assessment of antioxidant activity of the ethanolic extract of purslane leaves was carried out by the estimation of total phenolic content, loss of β-carotene and antioxidant activity. The heating (173±2°C for 24 h; 8 h heating cycles per day) performance of soybean oil incorporated with purslane leaves extract was evaluated in terms of peroxide value, free fatty acid, total polar material and fatty acid composition. The thermal stability of the oils was evaluated using differential scanning calorimeter. The *poori* was prepared to check the acceptability of the oil. Results suggest that leaves extract of purslane (1500 ppm) may be used for obtaining reasonable thermal stability of soybean oil with acceptable sensory characteristics of the product. Although TBHQ showed almost similar thermal stability with leaves extract of purslane (1500 ppm), natural anti-oxidant is more preferred over synthetic anti-oxidant.

Key words: purslane leaf extract, TBHQ, anti-oxidant potential, edible oils

1 Introduction

Recently the interest in use of natural antioxidant has gained momentum as that natural antioxidants are safer and superior than processed / artificial ones. Many sources of natural antioxidants are present such as spices, herbs, tea leaves, oilseeds, cereals, cocoa beans, grains, fruits, vegetables, enzymes, proteins etc. Purslane leaf is a good plant source of nutritional benefits with high antioxidant properties. It is one of the richest green plant sources of omega-3 fatty acids. Purslane leaves function as a potent antioxidant in lipid systems. The oxidative reaction is responsible for rancid odors and flavors of fats and oils which reduce the nutritional quality of the foods. Oxidation reactions consist of auto-oxidation, photo-oxidation, enzymatic oxidation and ketonic oxidation, whereas auto-oxidation, in general, is the deterioration of quality during storage of edible oils. Auto-oxidation is the reaction between oxygen and unsaturated fatty acids *via* an auto-catalytic procedure consists of a free radical chain mechanism. This chain includes beginning, propagation, and termination reactions that could be cyclical once started:

Initiation: \[ \text{RH} \rightarrow \text{R}^* \]

Propagation \[ \text{R}^* + \text{O}_2 \rightarrow \text{ROO}^* \]

\[ \text{ROO}^* + \text{RH} \rightarrow \text{ROOH} + \text{R}^* \]

\[ \text{R}^* + \text{R}^* \rightarrow \text{R} - \text{R} \]

Termination \[ \text{ROO}^* + \text{R}^* \rightarrow \text{ROOR} \]

\[ \text{ROO}^* + \text{ROO}^* \rightarrow \text{Non - radical products} \]

In the beginning, free radicals are formed from the substrate. Antioxidants are components which stop auto-oxidation of oils and fats by giving their hydrogen to free radicals formed in the beginning and formulate the stages of auto-oxidation by following reactions (AH is antioxidant molecule):
ROO• + AH $\rightarrow$ ROOH + A•
R• + AH $\rightarrow$ RH + A•
RO• + AH $\rightarrow$ ROH + A•
ROO• + A• $\rightarrow$ ROOA
RO• + A• $\rightarrow$ ROA
A• + A• $\rightarrow$ A – A
(Source: Young and Woodside$^2$)

There are several variables which affect the auto-oxidation reaction such as unsaturation, temperature, the presence of oxygen, light, moisture, heavy metals and antioxidants. Antioxidants prevent free radical-induced cell and biological targets damage by preventing the formation of radicals, scavenging them, or by promoting their decomposition$^2$. Usually, they can be classified as two main groups, natural and synthetic antioxidants. Among the synthetic types, the most frequently used are propyl gallate (PG) and tert-butyl hydroquinone (TBHQ). In most countries, the usage level of synthetic antioxidants is regulated and the safety of toxicity studies. Mainly the interest on naturally occurring antioxidants is developed because of the trend to minimize or avoid the use of synthetic food additives$^3$.

In recent years, phenolic compounds of plant origin have attracted considerable attention due to their beneficial functional and nutritional effects including antioxidant and antimicrobial activity$^4$. The literature on the use of natural plant extracts with antioxidant activities in various edible oils is scanty. This work has been done to reduce the application of synthetic compounds as antioxidants because of their harmful health effects. The present study concerns about the stabilization of oil with natural antioxidants during heating / frying. The calorimetric analysis is generally used to check the thermal stability of edible oils as thermal decomposition depends upon fatty acids composition and presence of antioxidants. The report on the antioxidant potential of purslane leaves extract for the stabilization of soybean oil during heating is scanty.

Poori is an Indian fried bread that is traditionally used during breakfast. It is made by deep frying of circular sheet prepared from the dough of wheat flour. Refined oil is used for the preparation of poori. The purpose of this research work was to explore the stability of antioxidant potential of purslane leaves extract towards heating stability of edible oils and sensory acceptability of the product (poori: an Indian fried bread) during frying.

2 Materials and Methods

2.1 Materials

Fresh leaves of purslane were procured from local market of Mysore. Refined soybean oil with no added antioxidant was procured from local market of Mysore, and was kept at $-20^\circC$ until analysis. All the chemicals and reagents procured from Sigma Aldrich, Bangalore, were of analytical grade. The solvents procured from Merck & Co. (Darmstadt, Germany), were of analytical grade. All the experiments were done in triplicates.

2.2 Preparation of leaf extract of purslane (LEP)

The purslane leaves were dried under the shed at 30 ± 2°C for 5 days. During solvent extraction method sample (dried purslane leaf, 10g) was mixed with 100 mL of solvent (Ethanol - water, 1:1, v/v). The solution was blended thoroughly in a shaker at 160 rpm at room temperature for 48 h. After extraction, the leaves extracts were filtered. The solvents were allowed to evaporate using a rotary evaporator (BUCHI, R-215, Switzerland) at 50°C. The concentrated leaf extract was added into soybean oil (SBO) at three different concentrations; 500 ppm, 1000 ppm and 1500 ppm (T1, T2, and T3). These samples were compared with control (blank; with no addition of extract) and the positive control sample (Tertiary butylhydroquinone, TBHQ, 100 ppm). The samples filled in glass bottles were kept at room temperature for further analysis.

2.3 Analysis of natural antioxidants

The total phenolic content (TPC) of purslane leaf extract was expressed in terms of gallic acid by the method followed by Anwar et al.$^5$ The β-carotene was estimated by the method followed by Taga et al.$^6$ In a flask 40 mg of linoleic acid and 400 mg of Tween 40 were added, about two milliliters of a solution of β-carotene in chloroform (1 mg/mL) was pipetted into it. Chloroform was evaporated using flash evaporator at 45°C for 4 min. Distilled water (100 mL) was added gradually to the semisolid residue with vigorous shaking to form an emulsion. About 5-mL aliquot of the emulsion was added to a tube containing 0.2 mL of the antioxidant solution (500 mg/L). The absorbance of the same was recorded at 470 nm against the blank containing mixture without β-carotene. Tubes were placed in a water bath at 50 ± 2°C, and the absorbance measurements were conducted again at 15-min intervals up to 120 min. The 1-diphenyl-2-picrylhydrazyl radical (DPPH) was used for the determination of free radical-scavenging activity of the extracts$^7$. Different concentrations of extract were added to an equal volume of ethanolic DPPH (100 μM) solution. Absorbance was taken at 517 nm at room temperature after 15 min.
2.4 Determination of free fatty acids, peroxide value and total polar material

The determination of free fatty acids (FFA) and peroxide value (PV) for oxidative stability were carried out by the AOCS\textsuperscript{10}. The thermo-oxidative degradation level of the oil was measured by the changes in total polar material (TPM). The TPM present in oils was measured by instrumental method (Fri-Check, bvba, Belgium) based on changes in viscosity of oil during heating\textsuperscript{8,10}.

2.5 Determination of fatty acids composition

Fatty acid methyl esters (FAMEs) were prepared based on standard IUPAC method\textsuperscript{11} and analysed using gas chromatography (Shimadzu, 2010) equipped with FID using fused silica Rtx-2330 column (Restek made, 30 m $\times$ 0.32 mm ID and 0.20 $\mu$m film thickness). Injector port and detector temperature were set up at 230 and 250$^\circ$C, respectively. The N$_2$ was used as a carrier gas. Initially, column temperature was maintained at 120$^\circ$C, followed by increasing to 220$^\circ$C in 20 min, and holding the same for 10 min. The fatty acids were identified by comparing their fragmentation pattern and retention time with authentic standards.

2.6 Heating studies of oils

The samples (50 g) of each variation were heated continuously in 100 mL beakers on a hot plate at 173 $\pm$ 2$^\circ$C for 24 h (8 h heating cycles per day)\textsuperscript{12}. The temperature was monitored using electronic temperature indicator (HTA, Bangalore). Samples of oil with extract and pure oil were taken for every 8 h heating cycle and kept in a refrigerator after cooling at room temperature for further analyses.

2.7 Product formulation: preparation of poori

The product is made by kneading the wheat flour (1 kg) with lukewarm water (650 ml) in a Hobart mixer (Hobart, England) until the soft, non-sticky dough is formed, preparing equal sized (0.02 kg) balls from the dough, shaping the raw poori (av. dia. 0.08 m, thickness 0.003 m) using rolling board with rolling pin and frying the same. The frying was carried out at atmospheric pressure at 173 $\pm$ 2$^\circ$C for 5 min using laboratory scale deep-fat fryer (Mini Master Fryer, Continental, India)\textsuperscript{3} with 2 l oil. The temperature of the oil was measured by T-type thermocouple.

2.8 Determination of melting temperature and enthalpy

The melting temperature of the native oil and oils with TPM present in oils was measured by instrumental method (Fri-Check, bvba, Belgium) based on changes in viscosity of oil during heating\textsuperscript{8,10}.

2.9 Sensory evaluation of the product

Sensory evaluation of the product (poori) was carried out at our laboratory consists of booth room maintained at 22 $\pm$ 2$^\circ$C under fluorescent light. The samples of poori were randomly served to a panel of ten trained judges (coded with random number) who are regular consumer of similar food products. All the orders of servings were fully randomized. The prescribed method\textsuperscript{14} of nine-point hedonic scale with sensory attributes such as appearance, taste, color, after taste, overall acceptance was followed for the sensory evaluation. Assessment of preferences coded with numerical values as extremely like (9), rather like (8), quite like (7), neither like nor dislike (6), dislike a little (5), rather dislike (4), dislike very much (3), highly dislike (2) and extremely dislike (1), was used for sensory evaluation.

2.10 Statistical analysis

All investigations were done in triplicates. The outcome was represented as mean $\pm$ standard deviation. Data were statistically analyzed by ANOVA for comparison of means using Microsoft Excel 10 for significance ($p<0.05$).

3 Results and Discussion

3.1 Accelerated oxidation studies

The phenolic compounds are considered to be the most important phytochemicals responsible for antioxidant potential of plant materials. The purslane is a great source of ascorbic acid and flavonoids which also have the antioxidant activity. The polyphenolic compounds are broadly spread in different parts of the plants\textsuperscript{50}. The study demonstrated that in many plant species, the total phenolic content and antioxidant activity are positively correlated with each other\textsuperscript{51}.

The Table 1 showed the ability of purslane leaf extracts to scavenge DPPH radical at concentrations of 500 to 1000 ppm and 1500 ppm are. The result showed that scavenging-free radicals also increased till 1500 ppm (41.6% to 87.1%) which is due to an increase in concentration of total polyphenolic compounds (85.3 mg GAE/g to 151.7 mg GAE/g) (Table 1) in the extracts\textsuperscript{17}. As the concentrations of phenolic compounds increases, the amount of hydroxyl groups present in the reaction medium is also increased. Consequently, chances of hydrogen donation to free radicals are increased\textsuperscript{49}. The outcome of the work revealed that LEP at maximum concentration of 1500 ppm had the acceptable range.

The antioxidant activities of the purslane leaf extracts, measured by bleaching of $\beta$-carotene are indicated in Table 1. It is found that extract of purslane leaf at 500, 1000 and 1500 ppm had the significant antioxidant activities. Therefore, LEP with different ratio was used to assess its effect on the stability of soybean oil during the heating process.
the changes in the fatty acid composition of oils due to observe the effect of various heating conditions on fatty acids profile. The fatty acid profile changes for all the individual fatty acids of soybean oil and effect of heating on fatty acids with two or three double bonds after the heat treatment is given to oils and fats. In the present study, the level of polyunsaturated fatty acid values (PUFA) was found to decrease the level of saturated fatty acids (SFA) was found to increase during heating. The estimated concentration of the unsaturated fatty acids showed that oil with TBHQ and with LEP had a good unsaturation level as compared to control oil. After 16 h of heating, unsaturation level was found to decrease more in all the oils in comparison to samples with added synthetic antioxidants. After 24 h the level of PUFA in control was found to significantly ($p<0.05$) decrease from 53.1% to 33.8%, in positive control 54.2% to 42.6%, but less degraded from 53.1% to 40.4% in oil with the addition of LEP (1500 PPM, T3). Similarly, the MUFA content was found to decrease in control 36.7% to 28.4%, positive control 36.4% to 33.2% and in T3 38.7% to 36.6%. At the same time there is an increase in SFA in all the samples. After 24 h the SFA was observed in control 8.9% to 28.3%, positive control 8.8% to 19.5% and T3 8.5% to 19.6%.

3.2 Effect of purslane leaves extract on fatty acid composition

Gas chromatography (GC) is a helpful methodical procedure to observe the effect of various heating conditions on the changes in the fatty acid composition of oils. The Table 2(a, b) indicated the fatty acids composition of individual fatty acids of soybean oil and effect of heating on fatty acids profile. The fatty acid profile changes for all the oils with control, positive control with TBHQ and T3 (based on acceptability range). Many researchers found that the changes occur in fatty acids with two or three double bonds after the heat treatment is given to oils and fats. In the present study, the level of polyunsaturated fatty acid values (PUFA) was found to decrease the level of saturated fatty acids (SFA) was found to increase during heating. The estimated concentration of the unsaturated fatty acids showed that oil with TBHQ and with LEP had a good unsaturation level as compared to control oil. After 16 h of heating, unsaturation level was found to decrease more in all the oils in comparison to samples with added synthetic antioxidants. After 24 h the level of PUFA in control was found to significantly ($p<0.05$) decrease from 53.1% to 33.8%, in positive control 54.2% to 42.6%, but less degraded from 53.1% to 40.4% in oil with the addition of LEP (1500 PPM, T3). Similarly, the MUFA content was found to decrease in control 36.7% to 28.4%, positive control 36.4% to 33.2% and in T3 38.7% to 36.6%. At the same time there is an increase in SFA in all the samples. After 24 h the SFA was observed in control 8.9% to 28.3%, positive control 8.8% to 19.5% and T3 8.5% to 19.6%.

Table 1  Antiradical activities, total phenol content and β-carotene assays of purslane leaf extracts.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>T1</th>
<th>T2</th>
<th>T3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Antiradical Activity (%)</td>
<td>38.4 ± 0.1d</td>
<td>41.6 ± 0.4d</td>
<td>70.8 ± 0.9c</td>
<td>87.1 ± 0.6d</td>
</tr>
<tr>
<td>TPC Content (mg GAE/g)</td>
<td>85.3 ± 0.4c</td>
<td>91.3 ± 0.3b</td>
<td>134.04 ± 0.2c</td>
<td>151.7 ± 0.2d</td>
</tr>
<tr>
<td>β- Carotene (μmol/L)</td>
<td>190 ± 0.5a</td>
<td>260 ± 1.1b</td>
<td>320 ± 0.8c</td>
<td>460 ± 0.9d</td>
</tr>
</tbody>
</table>

where, T1, T2, T3: addition of natural antioxidant LEP at 500 ppm, 1000 ppm and 1500 ppm, respectively. Different letters within the same row in the respective tables indicate significant differences ($p < 0.05$). LEP-Leaf extract of purslane, SFA-saturated fatty acids, MUFA-monounsaturated fatty acids, PUFA-polyunsaturated fatty acids.

Table 2 (a)  Fatty acid profile of individual fatty acids of soybean oil.

<table>
<thead>
<tr>
<th>Fatty acids</th>
<th>(%) relative area</th>
</tr>
</thead>
<tbody>
<tr>
<td>Myristic acid</td>
<td>0.5 ± 0.1</td>
</tr>
<tr>
<td>Palmitic acid</td>
<td>5.2 ± 0.2</td>
</tr>
<tr>
<td>Stearic acid</td>
<td>3.2 ± 0.1</td>
</tr>
<tr>
<td>Oleic acid</td>
<td>36.7 ± 0.7</td>
</tr>
<tr>
<td>Linoleic acid</td>
<td>46.0 ± 0.4</td>
</tr>
<tr>
<td>Linolenic acid</td>
<td>7.1 ± 0.6</td>
</tr>
</tbody>
</table>

Table 2 (b)  Fatty acid profile of SFA, MUFA and PUFA of fresh and heated oils.

<table>
<thead>
<tr>
<th>Duration of heating (h)</th>
<th>Control</th>
<th>Positive Control</th>
<th>T3</th>
<th>Control</th>
<th>Positive Control</th>
<th>T3</th>
<th>Control</th>
<th>Positive Control</th>
<th>T3</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>SFA</td>
<td>MUFA</td>
<td>PUFA</td>
<td></td>
<td></td>
<td></td>
<td>SFA</td>
<td>MUFA</td>
<td>PUFA</td>
</tr>
<tr>
<td>0</td>
<td>8.9 ± 0.2a</td>
<td>8.8 ± 0.1a</td>
<td>8.5 ± 0.3a</td>
<td>36.7 ± 0.1b</td>
<td>36.4 ± 0.2c</td>
<td>36.6 ± 0.1b</td>
<td>53.1 ± 0.2a</td>
<td>54.2 ± 0.1e</td>
<td>53.1 ± 0.1c</td>
</tr>
<tr>
<td>8</td>
<td>14.8 ± 0.3d</td>
<td>12.1 ± 0.1d</td>
<td>18.7 ± 0.2c</td>
<td>34.1 ± 0.5d</td>
<td>35.2 ± 0.1f</td>
<td>34.5 ± 0.2c</td>
<td>48.7 ± 0.3c</td>
<td>51.8 ± 0.2f</td>
<td>50.8 ± 0.1d</td>
</tr>
<tr>
<td>16</td>
<td>18.6 ± 0.1f</td>
<td>14.0 ± 0.2g</td>
<td>15.3 ± 0.2e</td>
<td>30.6 ± 0.1e</td>
<td>35.6 ± 0.1h</td>
<td>37.0 ± 0.3g</td>
<td>41.3 ± 0.1g</td>
<td>45.9 ± 0.3c</td>
<td>44.4 ± 0.1i</td>
</tr>
<tr>
<td>24</td>
<td>28.3 ± 0.1e</td>
<td>19.5 ± 0.1h</td>
<td>19.6 ± 0.1n</td>
<td>28.4 ± 0.3i</td>
<td>33.3 ± 0.1i</td>
<td>38.7 ± 0.1l</td>
<td>33.8 ± 0.2j</td>
<td>42.6 ± 0.1l</td>
<td>40.4 ± 0.2k</td>
</tr>
</tbody>
</table>

where, control: without adding any antioxidant; positive control: addition of synthetic anti-oxidant (TBHQ, 100ppm), T3: addition of natural antioxidant LEP at 1500 ppm. Different letters within the same row in the respective Tables (SFA, MUFA, PUFA) indicate significant differences ($p < 0.05$). LEP-Leaf extract of purslane, SFA-saturated fatty acids, MUFA-monounsaturated fatty acids, PUFA-polyunsaturated fatty acids.

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industry as it leads to the development of off-flavor in oils and fried foods. Changes in the FFA content of the oil samples are presented in Fig. 1. The FFA is a marker designed for estimation of oil degradation while heating/frying. The FFA showed an increasing trend during heating period to the end of the experiment in every sample. The total FFA at the end of 24 h of heating for soybean oil containing (a) 500, (b) 1000 and (c) 1500 ppm of LEP, TBHQ, and SBO reached to 6.8, 5.9, 5.2, 4.9, and 7.7, respectively. However, the FFA values of control oil were found to be significantly \( p < 0.05 \) different than oil samples treated with natural (T1, T2, T3) and synthetic (TBHQ) antioxidants.

### 3.4 Effect of purslane leaves extract on peroxide value

Changes in PV of the oil samples during the heating process at 173 ± 2°C are shown in Fig. 2. The PV of control oil is reported to be \( < 2 \) meq \( \text{O}_2/\text{kg} \). It was noticed that the extract is responsible for the significant \( p < 0.05 \) reduction of PV of oils as compared to control oil. Initially, during heating, PV of SBO is observed to rise from 0 to 18 h, for TBHQ, for LEP at 1000 and 1500 ppm from 0 to 12 h and reduced later on. Results of PV with peak value were shown as: TBHQ (1.4 meq \( \text{O}_2/\text{kg} \) after 12 h), LEP at 500 ppm (2.9 meq \( \text{O}_2/\text{kg} \) after 12 h), LEP at 1000 ppm (1.87 meq \( \text{O}_2/\text{kg} \) after 18 h), LEP at 1500 ppm (1.67 meq \( \text{O}_2/\text{kg} \) after 12 h), and SBO (4.1 meq \( \text{O}_2/\text{kg} \) after 12 h). This trend was corroborated with the previous study. The rapid rise in PV of control oil showed that it is more susceptible to oxidation and degradation as well. While oils with 500, 1000, 1500 ppm of LEP and positive control indicated greater ability to prevent an increase of PV as compared to the other oil samples. The only analysis of PV is not a convincing factor to evaluate oils’ oxidative changes while the heating/deep frying method, as peroxidase under frying conditions is unstable and get changed into further compounds such as carbonyl and aldehyde that cause PV abatement.

### 3.5 Effect of purslane leaves extract oil stability

Determination of total polar material (TPM) is one of the main tests and a suitable standard method to examine the thermal oxidative deterioration of the oils during heating/frying. Oil breaks down to generate hydroperoxides, acids and other free radicals, which may further cause polar molecules. This experiment helps to find out the level of polar, oxidized components in a sample. Faster rate of formation of polar components is a cumulative indication of degradation of oil. Figure 3 showed changes in TPM content of the oil samples during the deep frying process. In many European countries, the TPM value is used as a
marker for major oil degradation and it is adequate at maximum of 25-27 for used frying oil. The fresh soybean oil had a TPM content of 5.6, reflecting the good quality of the oil used, as TPM content of unused oils normally ranges between 0.4-6.4. The TPM was found to increase during frying and had high correlation coefficient with heating time. The TPM of the oils containing LEP of 500 and 1000 ppm and 1500 ppm, TBHQ and SBO after 24 h of frying were found to be 24.14, 20.03, 14.5, 13.1, and 28.2, respectively. This may be due to the increase in viscosity during frying/heating. These results showed that SBO reached beyond 25 TPM degradation limit for regulation purposes after 24 h of heating. Therefore, the thermo-oxidative stability of soybean oil was observed to improve considerably in the presence of LEP and synthetic antioxidant as compared to control oil. The study is supported with the earlier work in which stability of the oil was influenced with the antioxidant effect of rosemary extract additives.

3.6 Melting profiles of soybean oils containing TBHQ and LEP

The melting profile of the control oil, positive control oil and the oil enriched with LEP (1500 ppm) after heating for a period of 24 h showed a single melting peak (−9.4°C, ΔH 1.2 J/g; 1.8°C, ΔH 10.3 J/g; and −3.1°C, ΔH −1.7 g, respectively) (Table 3). The reason could be due to the presence of higher levels of unsaturated fatty acids. However, it is observed that the enthalpy (−1.7 J/g) for the oil with LEP was found to decrease in comparison to the enthalpy (1.2 J/g) of control oil. This could be due to the lower degradation of the oil with LEP (1500 ppm) with the formation of lower amount of saturated fatty acids (19.6%) as compared to control oil (28.3%) during heating.

3.7 Sensory evaluation of the product

The results of the sensory evaluation of poori obtained by frying using SBO with different concentrations of LEP as adjudged by a panel of ten trained judges are shown in Fig. 4. Addition of LEP up to T2 (1000 ppm) level did not have any adverse result on taste, smell and color value. However, when the addition level was found to increase up to T3 level (1500 ppm), the sensory scores of these parameters showed no significant (p > 0.05) differences. Some of the panelists have observed for T3 having phenolic smell and taste, which resulted in lower smell and taste scores.

Table 3 Melting Temperature and enthalpy of native oil and oil with LEP.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Onset (℃)</th>
<th>Peak (℃)</th>
<th>End set (℃)</th>
<th>Enthalpy (ΔH, J/g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control oil</td>
<td>−19.2</td>
<td>−9.4</td>
<td>2.5</td>
<td>1.2</td>
</tr>
<tr>
<td>Oil heated upto 24 h with TBHQ</td>
<td>−8.7</td>
<td>1.8</td>
<td>9.1</td>
<td>10.3</td>
</tr>
<tr>
<td>Oil heated upto 24 h with LEP (1500 ppm)</td>
<td>−14.6</td>
<td>−3.1</td>
<td>−1.0</td>
<td>−1.7</td>
</tr>
</tbody>
</table>

where, LEP-Leaf extract of purslane, TBHQ- Tertiary butylhydroquinone
The reason for the low colour score of T3 was due to the dark green color of LEP which has a slight but definite effect on this parameter.

4 Conclusion

Food processing operations require antioxidants that can sustain high temperatures during baking, cooking or frying, and provide protection to finished products. The ethanolic extract of purslane leaves showed an excellent antioxidant activity in soybean oil in a dose-dependent manner during accelerated oxidation during heating. The strong antioxidant activity of LEP in soybean oil was emphasized on the presence of potent anti-oxidative compounds with high thermal stability. Thus, the extract of purslane leaves can be used as an alternative source of natural antioxidants for increasing the stability of oils and oil-containing foods. Therefore, this study showed that the purslane has a good potential to be explored as a source of natural antioxidant.

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References

10) Gertz, C. Chemical and physical parameters as a quality indicator of used frying fat. Sponsored by the German Society for Fat Research, Deutsche Gaselchachta fur Fettwissenschaft (DGF), Hagen-Westfalia, Germany (March 20-21) (2000).
19) Kowalski, R. GC analysis of changes in the fatty acid composition of sunflower and olive oils heated with quercetin, caffeic acid, protocatechueic acid. Acta
Chromatogr. 18, 15-23(2007).
29) Vicente, G.; Martin, D.; Garcia-Risco, M.R.; Fornari, T.; Reglero, G. Supercritical carbon dioxide extraction of antioxidants from rosemary (Rosmarinus officinalis) leaves for use in edible vegetable oils. J. Oleo Sci. 61, 689-697(2012).