Degradation of Phenothrin in Stored Wheat Grains

Kenji NAMBU, Yoshiyuki TAKIMOTO and Junshi MIYAMOTO

Research Department, Pesticides Division, Sumitomo Chemical Co., Ltd.,
Takarazuka, Hyogo 665, Japan

(Received October 11, 1980)

¹⁴C-Labeled (+)-*trans* and (+)-*cis* isomers of phenothrin [3-phenoxybenzyl (±)-*cis*, *trans*-chrysanthemate] were each applied at the concentration of 4 ppm to wheat grains with 11% moisture content, and stored at 15 or 30°C in the dark. Both *trans* and *cis* isomers were decomposed slowly, and 79 and 87% of the applied radiocarbon remained intact in the grains after 12-month storage at 30°C, respectively. Both isomers were majorly metabolized via hydrolysis of ester linkage, oxidation of the benzyl alcohol to the benzoic acid and methylation of the benzoic acid. The joint application with piperonyl butoxide and fenitrothion inhibited the degradation of phenothrin isomers to some extent. The phenothrin isomers and their decomposition products were mainly located at seed coat during 12-month storage, and the residue levels of both isomers in flour and bran were 0.77 and 11.4 ppm, respectively. The phenothrin residues in flour somewhat decreased through the baking process, leaving 0.57 ppm of phenothrin isomers in bread. These findings were compared with those of ¹⁴C-malathion which was rapidly decomposed to desmethylmalathion, malathion-monoacids, -diacid and ¹⁴CO₂, with respective half-lives of 8 and 1.1 months at 15 and 30°C.

INTRODUCTION

Phenothrin [3-phenoxybenzyl (±)-*cis*, *trans*-chrysanthemate] is one of the new synthetic pyrethroids and currently used in household, public health and livestock production because of its outstanding insecticidal activity and its extremely low acute mammalian toxicity.¹⁾ Recently this insecticide proved to have an advantage over pyrethrins and bioresmethrin in cost/efficiency and reliable availability as a potential grain protectant.^{2,3)} This compound combined with piperonyl butoxide and fenitrothion was found to control a broad spectrum of insect pests including organophosphorus-resistant strains.^{2,3)} Although metabolic fates of phenothrin in rats,⁴⁻⁶⁾ plants⁷⁾ and soil⁷⁾ were already established, its fate in stored grains has not been clarified. This report deals with the degradation and distribution of (+)-*trans* and (+)-*cis* isomers of phenothrin in stored wheat grains and also their decomposition through processing and cooking.

MATERIALS AND METHODS

1. Chemicals

(+)-*Trans* and (+)-*cis* isomers of phenothrin labeled with ¹⁴C at benzyl-methylene carbon in the alcohol moiety were synthesized by Hazue and Kamata⁸⁾ of Sumitomo Chemical Co. The specific activity of both preparations was 7.3 mCi/mmol with radiochemical purity of more than 99%, as evidenced by thin-layer chromatography (*tlc*). Malathion [*O,O*-dimethyl *S*-(1,2-dicarboethoxy)ethyl phosphorodithioate] labeled with ¹⁴C at 1,2-ethyl positions was purchased from Radiochemical Centre, Amersham, England and its specific activity was 2.78 mCi/mmol with radiochemical purity of 98%.

The following unlabeled authentic compounds were synthesized in the authors' laboratory^{4,7,9)}: 3-phenoxybenzyl (+)-*trans*-chrysanthemate [*t*-phenothrin], 3-phenoxybenzyl (+)-*cis*-chrysanthemate [*c*-phenothrin], 3-phenoxybenzyl alcohol [PBalc], 3-phenoxybenzoic acid [PBacid], methyl 3-phenoxybenzoate [PBacid-Me], 3-phenoxybenzyl (+)-2,2-dimethyl-3-*trans*-formyl-cy-

clopropane-1-carboxylate [formyl-*t*-phenothrin], 3-phenoxybenzyl (+)-2,2-dimethyl-3-*cis*-formyl-cyclopropane-1-carboxylate [formyl-*c*-phenothrin], malathion, *O*-hydrogen *O*-methyl *S*-(1,2-dicarboethoxy)ethyl phosphorodithioate [desmethylmalathion], *O,O*-dimethyl *S*-(1-carboxy-2-carboethoxy)ethyl phosphorodithioate [malathion α -monoacid], *O,O*-dimethyl *S*-(1-carboethoxy-2-carboxy)ethyl phosphorodithioate [malathion β -monoacid], *O,O*-dimethyl *S*-(1,2-dicarboxy)ethyl phosphorodithioate [malathion diacid].

Piperonyl butoxide [PBO] with the purity of 98.4% was purchased from Takasago Perfumery Co., Hiratsuka, Japan. Fenitrothion or Sumithion® [*O,O*-dimethyl *O*-(3-methyl-4-nitrophenyl)phosphorothioate] of technical grade with the purity of 96.8% was synthesized by Sumitomo Chemical Co. ^{14}C -*t*-phenothrin, ^{14}C -*c*-phenothrin or ^{14}C -malathion in emulsion was prepared as follows; the ^{14}C -preparation, Sorpol 3005X (Toho Chemical Co., Tokyo, Japan) and xylene were mixed at the rate of 1/1/9 (w/w), and diluted suitably with water. An emulsion of ^{14}C -phenothrin plus PBO or ^{14}C -phenothrin plus PBO plus fenitrothion was also prepared from each mixture of ^{14}C -phenothrin-PBO-Sorpol 3005X-xylene (1/5/1/9) and ^{14}C -phenothrin-PBO-fenitrothion-Sorpol 3005X-xylene (1/5/1/1/9).

2. Treatment of Wheat Grains

Wheat grains, variety Horoshiri, harvested in Hokkaido, Japan, in August, 1978, with residues of less than the detectable amount (2 ppb) of both malathion and fenitrothion were kept at $20 \pm 2^\circ\text{C}$ for 3 weeks over saturated magnesium nitrate solution to adjust the moisture content to 10.5%. The grains were treated with each ^{14}C -preparation in emulsion at the rate of 1% relative to grain weight in October, 1978, then mixed well and air-dried. The application rates of phenothrin, phenothrin plus PBO, phenothrin plus PBO plus fenitrothion and malathion were 4, 4+20, 4+20+4 and 15 ppm, respectively. Each treated sample was placed in a stoppered glass vessel (i.d. 14 cm, height 14 cm) and kept at 15 ± 2 or $30 \pm 2^\circ\text{C}$ in the dark. To trap the volatile radioactive products, a 50-ml beaker

containing 30 ml of 6 N potassium hydroxide solution was placed inside each vessel.

The moisture content of the wheat grains was measured at intervals by keeping them at 120°C for 4 hr.¹⁰⁾ Immediately after application of the ^{14}C -preparations, the moisture content of the grains was 11.0%. Thereafter it gradually increased with time, amounting to 12.8 and 12.0% at 15 and 30°C after 12 months, respectively.

3. Analysis of Phenothrin Isomers, Malathion, Fenitrothion and Their Metabolites in Wheat Grains

At specified intervals a 10 g-aliquot of wheat grains was powdered with a mill (Kyoritsu-Riko Co., Tokyo, Japan) for 2 min and 30 ml methanol was added. The mixture was mechanically shaken and kept standing overnight at 4°C . The solvent phase was separated by centrifugation and the residue was extracted further four times with 30 ml methanol. The combined methanol extracts were analyzed for ^{14}C and metabolites by liquid scintillation counting and *tlc* as reported previously.⁷⁾ The *tlc* *Rf* values for phenothrin isomers, malathion and their metabolites are listed in Table 1.

To observe grossly the distribution of radio-carbon, the grains were embedded in paraffin and cut transversely and longitudinally, then the sections were exposed to X-ray films for two weeks.¹¹⁾

For determination of fenitrothion residues in the grains, aliquots of methanol extracts obtained above were subjected to gas-liquid chromatography in the following conditions: apparatus; Shimadzu GC-5A equipped with KBr single crystal FTD,¹²⁾ column; 5% XE-60 on Gas Chrom Q (100-120 mesh) packed into ϕ 3 mm $\times$ l 1.0 m glass column, gas; carrier gas (He) 48 ml/min, hydrogen 27 ml/min, air 0.3 l/min, temperature; column 186°C , injection port 275°C . The retention time of fenitrothion was 3.6 min.

4. Decomposition of Phenothrin Isomers through Processing and Cooking

The flour and bran were prepared as in the following. Thirty-five grams of the grains were mixed with 2 ml of distilled water and stirred with a glass rod to peel off pericarps

Table 1 *Tlc Rf* values for phenothrin isomers, malathion and their metabolites.

Compound	<i>Rf</i> in solvent systems ^{a)}								
	A	B	C	D	E	F	G	H	I
<i>c</i> -Phenothrin	0.61	0.54	0.68	0.47					
<i>t</i> -Phenothrin	0.59	0.54	0.68	0.43					
Formyl- <i>c</i> -phenothrin	0.36	0.31	0.55						
Formyl- <i>t</i> -phenothrin	0.38	0.30	0.57						
PBalc	0.26	0.19	0.35						
PBacid	0.34	0.05	0.46						
PBacid-Me	0.48	0.43	0.60						
Malathion					0.62	0.63	0.61	0.63	0.86
Malathion α -monoacid					0.45	0.46	0.40	0.34	0.76
Malathion β -monoacid					0.46	0.47	0.43	0.40	0.76
Malathion diacid					0.24	0.36	0.23	0.18	0.62
Desmethylmalathion					0.06	0.16	0.02	0.07	0.40

^{a)} A: *n*-hexane/toluene/acetic acid (3/15/2, v/v), B: *n*-hexane/acetone (4/1), C: benzene saturated with formic acid/ether (10/3), D: *n*-hexane/ether (50/1), 7 developments, E: benzene saturated with formic acid/ethyl acetate/acetic acid (8/2/1), F: toluene/ethyl formate/formic acid (5/7/1), G: benzene/ether/acetic acid (8/2/1), H: benzene/ethyl acetate/acetic acid (50/50/3), I: toluene/ethyl acetate/isopropanol/acetic acid (8/12/5/3).

from the grains. Germs were separated from the peeled grains by picking them up with a pincette. Then the grains were powdered with a coffee mill (Nihon Philips Co., Tokyo, Japan) for 2 min and the powdered endosperm (flour) was obtained by passing through a 100-mesh sieve to separate from aleurone layers. The germs, pericarps and aleurone layers were combined and the fraction was designated as bran. Each flour and bran fraction was extracted with methanol and analyzed as described above.

The bread was prepared as follows. Dry yeast (0.3 g), sugar (1.1 g), salt (0.27 g), skim milk (0.5 g), butter (1.0 g) and distilled water (12 ml, 40°C) were added to the flour (20 g), then mixed well and subjected to fermentation (1st, 37°C for 60 min; 2nd, 37°C for 30 min) and baking (200°C for 30 min) with an oven (Type TO-120, Hitachi, Tokyo, Japan). The bread was powdered with a mill and extracted with methanol using a Polytron® (Kinematica GmbH, Luzern, Steinhofhalde, Switzerland). The methanol extracts were concentrated and then were partitioned between acetonitrile and petroleum ether. The acetonitrile layer was analyzed as described above.

All experiments were duplicated. The results are shown as the average values of the

duplicate experiments.

RESULTS

1. Degradation of Phenothrin Isomers in Stored Wheat Grains

As shown in Fig. 1, both isomers of phenothrin were decomposed slowly in the grains even at higher temperature storage. After 12-month storage at 30°C, 79 and 87% of the applied radiocarbon were recovered as intact *t*- and *c*-phenothrin, respectively, and *t*-

Fig. 1 Residue amounts of phenothrin isomers, malathion and fenitrothion in wheat grains stored at 15°C and 30°C.

Table 2 Decomposition of *t*-phenothrin in wheat grains stored at 15°C and 30°C.

Compounds	Tem- perature	Months	% of the applied ¹⁴ C						
			Methanol extracts				Bound	KOH- trap	Total
			Pheno- thrin	Formyl- <i>t</i> -phen.	PBalc	PBacid	PBacid -Me		
<i>t</i> -Phenothrin (4 ppm)	15°C	0	95.3	1.5	0.3	<0.1	<0.1	0.7	100.3
		1	95.1	1.1	0.2	<0.1	<0.1	1.0	98.7
		3	94.6	1.2	0.4	<0.1	<0.1	1.6	99.1
		6	93.1	0.9	0.4	<0.1	<0.1	1.9	97.3
		12	92.0	0.6	0.6	0.4	<0.1	2.1	96.6
	30°C	1	97.5	0.4	0.3	<0.1	<0.1	1.1	100.5
		3	95.1	0.3	0.8	0.4	<0.1	1.5	99.1
		6	95.2	0.3	1.8	1.2	1.5	1.9	103.4
		12	78.9	0.3	3.0	5.4	5.5	2.6	96.9
<i>t</i> -Phenothrin + PBO (4+20 ppm)	30°C	0	93.7	2.0	0.2	<0.1	<0.1	0.7	97.7
		1	98.6	0.9	0.3	<0.1	<0.1	1.5	102.1
		3	95.2	0.6	0.6	<0.1	<0.1	2.1	99.1
		6	91.5	0.4	1.5	1.8	1.6	2.2	99.8
		12	82.5	0.2	1.8	4.3	3.8	3.1	97.0
<i>t</i> -Phenothrin + PBO + Fenitrothion (4+20+4 ppm)	30°C	0	95.5	1.1	0.2	<0.1	<0.1	0.9	99.3
		1	94.6	0.5	0.2	<0.1	<0.1	1.3	97.6
		3	95.0	0.5	0.4	<0.1	<0.1	2.1	98.8
		6	91.5	0.5	0.8	0.6	1.0	1.4	97.0
		12	89.1	0.3	1.0	1.5	2.4	2.5	98.2

Table 3 Decomposition of *c*-phenothrin in wheat grains stored at 15°C and 30°C.

Compounds	Tem- perature	Months	% of the applied ¹⁴ C								
			Methanol extracts					Bound	KOH- trap	Total	
			Pheno- thrin	Formyl- <i>c</i> -phen.	PBalc	PBacid	PBacid -Me				Unknown products
<i>c</i> -Phenothrin (4 ppm)	15°C	0	96.4	0.4	0.3	<0.1	<0.1	2.5	0.6	<0.1	100.2
		1	95.2	0.2	0.2	<0.1	<0.1	0.7	0.1	<0.1	97.0
		3	95.1	0.5	0.4	<0.1	<0.1	1.3	1.2	<0.1	98.5
		6	93.4	0.4	0.6	<0.1	<0.1	2.6	0.8	<0.1	97.8
		12	91.7	0.5	0.8	0.5	<0.1	2.5	1.1	<0.1	97.1
	30°C	1	100.1	0.2	0.3	<0.1	<0.1	0.7	0.8	<0.1	102.1
		3	98.3	0.4	0.7	<0.1	<0.1	1.2	1.3	<0.1	101.9
		6	95.1	0.3	2.0	0.6	1.4	2.3	1.0	<0.1	102.7
		12	86.6	0.3	1.6	1.9	2.8	1.3	2.0	<0.1	96.5
<i>c</i> -Phenothrin + PBO (4+20 ppm)	30°C	0	95.6	1.4	0.2	<0.1	<0.1	2.0	0.8	<0.1	100.3
		1	98.8	0.3	0.3	<0.1	<0.1	0.7	1.2	<0.1	101.3
		3	100.7	0.6	0.6	<0.1	<0.1	0.9	1.8	<0.1	104.6
		6	88.8	0.4	0.9	0.9	0.9	1.8	1.9	0.1	94.8
		12	88.4	0.2	1.0	2.3	2.0	1.6	2.3	0.3	98.1
<i>c</i> -Phenothrin + PBO + Fenitrothion (4+20+4 ppm)	30°C	0	97.8	1.0	0.2	<0.1	<0.1	2.1	0.9	<0.1	102.0
		1	95.9	0.3	0.3	<0.1	<0.1	0.8	1.3	<0.1	98.6
		3	97.3	0.5	0.4	<0.1	<0.1	1.0	2.0	<0.1	101.2
		6	93.0	0.3	0.7	0.4	0.5	1.8	1.7	<0.1	98.4
		12	90.9	0.3	0.8	0.9	1.9	1.1	2.0	0.1	98.0

Fig. 2 Radioautograms of transverse and longitudinal sections of wheat grains treated with ^{14}C -phenothrin isomers and ^{14}C -malathion and stored at 30°C for 1, 6 and 12 months.

phenothrin was somewhat more degradable than *c*-phenothrin (Tables 2 and 3). At lower temperature storage, decomposition of both isomers was negligible. The joint application of phenothrin isomers with PBO or PBO plus fenitrothion did not significantly affect the residual lives of both isomers for up to 6 months. But after 12 months, PBO and PBO plus fenitrothion gave the higher recovery of *t*-phenothrin by 4 and 10% of the applied radiocarbon at 30°C , respectively. This tendency was also observed in the case of *c*-phenothrin.

The degradation products obtained from phenothrin isomers were isolated and then identified by two-dimensional *tlc* cochromatography with authentic standards. PBacid-Me was further confirmed not to be an artifact by extraction with acetone or benzene instead of methanol. Among these products identified, ester-hydrolyzed products such as PBalc, PBacid and PBacid-Me increased gradually, and the sum of these metabolites amounted to 6–14% after 12-month incubation at 30°C . These metabolites were produced in larger amounts from *trans* isomer than *cis* isomer and also at 30°C than 15°C . PBO and PBO plus fenitrothion decreased the production of these metabolites by 1–4 and 3–9% of the applied radiocarbon after 12 months, respectively. Other than the above decomposition products,

formyl-*t*- and formyl-*c*-phenothrin were produced at the early stage of incubation, amounting maximally to 2% of the applied radiocarbon, but their contents decreased with time. Unknown products which contained at least five spots on *tlc*, and volatile radioactive products were detected in small amounts.

2. Distribution of ^{14}C in Wheat Grains and Decomposition of Phenothrin Isomers through Processing and Cooking

As shown in Fig. 2, the distribution of radiocarbon in wheat grains did not change substantially during 12-month storage. The radiocarbon was mainly located at seed coat including the crease area, while relatively little radiocarbon was found in germ and endosperm regions. Actually the contents of phenothrin isomers in flour obtained with our milling procedure were 13 to 15 times lower than those in bran (Table 4). Furthermore the phenothrin residues in flour decreased through the baking process to some extent. No *trans/cis* isomerization was observed during baking. Most of the decomposition products in bread were ester-hydrolyzed products, and *c*-formyl-phenothrin was detected in a trace amount (Table 5).

Table 4 Residues of phenothrin isomers prior to and following processing and cooking.

	Content, ppm ^{a)}			
	<i>t</i> -Phenothrin		<i>c</i> -Phenothrin	
	Storage period (months)			
	6	12	6	12
Wheat grain ^{b)}	3.78	3.13	3.64	3.32
Flour	0.79 (73%) ^{c)}	0.76 (76%)	0.77 (72%)	0.77 (75%)
Bran	11.4 (27%)	11.3 (24%)	9.75 (28%)	11.4 (25%)
Bread	0.69	0.57	0.66	0.57

^{a)} μg phenothrin /g (wet weight).^{b)} The wheat grains were treated with ^{14}C -phenothrin isomers at 4 ppm alone and stored at 30°C in the dark.^{c)} Wet weight ratio of milled fractions.

Table 5 Contents of phenothrin isomers and their decomposition products in flour and bread.

Compound		Storage period (months)	Content, ppm ^{a)}					
			Phenothrin	Formyl-phenothrin	PBalc	PBacid	PBacid-Me	Unknown products
<i>t</i> -Phenothrin	Flour	6	0.79	<0.01	0.02	0.01	0.01	0.01
		12	0.76	<0.01	0.03	0.06	0.08	0.04
	Bread	6	0.69	<0.01	0.03	0.02	0.01	0.04
		12	0.57	<0.01	0.02	0.03	0.05	0.03
<i>c</i> -Phenothrin	Flour	6	0.77	0.02	0.01	0.01	<0.01	0.02
		12	0.77	0.01	0.01	0.02	0.03	0.02
	Bread	6	0.66	0.02	0.03	0.01	0.01	0.03
		12	0.57	0.01	0.01	0.01	0.01	0.03

^{a)} μg phenothrin equivalent/g flour or bread (wet weight).

Table 6 Decomposition of malathion in wheat grains stored at 15°C and 30°C.

Temperature	Months	% of the applied ^{14}C								
		Methanol extracts					KOH-trap			
		Malathion	Malathion monoacids	Malathion diacid	Desmethyl malathion	Unknown products	Bound	$^{14}\text{CO}_2$	Unknown products	Total
15°C	0	94.0	1.7	<0.1	0.4	2.3	1.5	<0.1	<0.1	99.9
	1/2	88.9	2.9	<0.1	2.4	1.9	2.7	<0.1	0.1	98.9
	1	86.0	4.1	0.3	4.5	2.0	3.7	<0.1	0.2	100.8
	3	71.2	6.6	0.2	9.5	2.8	7.6	0.3	0.5	98.7
	6	54.2	8.0	0.7	15.9	3.0	11.8	0.5	1.2	95.3
	12	34.2	7.4	0.5	21.8	5.1	20.3	1.0	2.0	92.3
30°C	1/2	72.3	6.1	0.3	10.9	3.5	9.4	0.6	0.5	103.6
	1	49.9	6.0	0.5	19.6	3.4	14.4	1.3	1.0	96.1
	3	21.5	4.0	0.5	26.9	6.3	31.3	2.3	1.9	94.7
	6	11.0	2.4	0.8	29.0	8.8	37.8	3.8	2.1	95.7
	12	4.5	1.8	0.4	23.5	9.1	50.3	6.2	2.5	98.3

3. Degradation of Fenitrothion and Malathion in Stored Wheat Grains

As shown in Fig. 1, fenitrothion combined with PBO and each phenothrin isomer was rapidly decomposed with respective half-lives of about 12 and 1.5 months at 15 and 30°C. Malathion was decomposed a little faster than fenitrothion. The half-lives of malathion were about 8 and 1.1 months at 15 and 30°C, respectively. Malathion was decomposed majorly to desmethylmalathion, amounting to 29% at its maximum, and to malathion monoacids and diacid, as shown in Table 6. No malaoxon residue was detected throughout incubation at both temperatures. Volatile radioactive products increased gradually with time, amounting to 9% of the applied radiocarbon after 12-month incubation at 30°C. Most of these products were identified as $^{14}\text{CO}_2$. The unextractable residues (Bound in Tables) also increased with time, amounting to 20–50% of the applied radiocarbon after 12 months, but further characterization of these residues was not conducted.

The distribution pattern of ^{14}C -malathion residues is similar to that of ^{14}C -phenothrin residues, as shown in Fig. 2.

DISCUSSION

Phenothrin isomers were decomposed much

more slowly than fenitrothion and malathion in stored wheat grains. The degradation of pyrethroids on stored wheat grains was studied under similar conditions. The respective half-lives of pyrethrin I and allethrin in the grains with 15% moisture content at 30°C were 3 and 4 weeks.¹³⁾ The half-lives of bioresmethrin at 30–35 and 20°C were 8–10 and more than 26 weeks, respectively.¹⁴⁾ From these results, the degradation of phenothrin isomers in wheat grains seems to be much slower than these chrysanthemic esters.

Phenothrin isomers were mainly decomposed via cleavage of the ester linkage to PBalc which was subsequently oxidized to PBacid. PBacid was further converted to PBacid-Me. Both isomers in part underwent cleavage of the isobutenyl double bond to yield formyl-*t*- and formyl-*c*-phenothrin, probably due to air oxidation.⁷⁾ No *trans/cis* isomerization occurred during storage. Oxidation products at methyl groups of the isobutenyl moiety or ring hydroxylation products at 2'- and 4'-positions of 3-phenoxybenzyl moiety, which were found in metabolic studies with rats,^{4–6)} were not detected in the present experiment. The degradation pathways for phenothrin isomers in stored wheat grains are proposed in Fig. 3. Rawlands¹³⁾ showed that pyrethrin I and allethrin were majorly de-

Fig. 3 Proposed degradation pathways for phenothrin isomers in stored wheat grains.

graded to the chrysanthemic acid and the corresponding alcohols and no oxidative products were detected in stored wheat grains. From these findings it seems that little or no oxidative metabolism of chrysanthemic esters occurs in the first steps, hydrolytic degradation being the usual pathway in stored wheat grains. The preferential hydrolysis was observed in the decomposition of malathion in the present study, where no malaoxon was detected.

The radioautograms of sections of wheat grains (Fig. 2) show that little radiocarbon was located in endosperm, however the contents of phenothrin isomers in flour were relatively high. The gap is presumed to be due to contamination of seed coat at the crease area where the radiocarbon was localized into flour during our milling process. In a normal commercial flour milling, only a very small portion of the crease area will contaminate into flour.¹⁵⁾ In fact, the malathion contents in flour were 10–14% of those in wheat grains when a milling machine was used, as reported by Kawamura *et al.*¹⁶⁾ The malathion contents in flour obtained with our milling procedure were about 22% of those in wheat grains.¹⁷⁾ Therefore, in practice the contents of phenothrin isomers in flour appear to be lower than the figures that we found.

These laboratory findings suggest that phenothrin isomers applied to wheat grains seem to remain intact at seed coat and exhibit an excellent insecticidal activity against stored grain pests for a prolonged period. Most of the intact residues on the grains are likely to be removed during milling process and a low level of phenothrin residues will appear in flour or bread.

REFERENCES

- 1) K. Fujimoto, N. Itaya, Y. Okuno, T. Kadota & T. Yamaguchi: *Agric. Biol. Chem.* **37**, 2681 (1973)
- 2) J. H. Ardley & G. R. H. Halls: Chemical Marketing and Economics Division of the American Chemical Society, Preprint, Honolulu, Hawaii, pp. 38–54, 1979
- 3) J. H. Ardley, M. Bengston, J. M. Desmarchelier & P. Skidmore: *J. Stored Prod. Res.* in press
- 4) J. Miyamoto, T. Suzuki & C. Nakae: *Pestic. Biochem. Physiol.* **4**, 438 (1974)

- 5) T. Suzuki, N. Ohno & J. Miyamoto: *J. Pesticide Sci.* **1**, 151 (1976)
- 6) H. Kaneko, H. Ohkawa & J. Miyamoto: *J. Pesticide Sci.* **6**, 169 (1981)
- 7) K. Nambu, H. Ohkawa & J. Miyamoto: *J. Pesticide Sci.* **5**, 177 (1980)
- 8) M. Hazue & T. Kamata: unpublished
- 9) T. Suzuki & J. Miyamoto: *Pestic. Biochem. Physiol.* **8**, 186 (1978)
- 10) S. W. Pixton: *J. Stored Prod. Res.* **3**, 35 (1967)
- 11) Y. Takimoto, M. Ohshima & J. Miyamoto: *J. Pesticide Sci.* **3**, 277 (1978)
- 12) Y. Sato, J. Miyamoto & S. Suzuki: *Botyu-Kagaku* **33**, 8 (1968)
- 13) D. G. Rawlands: "Residue Reviews," ed. by F. A. Gunther, Vol. XXXIV, Springer-Verlag, New York, p. 116, 1971
- 14) FAO/WHO: "1975 Evaluation of Some Pesticide Residues in Food," p. 67, 1976.
- 15) J. Moss: Private communication (1979)
- 16) K. Kawamura, M. Takeda, M. Uchiyama, K. Sakai & H. Ishikawa: *Shokuhin-Eiseishi* **21**, 70 (1980)
- 17) K. Nambu, Y. Takimoto & J. Miyamoto: unpublished

要 約

フェノスリンの貯蔵中の小麦穀粒における分解

南部健二, 滝本善之, 宮本純之

¹⁴C で標識したフェノスリンの (+)-トランスおよび (+)-シス体を水分含量 11% の小麦穀粒におおの 4 ppm の濃度で処理し, 15 または 30°C の暗所に貯蔵した。トランスおよびシス両異性体は徐々に分解し, 12カ月後には 30°C においてそれぞれ添加量の 79 および 87% が小麦穀粒に残存していた。両異性体ともおもにエステル結合の加水分解, ペンジルアルコールの安息香酸への酸化さらに安息香酸のメチル化を経て代謝された。ビペロニルブトキシサイドおよびフェニトロチオンとの混合処理によりフェノスリンの分解はいくぶん抑えられた。フェノスリンとこれらの分解物は 12 カ月間おもに種皮に存在し, 製粉により, 小麦粉およびふすま中の両異性体の残留量はそれぞれ 0.77 および 11.4 ppm となった。この小麦粉を製パンすると残留量は 0.57 ppm に減少した。一方, ¹⁴C-マラチオンは小麦穀粒中 15 および 30°C においてそれぞれ半減期 8 および 1.1 カ月の速度で分解し, デスメチル体, モノカルボン酸体, ジカルボン酸体および ¹⁴CO₂ を生成した。