STUDIES ON THE PHYSICAL DEPENDENCE POTENTIAL OF ANALGESICS

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Abstract—Influence of morphine on the fine structure of mitochondria in the cell of zona fasciculata of adrenal cortex in mice treated chronically with morphine pellets was studied using the electron microscope. The transformation of mitochondrial structure was observed 12 hours after morphine pellet implantation and the degree of transformation reached a maximum at 48 hours. These changes, however, disappeared within 4 days. On the 4th day after implantation, removal of the pellet or levallorphan challenge resulted in alteration of the mitochondria with evidence of withdrawal syndrome. Reinjection of morphine to the mice immediately after removal of the pellet, however, prevented the appearance of such mitochondrial transformation. Chlorpromazine or sodium pentobarbital did not affect on the transformation of mitochondria.

We have already reported that abrupt withdrawal of morphine or challenge of either levallorphan or naloxone in morphine dependent rats resulted in a characteristic transformation of intramitochondrial structure in the adrenal cortex (1-4). The time course of appearance of these changes coincides with that of marked loss in body weight after withdrawal, the latter considered to be one of the best indicators of physical dependence on morphine (5-7). When morphine was readministered, normal structures of the mitochondria were readily restored.

From these results, we suggested that there is a close relationship between these changes and the physical dependence potential and that the transformation of mitochondria may be one of the best indices of physical dependence in rats.

Way et al (8) used the naloxone induced jumping behavior to quantify the degree of physical dependence in morphinized mice. They reported an inverse relationship between the degree of dependence and the amount of naloxone required to induce the jumping responses. Since then, this method has been widely used in many laboratories (7-13). Should the appearance of mitochondrial alteration observed in morphine dependent rats be evident in morphine pellet implanted mice in which it has been confirmed that acute dependence on morphine had developed, it is conceivable that a close relationship between the transformation of mitochondria and the dependence potential would prove to be a phenomena common to all rodents and the mitochondrial structure would be one of the best indicators of the degree of physical dependence potential.

We carried out experiments to determine whether or not such transformation of mitochondria is indeed produced in morphine dependent mice after abrupt withdrawal of the drug.
MATERIALS AND METHODS

Male ICR mice from CLEA Japan INC. (Tokyo) weighing 27 to 30 g were housed in our laboratory for at least 1 week prior to experimentation. Food and water were provided ad libitum.

Morphine pellets used in this experiment were prepared by the method of Way et al (8). Such consisted of morphine base, 75 mg; microcrystalline cellulose, 75 mg; fumed silicon dioxide, 0.75 mg; and calcium stearate, 1.5 mg. Placebo pellets consisted of 75 mg of lactose instead of the morphine base. The 7 mm pellets were implanted s.c. in the dorsal region of mice under ether anesthesia.

To confirm the development of tolerance to and physical dependence on morphine in the pellet implanted mice, the following preliminary experiments were performed.

The pellets were removed at various days after implantation and the absorption rate from the morphine pellets was determined by quantifying the amount of drug remaining at the implantation site (14). Twenty mg/kg of morphine was then injected 6 hours after removal of the morphine pellet and the grade of tolerance was assessed.

The degree of tolerance was measured using an electro-shock method (15) by plotting the mean analgesic response time against the time after injection of morphine on section paper and determined the area under the response curve.

The degree of physical dependence on morphine was assessed by the decrease in body weight and the withdrawal jumping behavior after abrupt withdrawal of morphine or challenge with 10 mg/kg of levallorphan.

In these experiments, we confirmed the following: 1) About 22 mg/kg of morphine was absorbed from the pellet during the initial 4 days and a particularly large amount was absorbed during the 1st and 2nd days. 2) The development of tolerance after morphine pellet implantation was discernible within 24 hr, but progressively increased with increasing time, reaching the maximal state on the 3rd or 4th days. 3) Marked loss of body weight and characteristic withdrawal jumping behavior were induced by abrupt withdrawal of morphine or the levallorphan challenge. In the abrupt withdrawal group, maximal effects occurred at 10 hr (loss of body weight) and 6 hr (jumping), respectively, but in the mice in the levallorphan challenged group maximal effects were seen at 2 hr (loss of body weight) and 15 min (jumping), respectively.

These results are good agreement with those of Maggiolo and Huidobro (16), Huidobro (17) and Way et al (8), and indicate that the development of tolerance to and dependence on morphine were thoroughly induced in these mice. Electron microscopic studies were then done using those mice.

The animals were anesthetized with pentobarbital and prefixed with perfusion of formaldehyde (2%) and glutaraldehyde (2.5%), solution adjusted to pH 7.4 with 0.1 M of Sörensen phosphate buffer which was given through left cardiac ventricle.

The adrenals were removed immediately after perfusion and postfixed with 1% buffered osmium tetraoxide for 3 hr at 4 C. Dehydration was carried out using a graded series of chilled ethanol. The blocks were then embedded in Epon 812 and sections were cut with
glass knives in a Porter Blum MT-I and Reichert Om-U2 ultramicrotome after polymerization in an incubator. To identify the outer region of the zona fasciculata, semi-thin sections (1 μm) were cut from the Epon block, stained with toluidine blue and viewed under light microscope. The ultrathin sections were stained with uranylacetate—lead citrate, and examined under a JEM-100U electron microscope. Ten photographs at relatively low magnification from various regions of outer zone of zona fasciculata were taken and the results examined.

RESULTS

Fine structure of adrenal cortex in normal mice

Zona glomerulosa forms the outermost layer of the adrenal cortex and consists of 3–4 cell layers. It is located between the capsule which is composed of a few fibrous cells with many collagenous fibers and the zona fasciculata. These layers have irregular diamond shaped or cylindrical cells which have an irregular ovoid nucleus, elliptical mitochondria with a comb-like inner structure, lipid droplets and a relatively clear cytoplasm. These cell layers gradually shifted to the zona fasciculata through 1 or 2 layers of the intermediate zone.

As shown in Fig. 1, in the cells of zona fasciculata of normal mice, there are many dark cells which are compactly filled with mitochondria, smooth endoplasmic reticulum and lipid droplets. Characteristic mitochondria are nearly globular in shape with a diameter of about 0.5 μm, but their inner structure varies from cell to cell, i.e., vesicular, tubular, tubulo-vesicular or mitochondrial ghost which decrease in electron density of the matrix and have the appearance of empty mitochondria. Therefore, the inner structure of mitochondria in zona fasciculata of normal mice is considerably different from that of normal rats (1). Size of the mouse mitochondria is slightly smaller and the inner structure is more irregular than that seen in rats. This tendency is more evident in the inner zone of zona fasciculata.

Fig. 1. Electron micrograph of adrenocortical cells of zona fasciculata in control mouse. Mt: mitochondria, Ly: lysosome, Li: lipid droplet, N: nucleus
or in zona reticularis. About 80–85% of mitochondria in zona fasciculata revealed a vesicular or tubulo-vesicular structure with high electron density, and the remaining 15–20%, of mitochondria showed a tubular or empty type with low electron density, in other wards, transformed mitochondria as seen in dependent rats were also observed even in normal mice.

Effects of the morphine pellet implantation

Twelve hr after implantation of morphine pellets, the cells of zona fasciculata became relatively clear with an increasing endoplasmic reticulum. Mitochondria of these cells began to swell and at the same time intramitochondrial structure became tubular or had a concentric lamellar structure with an empty area in their matrix. These altered mitochondria nearly doubled in the number during this time and then gradually increased. A

![Fig. 2. Mitochondria of zona fasciculata of adrenal cortex in mouse implanted morphine pellet for 24 hours.](image1)

![Fig. 3. Mitochondria of zona fasciculata of adrenal cortex in mouse implanted morphine pellet for 96 hours.](image2)
maximum state was reached (about 60%) at 48 hr as shown in Fig. 2. Three days after pellet implantation, these altered mitochondria diminished rapidly despite the continuance of pellet implantation and a normal state was reverted to within 4 days (Fig. 3).

We also investigated the influence of placebo pellet in normal mice, and no considerable changes were found. These findings indicate that the operative injury with the pellet implantation did not affect the transformation of mitochondria (Fig. 4).

Effects of the morphine pellet removal or levallorphan challenge

Four days after morphine pellet implantation when morphological changes disappeared and a normal appearance was seen, pellets were removed and the intramitochondrial structure was investigated at various intervals. Three hr after removal of the pellet, the mitochondria began to swell and inner membrane of some mitochondria showed a tubular or lamellar structure. These changed mitochondria gradually increased but mitochondrial ghosts were comparatively few at this time. Six hr later, such diminished temporarily, but gradually increased thereafter and reached a maximum state at 18 hr after removal of the pellet (Fig. 5).

Fig. 4. Transformation of mitochondrial structure in zona fasciculata of adrenal cortex in mice after implantation of morphine pellet or placebo pellet.

Fig. 5. Eighteen hours after removal of morphine pellet implanted for 96 hours.
One of the characteristic features of these mitochondria was a decrease in electron density of the mitochondrial matrix and an increase in the number of empty mitochondria. These mitochondrial ghosts or empty mitochondria formed the majority in altered mitochondria. Such began to decrease after 24 hr and normal structures were reverted to within 72 hr.

Since the tubular or empty mitochondria were transiently observed in normal mice 3 hr after removal of the placebo pellet, the appearance of altered mitochondria seen after removal of morphine pellet apparently increased within 18 hr and gradually decreased thereafter.

In the next experiment, when 10 mg/kg of levalorphan was administered without removal of the pellet from mice in which the morphine pellet had been implanted 4 days previously, no significant changes had occurred within 3 hr, but 6-12 hr later, similar changes as seen in the abrupt withdrawal mice were evident (Fig. 6, 7). At this time, empty mito-
Chondria were also noted to be in the majority in the altered mitochondria. The appearance of these changed mitochondria induced by levallorphan challenge occurred earlier than that caused by abrupt withdrawal and disappeared within 48 hr. A slight increase of smooth endoplasmic reticulum accompanied by the transformation of mitochondria was observed, but there were no significant changes in nucleus, Golgi apparatus and lipid droplets.

Effects of readministration of morphine

When 80 mg/kg s.c. of morphine was readministered to the mice in which the morphine pellet had been removed 6 hr previously, not only was the appearance of altered mitochondria inhibited, but a normal appearance was reverted to within 4–6 hr. Two hr after readministration, electron density of mitochondrial matrix increased significantly and at the same time vesicular or tubulo-vesicular structure began to increase and smooth endoplasmic reticulum became more evident (Fig. 8). Eight hr after readministration, however, the number of altered mitochondria showed a slight increase (Fig. 9).

**Fig. 8.** Two hours after administration of morphine at 6 hours after removal of morphine pellet.

**Fig. 9.** Effect of morphine administration on the transformation of mitochondria followed by removal of morphine pellet or placebo pellet.
As described above, a slight and transient change of mitochondrial structure occurred after removal of placebo pellet. At this time, changed mitochondria increased even more with administration of 20 mg/kg s.c. of morphine.

Effects of various agents on the transformation of mitochondria

Four days after implantation of the morphine pellet, mice were given 20 mg/kg of chlorpromazine s.c. or 30 mg/kg of sodium pentobarbital i.p. These mice were then challenged with 10 mg/kg of levallorphan. Six hr later, the mitochondrial inner structure was investigated.

After injection of chlorpromazine or sodium pentobarbital, percentages of the altered mitochondria were 51.7 and 54.1, respectively. These agents alone apparently had no effect on the mitochondrial structure, therefore probably played no role in the transformation of mitochondria.

DISCUSSION

Morphine pellet implantation was first described by Maggiolo and Huidobro (16) and later, Way et al (8) improved this method. These authors reported that tolerance to and physical dependence on morphine rapidly developed by implantation of morphine pellet and that such reached a maximum state at 2–4 days. Our data in the present experiments are in good agreement.

We previously reported that there is a close relationship between the dependence potential of narcotic analgesic drugs and the transformation of mitochondria of zona fasciculata in rats and that the latter should be one of the best indices of physical dependence on morphine (1–4). This assumption was confirmed also in mice, in the present experiments (Fig. 10).

It is well known that the appearance of transformed mitochondria exhibits a lowering of corticosteroid biosynthesis (2, 18–22). This decrease may be induced during the withdrawal period in morphine dependent mice as well as rats. If such is indeed correct, then why is the transformation of mitochondria, similar to that observed during the period of

![Fig. 10](image_url)  
Comparison of the schematic representation of ultrastructural changes of zona fasciculata mitochondria of mouse and rat adrenal cortex under various treatments.
withdrawal, induced in the early stage of pellet implantation and thereafter disappears? What possible explanation is there for the discrepancy between the findings that a single dose of morphine stimulates the hypophyseo-adrenocortical axis (23-25) and the results of our present experiments?

Yano et al. (1-4) found that the transformation of mitochondria seen in withdrawal period was induced at 6-12 hr after a single injection of morphine, but they did not give a full explanation for this phenomenon.

Yamamoto et al. (26-28) gave a continuous infusion of morphine to rats in order to elucidate this contradictory phenomenon. They reported that the tolerance to and dependence on morphine rapidly developed in a short period when this method was used, but that the time course of the disappearance of tolerance did not always coincide with that of dependence. They also suggested that the alterations of mitochondria may thus be the result of differences in duration of tolerance and physical dependence.

Thus, the transformation of mitochondria observed 1-2 days after implantation of morphine pellet in mice may be due to over development of dependence rather than that of tolerance and the following disappearance of altered mitochondria may be the result of a development of tolerance which parallels that of dependence.

REFERENCES


