Effects of Bisphenol A (BPA) on Placentation and Survival of the Neonates in Mice

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Abstract. The effects of bisphenol A (BPA) on placentation have not been fully determined. The aim of this study was to clarify the structural changes of the placenta, abortion rate, and survival of neonates after BPA administration in mice. BPA (10 mg/kg/day) was administered to pregnant mice (BPA mice) subcutaneously from the first day of pregnancy (Day 0) to Day 7 (8 days total). The number of embryos and weights of whole uteri were measured on Days 10 and 12. Morphological changes in the placentae were examined by light microscopy on the corresponding days of pregnancy. The number of neonates was also counted. Survival rates were periodically calculated for neonates from the first day after parturition (P-Day 0) to P-Day 56. The number of embryos and weight of the uterus on Days 10 and 12 were significantly decreased by BPA injection. No notable differences were recognized between the left and right uteri. The proportion of the labyrinthine zone per whole placenta in the BPA mice became lower than that in the controls, and that of the metrial gland was higher in the BPA mice. The intervillous spaces of the placenta were narrower in the BPA mice. Degenerative changes were found in the trophoblastic giant cells and spongiotrophoblast layers of the BPA mice. The number of BPA mouse neonates was drastically decreased within 3 days after birth, and no mice survived after P-Day 56. The results suggest that BPA not only disrupts placental functions and leads to abortion through chronic stimulation of gene expression by binding to DNA but that it also affects the mortality of neonates through indirect exposure of embryos.

Key words: Bisphenol A, Placenta, Neonate, Mouse, Survival rate

Bisphenol A (BPA: 2,2-Bis(4-hydroxy-phenyl)propane) interacted with estrogen receptors (ERs) and has weak estrogen activity (approximately 1/10,000 of 17β-estradiol affinity to ERs) [1]. BPA, which is an endocrine disruptor, has been used as a basic ingredient in polycarbonate and/or epoxy resin [2, 3]. Therefore, plastic containers for household use and/or dental sealants contain BPA [4]. Problems due to exposure of humans to BPA via these products have arisen in recent years. For example, BPA was detected at concentrations of up to 30 µg/ml in saliva from patients who had dental sealants applied to their teeth [4].

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The effects of BPA on female reproductive organs and the weights of the uterus are increased by administration of high doses of BPA (up to 800 mg/kg) [5, 6]. Injection of BPA (500 mg/kg) to ovariectomized rats caused a 67% increase in uterine weight and proliferation of uterine stromal cells [7]. A medium dose of BPA (30 mg/kg) resulted in a 37% increase in the uterine weights of ovariectomized Sprague-Dawley rats [8]. Differences in response to BPA among rat strains and grafting of the capsule to release a low dose of BPA (0.3 mg/kg/day) subcutaneously increased the uterine weight and height of the uterine luminal epithelial cells in ovariectomized F344 rats [9]. On the other hand, no detectable changes were observed in the Sprague-Dawley rats used in the same experiment as the F344 rats [9].

The effects of BPA on the pregnant uterus have not been fully examined. Although DNA microarray has shown that the mRNAs of some nuclear receptors in the mouse placenta are increased by BPA administration [10], no detailed information is currently available about the effect of BPA on placentation and/or the fate of embryos and neonates. The present study shows the morphological changes in the pregnant uterus during placentation after BPA administration in mice. Embryo and neonate mortality was also investigated.

Materials and Methods

Ten to 12-week-old virgin mice (ICR strain; Clea Japan, Osaka, Japan) were used in this study. They were housed under a 12 h light and dark cycle and were provided with water and food ad libitum. The first day of pregnancy (Day 0) was defined as the day when a vaginal plug was detected. The first day after birth was designated P-Day 0. All experimental procedures were based on the guidelines of the committee for animal welfare at Yamaguchi University.

BPA (Wako Chemicals, Osaka, Japan) dissolved in sesame oil (Sigma-Aldrich, St. Louis, MO, USA) was injected into the mice subcutaneously for 8 days from Day 0 to Day 7. Mice injected with vehicle from Day 0 to Day 7 were used as the control. Placentae were excised under pentobarbital anesthesia on Days 10 and 12. After counting the number of embryos and measuring the uterine weight, all placentas were fixed in Bouin’s solution. Uterine weight per body weight of the mother was calculated. In addition, the number of neonates was counted to calculate the survival rates of neonates for P-Day 0 to 3 and P-Day 56 using the following formula: the number of living neonates/the number of dead neonates. Values were evaluated by one-way ANOVA. For light microscopy, fixed tissues were dehydrated in a graded series of ethanol, infiltrated in xylene, and embedded in paraffin wax. Paraffin sections were prepared at 4 µm thickness and stained with hematoxylin and eosin. The thickness of the layers of the whole placenta, metrial gland, decidua basalis, and placental labyrinth were measured using sections with the maximum thickness for the layer of the whole placenta, and the proportions of each tissue were calculated as follows: metrial gland/whole placenta, desidua basalis/whole placenta, and placental labyrinth/whole placenta. Mean values for the thickness of each layer were calculated using 5 serial sections from 5 individuals. Each value was statistically analyzed by Welch’s t-test after examining equal variance by F-test.

Results

The number of embryos, uterine weights, and proportions of uterine weight per body weight of the mother are shown in Table 1. All indexes were significantly lower in the BPA mice than in the controls on Days 10 and 12. There were no differences between the left and right uteri (data not shown).

The metrial gland, decidua basalis, and placental labyrinth were easily discriminated for both the control and BPA mice (Fig. 1a and b), although the spongiosotrophoblastic layer could not be clearly located at low magnification in the BPA mice on Day 12. The placentae of the controls were larger than those of the BPA mice on Day 12 (Table 2), although no significant difference was detectable between the BPA mice and controls on Day 10. The difference in the proportions of each tissue were less conspicuous on Day 10 than on Day 12. The proportions of the metrial gland were significantly higher in the BPA mice than in the controls on Day 12. On the other hand, the proportions of the decidua basalis and placental labyrinth were
significantly lower in the BPA mice than in the controls on Day 12. There were no detectable changes in the proportions of the decidua basalis between the BPA and control mice on Day 10.

No abnormalities were found in formation of the network of fetal blood vessels in the BPA mice. On the other hand, the intervillous spaces, through which maternal blood flows, were narrowed in the BPA mice on Days 10 and 12 (Fig. 1c and d). Trophoblastic giant cells, which usually form a band at the boundary line between the decidua basalis and placental labyrinth in normal mice, were not sequentially aligned in the BPA mice (Fig. 1c and d). The BPA mice had fewer glycogen-containing cells and thinner spongiotrophoblastic layers than the controls (Fig. 1e and f). The survival rate of the control neonates was almost 100% within 56 days of birth. Approximately two thirds of the BPA neonates that were born alive died within a day after birth (Fig. 2). Most of the neonates were unable to survive for three days. None survived at 56 days after birth.

**Discussion**

This study revealed that administration of BPA at a dose of 10 mg/kg from Day 0 to 7 leads to reductions in the number of embryos and uterine weight and influences placentation, especially development of the decidua basalis and trophoblastic layers. In addition, BPA decreased the number of neonates and increased their mortality.

Estrogen is important for successful pregnancy. Estrogen regulates migration of the embryo by affecting muscular layers in the oviduct [11] and induction of embryonic implantation to the uterus [12]. A single estrogen surge in the morning on Day 3 is the first signal for initiation of implantation in normal mice [13, 14]. Leukemia inhibitory factor (LIF), which can substitute for estrogen, is induced immediately after estrogen and is essential for successful implantation [12]. LIF leads to apoptotic cell death of the luminal epithelial cells at the point of embryo attachment to the uterus, and this is followed by decidualization and closure of the luminal epithelium at implantation sites [15, 16]. BPA might disrupt the motility of muscular layers of the oviduct and/or the above sequential phenomena of embryo implantation by binding estrogen receptors, ultimately resulting in a reduction of the number of embryos during pregnancy.

The decrease in uterine weight in this study contradicts the findings of previous reports of BPA administration experiments for rats [5, 6, 8, 9, 17]. The decrease in this study might have resulted from fetal loss during pregnancy. This study does not disprove the possibility that there are species differences between mice and rats in the terms of response of the uterus to BPA.

Although the thickness of the whole placenta decreased, the proportion of the metrial gland to the whole placenta increased in the BPA mice. The actual thickness of the metrial gland was thick compared with the controls (Table 1, Fig. 1a and b). Estrogen receptors are localized in the luminal epithelium, glandular epithelium, and/or stromal cells of the uterus from Days 1 to 8 [18, 19]. BPA probably stimulates stromal cells constantly, and BPA administration probably causes hyperplasia of the metrial gland after irreversible binding of its metabolites to the DNA of the stromal cells in each of these tissues [20]. There might be other mechanisms of BPA signaling in the decidua basalis and trophoblastic layers, since the reverse phenomenon to hyperplasia appeared between the decidua basalis and metrial gland. The effects of BPA on trophoblasts are probably indirect via constant stimulation of other cells by BPA because trophoblasts do not have estrogen receptors [18, 21].

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**Table 1. Effects of BPA on abortion and uterine weight**

<table>
<thead>
<tr>
<th>Day</th>
<th>Group</th>
<th>Number of embryos</th>
<th>Uterine weight (g)</th>
<th>Uterine weight (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>Control</td>
<td>16.2 ± 1.64a</td>
<td>2.184 ± 0.109b</td>
<td>5.31 ± 0.444c</td>
</tr>
<tr>
<td>10</td>
<td>BPA</td>
<td>4.6 ± 1.14a</td>
<td>0.542 ± 0.063b</td>
<td>1.40 ± 0.098c</td>
</tr>
<tr>
<td>12</td>
<td>Control</td>
<td>16.4 ± 2.61d</td>
<td>5.706 ± 0.657c</td>
<td>12.97 ± 1.740d</td>
</tr>
<tr>
<td>12</td>
<td>BPA</td>
<td>4.8 ± 0.84d</td>
<td>1.144 ± 0.038c</td>
<td>2.63 ± 0.221f</td>
</tr>
</tbody>
</table>

Values with the same superscript letters are significantly different (P<0.01). Values are shown as the mean ± SD (n=5).
The trophoblastic giant cells of the BPA mice were poorly developed in this study. It has been suggested that trophoblastic giant cells controlled maternal blood flow [21] and that their angiogenic factors are important for successful placentation [22, 23]. BPA possibly binds to cell species other than trophoblasts and disrupt cytokine networks involved in trophoblastic functions. The intervillous spaces, which are formed by trophoblasts and through which maternal blood
**Table 2.** Morphological changes in the placenta of BPA mice

<table>
<thead>
<tr>
<th></th>
<th>Whole placenta (µm)</th>
<th>Metrial gland (%)</th>
<th>Decidua basalis (%)</th>
<th>Placental labyrinth (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day 10 Control</td>
<td>1798 ± 81.31</td>
<td>25.2 ± 2.28&lt;sup&gt;a&lt;/sup&gt;</td>
<td>36.8 ± 1.83</td>
<td>38.0 ± 0.88&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Day 10 BPA</td>
<td>1903 ± 34.69</td>
<td>39.7 ± 1.29&lt;sup&gt;a&lt;/sup&gt;</td>
<td>38.4 ± 1.83</td>
<td>22.7 ± 1.11&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Day 12 Control</td>
<td>2401 ± 51.60&lt;sup&gt;c&lt;/sup&gt;</td>
<td>22.9 ± 1.36&lt;sup&gt;d&lt;/sup&gt;</td>
<td>33.0 ± 2.16&lt;sup&gt;e&lt;/sup&gt;</td>
<td>44.0 ± 1.90&lt;sup&gt;f&lt;/sup&gt;</td>
</tr>
<tr>
<td>Day 12 BPA</td>
<td>1814 ± 39.98&lt;sup&gt;c&lt;/sup&gt;</td>
<td>42.0 ± 0.99&lt;sup&gt;d&lt;/sup&gt;</td>
<td>27.1 ± 1.80&lt;sup&gt;e&lt;/sup&gt;</td>
<td>30.9 ± 1.50&lt;sup&gt;f&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

Values with the same superscript letters are significantly different (P<0.01). Values are shown as the mean ± SD (n=5).

The effects of BPA on cytokine production should be examined at the protein level. The reduction in the number of neonates and increased mortality of the BPA neonates are consistent with previous results when a high dose of BPA was administered to pregnant mice [25]. Polychlorinated biphenyl (PCB), diethylhexyl phthalate (DEHP), and/or dichloro-diphenyl-trichloroethane (DDT), which are representative endocrine disruptors, bind to estrogen receptors [7], decrease the number of embryos and neonates, and increase the mortality of neonates. Comparative morphological evaluation of the placenta with administration of not only BPA but also the other above-mentioned molecules is important for assessment of the effects of estrogen receptor binding disruptors on placentation.

BPA also influences parental care for neonates [26]. The extremely low survival rate in this study probably resulted from not only spontaneous disorder of the neonate by indirect exposure to BPA in the first trimester of pregnancy, but also behavioral abnormality by the mother, although we could not detect any behavioral disorders in the mothers in this study.

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**References**


