The Role of Tumor-Associated Macrophages on Serum Soluble IL-2R Levels in B-Cell Lymphomas

Akira Sakai1) and Noriaki Yoshida2,3)

Interleukin-2 receptor (IL-2R) is comprised of three different subunits (α, β, and γ chain) and is expressed on B cells and NK cells besides T cells. CD25 is also known as the IL-2Rα chain on cell membranes, while soluble IL-2R (sIL-2R) is generated by the proteolytic cleavage of the IL-2Rα chain. Levels of sIL-2R in sera are monitored as a marker of disease activity in patients with lymphoma. However, elevated serum sIL-2R levels are also found in inflammatory diseases, such as infectious diseases. Levels of sIL-2R in sera are thought to reflect tumor burden in adult T-cell leukemia/lymphoma due to the expression of CD25 on tumor cells. Conversely, sIL-2R is thought to be mainly derived from activated T cells infiltrating tumor tissues in diffuse large B-cell lymphoma (DLBCL) and follicular lymphoma (FL) because lymphoma cells except for a subtype of DLBCL are mainly negative for CD25. Matrix metalloproteinase-9, a protease capable of cleaving the membrane bound IL-2Rα chain, is mainly produced by tissue-associated macrophages. Increased macrophages in tumor tissues are reported to be associated with poor prognosis, especially in Hodgkin’s lymphoma. We found increased macrophages in DLBCL and FL compared with reactive lymphoid hyperplasia as well as a positive correlation between the levels of sIL-2R in sera and the number of macrophages in tumor tissues in FL and extranodal DLBCL. (J Clin Exp Hematop 54(1): 49-57, 2014)

Keywords: soluble interleukin-2, tumor-associated macrophage, matrix metalloproteinase

INTRODUCTION

In clinical practice, the levels of soluble interleukin-2 receptor (sIL-2R) in sera are monitored in patients that are suspected of having malignant lymphoma. sIL-2R is generated by the proteolytic cleavage of membrane bound IL-2Rα (CD25),1 and the levels of sIL-2R in sera have proven to be a useful marker of tumor burden in adult T-cell leukemia/lymphoma (ATL/L).1,2,3 caused by human T-cell leukemia virus type 1 infection of CD4+ T cells, due to CD25 being expressed mainly on CD4+ T cells. Subsequently, the levels of sIL-2R in sera are monitored in B-cell lymphomas as well, because of the theory that sIL-2R levels reflect disease activity.4,5 Furthermore, elevated serum sIL-2R levels are found in inflammatory diseases such as infectious diseases, and are related with disease activity. Recently, the levels of serum sIL-2R have been shown to have predictive value for patients with acute-type and lymphoma type ATL/L.6 However, sIL-2R is not included in the international prognostic index (IPI) for diffuse large B-cell lymphoma (DLBCL),7 or in the follicular lymphoma international prognostic index 2 (FLIPI2) for follicular lymphoma (FL).8 Therefore, it is important to elucidate which cell types release sIL-2R and how the levels of serum sIL-2R correlate with these diseases.

IL-2 AND IL-2R

Interleukin-2 (IL-2) was discovered in 1976 as a T-cell growth factor in the supernatants of activated T cells.9 IL-2 is essential in the development of CD4+ CD25+ Foxp3+ regulatory T (Treg) cells,10,11 which are induced from naïve T cells by T cell receptor stimulation in the presence of transforming growth factor-β (TGF-β) and IL-2.12 In the absence of IL-2, Treg cells cannot survive or expand in the thymus or in the peripheral blood.13-16 IL-2R comprises three different subunits: α, β, and γ chains (Fig. 1). Among these, the IL-2 receptor α chain (IL-2Rα, CD25, Tac antigen) is IL-2-specific and expressed on activated, but not non-activated lymphocytes.17 IL-2Rα binds IL-2 with low affinity and forms a complex with IL-2Rβ (CD122) and IL-2Rγ (common
cytokine receptor γ chain, γc, CD132). The IL-2Rβ/γc complex is mainly expressed on memory T cells and NK cells that bind IL-2 with intermediate affinity. All three receptor chains are co-expressed on activated T cells and Treg cells and bind IL-2 with high affinity. IL-2Rβ is also a key component of the IL-15 receptor, whereas IL-2Rγ is an essential component shared by the receptors for IL-2, IL-4, IL-7, IL-9, IL-15, and IL-21.18,19 IL-2 itself acts primarily on lymphoid populations, including T cells,20 B cells,21 and NK cells.22 IL-2 signals, via the heterodimerization of the IL-2Rβ and IL-2Rγ cytoplasmic domains, lead to the activation of at least three major signaling pathways: phosphoinositol 3-kinase (PI3-K)/AKT, Ras-MAP kinase, and the JAK-STAT pathways with JAK1 and JAK3.19,23,24

sIL-2R is generated by enzymatic cleavage and does not appear to be the product of a unique post-transcriptional splicing event.1 The levels of sIL-2R are elevated in sera and other biologic fluids from patients with a variety of cancers and autoimmune diseases, and sIL-2R levels have proven to be a useful marker of disease activity and progression.25,26 Interestingly, sIL-2R is not simply a serum component, as it is known to have a function. Recently, Yang et al. demonstrated the biologic relevance of sIL-2R in terms of its effects on IL-2-mediated T-cell function in follicular lymphoma (FL).27 They observed that serum sIL-2R levels were elevated compared with controls and that elevated pre-treatment sIL-2R levels were associated with poor outcome. Furthermore, they determined the effects of sIL-2R on IL-2 signaling and revealed that a sIL-2R-IL-2 complex promoted T-cell differentiation to inhibitory Treg cells, rather than to Th1 or Th17 cells (CD4+ T helper (Th) cells), and inhibited CD8+ T cells. Therefore, these findings indicate that sIL-2R plays an active biological role in the microenvironment of FL. In addition to our study,28 Yang et al. and others have shown that T cells expressing surface IL-2Ra (CD25) are the major source of sIL-2R in B-cell lymphomas including FL.29,30

**CLINICAL SIGNIFICANCE OF sIL-2R LEVELS IN B-CELL LYMPHOMAS**

It is reasonable to predict disease activity based on serum levels of sIL-2R in CD4+ T-cell lymphoma, including ATL/L in which tumor cells are CD25 positive (Fig. 2). However, B cells are mainly negative for CD25. In order to determine whether sIL-2R has prognostic value in B-cell lymphomas, we retrospectively analyzed DLBCL and FL patients (Fig. 3).28 Patients were divided into two groups according to pre-treatment sIL-2R levels (> 1,500 and ≤ 1,500). In DLBCL, patients with high sIL-2R showed poor prognosis when compared to patients with low sIL-2R. Conversely, patients with high sIL-2R in FL tended to have poor prognosis with no significant differences observed. We then performed a flow cytometric study to identify the types of cells expressing CD25 in B-cell lymphoma samples (Fig. 4). We found three representative patterns in DLBCL: First, most lymphoma cells (CD19+ cells) are positive for CD25, while T cells (CD3+ cells) are partially positive for CD25. Second, both lymphoma cells and T cells are partially positive for CD25. Third, most lymphoma cells are negative for CD25 and T cells are partially positive for CD25. In FL, T cells are...
partially positive for CD25 and lymphoma cells are partially positive for CD25 or mostly negative for CD25. In reactive lymphoid hyperplasia (RLH), T cells are partially positive for CD25 and B cells (CD19+ cells) are mostly negative for CD25. On the other hand, tumor cells in mantle cell lymphoma (MCL) are positive for CD25. Subsequently, analysis of the relationship between serum level of sIL-2R and the degree of CD25+ lymphoma cells or T cells and other clinical data showed that sIL-2R levels in sera do not precisely indicate tumor burden in DLBCL and FL.28

MECHANISM OF sIL-2R PRODUCTION

sIL-2R is generated by the enzymatic cleavage of IL-2Rα (CD25).1,31 However, expression of CD25 in tumor cells does not clarify the mechanisms responsible for the elevation of sIL-2R in DLBCL and FL.28 One of the matrix metalloproteinases (MMPs), MMP-9, attracts attention as a protease capable of cleaving IL-2Rα chains on cell membranes (Fig. 1). Additionally, MMP-2 is a gelatinase-containing gelatin binding repeats similar to MMP-9 and is thought to have the ability to cleavage IL-2Rα chains. Furthermore, these MMPs are produced by macrophages, neutrophils, and dendritic cells, such as inflammation related cells.32,33 We also confirmed that MMP-9 is able to cleave IL-2Rα chains depending on its concentration, using an ATL cell line (MT4) that does not express endogenous MMP-9.28 However, the detection of sIL-2R in the supernatants of MT4 cells depending on concentration, using an ATL cell line (MT4) that does not express endogenous MMP-9.28 Therefore, it is important to determine which cells generate and release MMP-9. According to previous studies,24-39 we speculated that the cleavage of IL-2Rα chains on lymphoma cells and activated T cells, except for tumor cells in ATL/L, using immunohistochemistry (IHC) (Fig. 5).28 It is noted that termed tumor-associated macrophages (TAMs) were positive for MMP-9 in IHC experiments (Fig. 5).28 However, the mechanism involved is not simple (Fig. 6); the number of TAMs, identified as CD68 (KP1)+ or CD163+ macrophages, in DLBCL and FL was significantly higher than in RLH.28 Evaluation of the correlation between levels of sIL-2R and MMP-9 in sera showed a positive correlation in FL but not in DLBCL.28 However, evaluation of the correlation between levels of sIL-2R in sera and number of TAMs showed a positive correlation in FL and extranodal DLBCL but not in all cases with DLBCL.28

TUMOR-ASSOCIATED MACROPHAGES (TAMs)

Macrophages that infiltrate tumor tissues are referred to as TAMs.40,41 Classically activated macrophages (M1) have tumoricidal activity and elicit tissue destructive effects following exposure to interferon γ (IFNγ). Alternatively activated macrophages (M2), in response to IL-4 and IL-13, have cancer related inflammatory (CRI) activity.42-44 In particular, studies in solid tumors reported that TAMs play critical roles in tumor growth, progression, invasion, metastasis, and angiogenesis.44 TAMs are identified as CD68+ or CD163+ cells. Although the KP1 clone for CD68 has been reported to be a less specific marker for macrophages,45 there are some studies that predict prognosis by analyzing the number of macrophages in tumor tissues.46-51 Interestingly, a study in classic Hodgkin’s lymphoma revealed that an increased number of CD68+ macrophages was correlated with a decreased progression-free survival as well as with an increased likelihood of relapse after autologous hematopoietic stem-cell transplantation, resulting in shortened disease-specific survival.48 Furthermore, a study using tissue microarray multivariate analysis showed increased CD68 or CD163 expression to be significant independent predictors of inferior failure free survival or overall survival (OS).50 On the other hand, analysis of increased CD31+ angiogenic sprouting in poor
Fig. 3. Overall survival stratified according to soluble interleukin-2 receptor (sIL-2R) levels (≤ 1,500 vs. > 1,500) in diffuse large B-cell lymphoma (DLBCL) (A) and follicular lymphoma (FL) (B). Serum sIL-2R levels were analyzed in previously untreated patients with DLBCL (n = 104) or FL (n = 30). The 5-year overall survival rates for patients with sIL-2R levels of ≤ 1,500 U/mL and > 1,500 U/mL were 76% and 62%, respectively (p < 0.05) in DLBCL, and 100% and 79.3%, respectively (p = 0.189) in FL.
prognosis FL demonstrated an association between increased sprouting and elevated numbers of infiltrating CD163+ macrophages.\textsuperscript{46} Furthermore, elevated CD68+ TAMs and an increased proportion of CD163+ TAMs were associated with a worse clinical prognosis in ATL/L.\textsuperscript{51} Based on the molecular features, using cDNA microarray, of tumor-infiltrating immune cells in FL, T cells-related gene expression was associated with favorable prognosis, whereas monocyte-related gene expression was associated with poor prognosis.\textsuperscript{52} Therefore, increased infiltration of macrophages in the tumor microenvironment is thought to be associated with poor prognosis, however, the mechanism remains unclear. TAMs produce various cytokines and chemokines. In particular, vascular endothelial growth factor (VEGF) and MMPs, including MMP-9, are also produced by TAMs and enhance angiogenesis, blood vessel permeability, and destruction of the extracellular matrix (ECM) in the tumor microenvironment.\textsuperscript{33,44} In mice study, IL-10 released from macrophages acts in a paracrine manner on Treg to maintain Foxp3 expression.\textsuperscript{53} Therefore, Treg might increase in the microenvironment where macrophages increase (Fig. 6). Altogether, TAMs might produce a permissive environment for lymphoma cell growth and survival.

**FUTURE PERSPECTIVES**

First, to elucidate the mechanism of sIL-2R production in lymphoma, it is necessary to reanalyze the relationship between lymphoma cells and the tumor microenvironment or tumor infiltrating cells. To date, primary therapeutic strategies target lymphoma cells according to phenotypic analyses and the histopathological classification. Furthermore, many
researchers focus on lymphomagenesis induced by genetic or chromosomal aberrations. However, given that the tumor microenvironment is a reservoir for minimal residual disease (MRD), which is a crucial problem associated with relapse disease, combination of drugs that kill lymphoma cells and modulate bystander cells might be promising. Therapies based on monoclonal antibodies to MMPs have already been attempted.\textsuperscript{34} We also found that down-regulation of MMP-9 activity by shRNA transfection decreases the matrigel invasion activity in MMP-9 producing ATL cell lines (not published data).

Second, we suggest that CD25\textsuperscript{+} DLBCL is a subtype of DLBCL according to our two-color (CD25/CD19) staining analysis (Fig. 4). Characterization of CD25\textsuperscript{+} DLBCL could be interesting because we found that recombinant IL-2 enhances the proliferation of CD25\textsuperscript{+}/CD19\textsuperscript{+} B cells, which were purified from the lymph nodes of patients with RLH (not published data). Therefore, IL-2 might play a critical role in CD25\textsuperscript{+} DLBCL.

**ACKNOWLEDGMENTS**

The authors would like to thank the members of The Hiroshima Lymphoma Study Group for the great collabora-
REFERENCES
11 Fontenot JD, Rasmussen JP, Gavin MA, Rudensky AY: A function for interleukin 2 in Foxp3-expressing regulatory T cells. Nat...


