Letter to the editor

Hyaline vascular Castleman’s disease representing 18 trisomy

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TO THE EDITOR

We herein report a case of Castleman’s disease (CD) hyaline vascular (HV) type representing 18 trisomy.

A 69-year-old Japanese male was admitted with a history of right cervical lymphadenopathy lasting for several months. CT demonstrated right cervical and bilateral hilar lymphadenopathy. The results of the full blood count with differential analysis were normal. The biochemical profile, including serum lactate dehydrogenase and interleukin-2 receptor levels, was normal. An excisional biopsy was performed. After the lymph node biopsy, he received rituximab therapy.

The biopsied specimen was 2.0 cm in diameter. In the low-power field, the lymphoid sinuses were absent (Fig. 1). Numerous lymphoid follicles were present throughout the cortex and medulla (Fig. 1). In the medium-power field, small atrophic germinal centers were surrounded by a broad mantle zone composed of concentric rings (Fig. 2a). Moreover, some of the lymphoid follicles contained more than one atrophic germinal center (Fig. 2a). Only a few germinal centers were penetrated by a sclerotic blood vessel (Fig. 2a). In the high-power field in small germinal centers, germinal center lymphocytes decreased in number and some of the follicular dendritic cells (FDCs) exhibited nuclear pleomorphism (Fig. 2b). In the interfollicular area, there were numerous high endothelial vessels with plump endothelial cells and sclerotic walls (Fig. 2a); small foci of plasmacytoid dendritic cells were also present (Fig. 2c). CD21 immunostaining demonstrated FDC networks with tight/concentric and expanded/disrupted patterns (Fig. 2d) in some of the lymphoid follicles.

Flow cytometry data from the biopsied specimen revealed a polyclonal B-cell population. However, conventional G-banded karyotype analysis demonstrated 46, X,-Y, +18, i(18)(q10) in 8 of the 20 cells examined (Fig. 3). A clonal band was observed by Southern blot analysis of the immunoglobulin heavy (IgH) chain gene (data not shown).

The immunohistochemical studies were re-examined. In a portion of the lymph node (Fig. 1, ring), aggregates of small lymphocytes (Fig. 4a) were positive for CD20 (Fig. 4b), CD43 (Fig. 4c), and bcl-2, but negative for CD3, CD5 (Fig. 4d), CD10, bcl-6, and CyclinD1.

In 1956, Castleman et al. described an entity involving localized mediastinal lymph node hyperplasia resembling thymoma. Since this original description, CD has been extended to include two entities: the classic HV type and a rare plasma cell (PC) type. The present case had characteristic histomorphological and immunohistochemical findings of HV type CD: (i) absence of lymphoid sinuses in the lesion, (ii) presence of abnormal lymphoid follicles, (iii) nuclear pleomorphism of FDCs, (iv) interfollicular vascularity, (v) presence of plasmacytoid dendritic cell clusters, and (vi) abnormal proliferation of FDCs. However, karyotype analysis demonstrated +18, which is one of the common clonal cytogenetic abnormalities of nodal marginal zone B-cell lymphoma (NMZBL). Southern blot analysis of the immunoglobulin heavy (IgH) chain gene also revealed a clonal band. CD20+ CD43+ aggregates in small lymphocytes in a portion of the lymph node were observed on immunohistochemical studies. Aberrant CD43+ expression by B cells suggested NMZBL. The etiology of HV CD remains unknown. Some authors have suggested that HV CD may be a FDC tumor. Indeed, clonal cytogenetic abnormalities in the FDCs in HV CD have been reported. Furthermore, HV CD and FDC sarcomas were found to be associated. However, the cytogenetic, molecular, and immunohistochemical findings indicate that this case may have been early stage NMZBL. Several malignant B-cell lymphomas have HV-CD-like morphology, including follicular lymphoma, mantle cell lymphoma, NMZBL, and diffuse large cell lymphoma. This case suggests that NMZBL may exhibit the same histomorphological findings as HV CD.

CONFLICT OF INTEREST

The authors declare no conflict of interest regarding this study.
Fig. 1. Numerous lymphoid follicles were seen throughout the cortex and medulla. Note the absence of lymphoid sinuses. The ring indicates aggregates of CD43-positive small lymphocytes. HE

Fig. 2. a. In the medium-power field, small atrophic germinal centers were surrounded by a broad mantle zone composed of concentric rings. Note that the lymphoid follicle contained two atrophic germinal centers. The germinal center was penetrated by a sclerotic blood vessel. Numerous high endothelial vessels exhibited plump endothelial cells and sclerotic walls in the interfollicular area. HE  
   b. In the high power field, note the decrease in germinal center cells and nuclear pleomorphism of FDCs. HE  
   c. Note the foci of plasmacytoid dendritic cells in the interfollicular area. Giemsa  
   d. CD21 immunostaining demonstrated the proliferation of FDCs (expanded/disrupted pattern).

Fig. 3. G-banded karyotype analysis demonstrated +18 (arrows).

Fig. 4. a. In a portion of the lymph node, aggregates of small lymphocytes without cytological atypia were observed. They were positive for CD20 (b) and CD43 (c) but negative for CD5 (d).
REFERENCES


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