**Rhodococcus equi in the Soil Environment of Horses in Inner Mongolia, China**

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**ABSTRACT.** Little is known about the distribution of *Rhodococcus equi* in the soil environment of native horses in China. One hundred and eight soil samples were collected from native-horse farms in the Hulun Beier grasslands of eastern Mongolia, the Xilin Goler grasslands of southern Mongolia, and Tongliao City in Inner Mongolia, China. The isolation rates of *R. equi* from soil samples from the Hulun Beier and Xilin Goler grasslands ranged from 25.9% to 30.0%. In contrast, isolation rates from soil samples from Tongliao City were as high as 82.3% and the mean number of *R. equi* in soil samples from Tongliao City was 10 times more than those of samples from the grasslands. The 488 isolates were examined using PCR for the presence of genes that encode virulence-associated 15–17 kDa antigen protein (VapA) and the 20 kDa antigen protein (VapB). All isolates were negative for virulence-associated proteins. Plasmid profiles of these avirulent isolates showed that cryptic plasmids of various sizes were present with an incidence of 13.3% to 21.5%. The results of the present study contrast with those of our recent study (J. Vet. Med. Sci. 67(7): 739–742, 2006), in which we reported that *R. equi* was absent from Mongolian horses in Ulaanbaatar, Mongolia. It is suggested that the difference between the results of these two studies is due to the mobile pasturing system in Mongolia and nonmobile pasturing system in Inner Mongolia.

**KEY WORDS:** Inner Mongolia, native horse, *Rhodococcus equi.*

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*Rhodococcus equi* is one of the most important causes of pneumonia of foals at the age of one to three months [1, 2]. *R. equi*, a soil organism, is ingested by many herbivores and has a worldwide distribution, especially in the feces and environment of horses [1, 2, 13]. The primary route of infection in foals is believed to be inhalation of virulent *R. equi* present in the foals’ environment. Virulence of *R. equi* has been associated with the presence of a virulence plasmid that encodes a pathogenicity island consisting of several virulence-associated proteins (Vaps) [8]. A variety of virulence plasmids have been identified in virulent isolates throughout the world based on size and distinct restriction endonuclease digestion patterns [4, 11, 12, 14, 16, 17, 20]. There are at least 12 virulence plasmids associated with *R. equi*, and the plasmid type is related to the geographic origin of the strain [4].

Based on our molecular epidemiologic studies on virulence plasmids in Japanese native horses and horses of other breeds, we hypothesized that the transmission of the virulent form of *R. equi* and the associated virulence plasmids to Japan may have occurred through the migration of native horses from the Mongolian grasslands via China and the Korean Peninsula [12]. The presence of a 90 kb type-II plasmid in Korean and Japanese isolates also partially supports a common origin and ancestry of the native horses of northeast Asia [17]. More recently, we investigated soil samples and the feces of foals raised by nomadic families camped in three areas less than 100 km from Ulaanbaatar, Mongolia [15]. However, no *R. equi* was isolated from the feces of Mongolian native horses. In the present study, we focused on the soil environment of horses in Inner Mongolia, China, which is adjacent to Mongolia, to investigate the presence of virulent *R. equi*.

One hundred and eight soil samples were collected from native horse farms in the Hulun Beier grasslands of eastern Mongolia, the Xilin Goler grasslands of southern Mongolia, and Tongliao city, which is in Inner Mongolia, China (Fig. 1). Of the 108 samples, 34 were collected from farms in Hailar and 30 were collected from farms in Xin Bulag Dong, the eastern and western regions of the Hulun Beier grasslands, respectively; 27 were collected from farms in the eastern region of the Xilin Goler grasslands; 17 were collected from farms in Tongliao city. According to a field survey of local veterinarians, the disease was not recognized, as there has been no outbreak of *R. equi* infection in foals.

A small spoon was used to scrape each soil sample from the surface of the ground. Each sample was placed in a sterile tube. At the laboratory of the Department of Veterinary Medicine, Changchun University of Agriculture and Animal Sciences, 1 g of the soil was serially diluted with a 10-fold volume of sterile saline. Each dilution was inoculated onto two plates of nalidixic-acid-novobiocin-acidione (cycloheximide)-potassium tellurite (NANAT) medium as described by Woolcock et al. [21]. The plates were incu-
bated at 30°C for two or three days. The *R. equi* colonies were counted, and the numbers of viable organisms per gram of feces or soil were estimated. Suspected colonies of *R. equi* were subcultured and examined for the vapA and vapB genes using the polymerase chain reaction (PCR). The target DNAs for PCR amplification were the published sequences of the 15–17 kDa antigen (VapA) gene and a 20 kDa antigen (VapB) gene (Genbank database accession numbers D21236l and D44469, respectively) from *R. equi* strains ATCC 33701 and 5, respectively [6, 10]. Primer 1 (5'-GACTCTTCACAAGACGGT-3') corresponded to the sense strand at position 6–23 in the sequence of the 15–17 kDa antigen gene, and primer 2 (5'-TAGGCGTTGTGC-5') corresponded to the antisense strand at position 569–552 [6]. Primer 3 (5'-AACGTAGTCGCGGTGAGAA-3') corresponded to the sense strand at position 240–258 in the sequence of the cloned fragment containing the 20 kDa antigen gene, and primer 4 (5'-ACCGAGACTTGAGCGACTA-3') corresponded to the antisense strand at position 1066–1048 [10]. PCR amplification was performed using previously described methods [9, 19]. Plasmid DNA was isolated from *R. equi* using the alkaline lysis method with some modifications, as described previously [20]. Samples of the plasmid preparations were separated on 0.7% agarose gels at approximately 5 V/cm for 2 hr.

Of the Hulun Beier grassland sites, *R. equi* was isolated from five of seven farms at Hailar (isolation rates: 20–50%; 25–525 colony forming units (CFU) per gram) and from three of six farms at Xin Bulag Dong (isolation rates: 40–80%; 25–500 CFU/g). *R. equi* was isolated from three of four farms at Huolinguole in the Xilin Goler grassland (isolation rates: 12.5–83.3%; 50–1650 CFU/g) and from four of four farms at Tongliao City (isolation rates: 80–100%; 50–8050 CFU/g). The isolation rates of *R. equi* from the grasslands of eastern Hulun Beier, western Hulun Beier, and Xilin Goler were 29.4%, 30.0%, and 25.9%, respectively (Table 1). In contrast, the isolation rate of samples from Tongliao City was as high as 82.3%, and the mean number of *R. equi* in soil samples from Tongliao City was 10 times greater than those from the grasslands.

As shown in Table 2, 488 isolates were examined using PCR for presence of the genes that encode virulence-associated 15–17 kDa antigen protein (VapA) and the 20 kDa antigen protein (VapB), and all isolates were negative for these proteins. Plasmid profiles of the 488 isolates revealed the presence of cryptic plasmids of various sizes with an incidence of 13.3–21.5%.

To test the hypothesis that the transmission to Japan of the virulent form of *R. equi* and its associated virulence plasmids may have occurred with the migration of native horses from the Mongolian grasslands via China and the Korean Peninsula, we sampled the environments of Cheju horses in Korea and Mongolian horses at Ulaanbaatar in Mongolia for isolation of *R. equi* [12, 15, 17]. Our recent study showed no evidence of *R. equi* in either fecal samples of Mongolian foals or soil samples collected from the environments in which they were held by 26 nomad families in Ulaanbaatar. Avirulent *R. equi* were isolated only from the feces of descendants of introduced Przewalski's Horses at Hustai National Park [15]. In the present study, we were able to isolate *R. equi* from soil environment of horses at the Hulun Beier and Xilin Goler grasslands of Inner Mongolia, but no virulent *R. equi* were found in those isolates. Moreover, the isolation rates and the mean number of *R. equi* from Tongliao City were significantly greater than those from the grasslands of Inner Mongolia. These data contrast with those of our recent study in which *R. equi* was absent from Mongolian horses in Ulaanbaatar, Mongolia [15].

*R. equi* is a soil saprophyte that is most commonly found in the superficial soil layer at concentrations of up to $10^4$

![Fig. 1. Location of sites where soil samples were collected in Inner Mongolia.](image-url)
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CFU/gram at horse-breeding farms in Japan and many countries where R. equi infestation occurs every year [1–3, 7, 13, 22]. The soil concentration of R. equi is highest in areas where horses graze and increases with the length of time that the pasture has been grazed by horses [3, 7, 22]. Traditionally, Mongols are pastoral nomads who move with their animals in a predictable pattern according to the seasons and the availability of forage. They never camp at the exactly same place as before. With the introduction of post-Maoist economic reforms, herders no longer have customary rights to land in Inner Mongolia, China [5]. Instead, the herding families are allowed small areas of pasture called kulums (enclosures). A typical family in Inner Mongolia has a mixed herd of 75–200 sheep, 2–40 cattle, 2–60 horses and 3 Bactrian camels [5]. We speculate that the difference between the isolation of R. equi reported in this study and that of our previous study is related to the mobile vs. nonmo-

Table 1. Isolation of Rhodococcus equi from soil in the environments of horse breeding areas in Inner Mongolia, China

<table>
<thead>
<tr>
<th>Area</th>
<th>No. of farms</th>
<th>No. of soil samples</th>
<th>No. of positive samples</th>
<th>Isolation rate (%)</th>
<th>No. bacteria per g soil (Mean ± SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Eastern region of Hulun Beier Grasslands (Hailar)</td>
<td>7</td>
<td>34</td>
<td>10</td>
<td>29.4^a</td>
<td>135 ± 183^a</td>
</tr>
<tr>
<td>Western region of Hulun Beier Grasslands (Xin Bulag Dong)</td>
<td>6</td>
<td>30</td>
<td>9</td>
<td>30.0^b</td>
<td>103 ± 150^b</td>
</tr>
<tr>
<td>Eastern region of Xilin Golier Grasslands (Huolinguole)</td>
<td>4</td>
<td>27</td>
<td>7</td>
<td>25.9^c</td>
<td>361 ± 588^c</td>
</tr>
<tr>
<td>Tongliao City</td>
<td>4</td>
<td>17</td>
<td>14</td>
<td>82.3</td>
<td>1645 ± 2190</td>
</tr>
<tr>
<td>Total</td>
<td>21</td>
<td>108</td>
<td>40</td>
<td>37.0</td>
<td>696 ± 1473</td>
</tr>
</tbody>
</table>

^a) P<0.01 compared with the percentage for Tongliao City (χ² test).
^b) P<0.05 compared with the percentage for Tongliao City (χ² test).
^c) P<0.05 compared with the mean for Tongliao City (Student’s t-test).
^d) P>0.05 compared with the mean for Tongliao City (Student’s t-test).

Table 2. Identification of virulence plasmids in isolates from Inner Mongolia

<table>
<thead>
<tr>
<th>Area</th>
<th>No. of isolates tested</th>
<th>PCR VapA</th>
<th>VapB</th>
<th>Cryptic plasmids (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Eastern region of Hulun Beier Grasslands</td>
<td>58</td>
<td>0</td>
<td>0</td>
<td>8 (13.8)^a</td>
</tr>
<tr>
<td>Western region of Hulun Beier Grasslands</td>
<td>29</td>
<td>0</td>
<td>0</td>
<td>4 (13.8)^a</td>
</tr>
<tr>
<td>Eastern region of Xilin Golier Grasslands (Huolinguole)</td>
<td>113</td>
<td>0</td>
<td>0</td>
<td>15 (13.3)^b</td>
</tr>
<tr>
<td>Tongliao City</td>
<td>288</td>
<td>0</td>
<td>0</td>
<td>62 (21.5)</td>
</tr>
<tr>
<td>Total</td>
<td>488</td>
<td>0</td>
<td>0</td>
<td>89 (18.2)</td>
</tr>
</tbody>
</table>

^a) P>0.05 compared with the percentage for Tongliao City (χ² test).

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