Isolation of Maedi/Visna Virus from a Sheep in Japan

Keisuke OGUMA1), Chiaki TANAKA1), Ryo HARASAWA2), Atsushi KIMURA3), Jun SASAKI4), Masanobu GORYO4) and Hiroshi SENTSUI1)*

1)Laboratory of Veterinary Epizootiology, Department of Veterinary Medicine, Nihon University, 1866 Kameino, Fujisawa, Kanagawa 252–0880, Japan
2)Laboratory of Veterinary Microbiology, Department of Veterinary Medicine, Faculty of Agriculture, Iwate University, 3–18–8 Ueda, Morioka, Iwate 020–8550, Japan
3)Veterinary Clinics of NOSAI Morioka, 28–2 Hosoda, Kamiohta, Morioka, Iwate 020–0053, Japan
4)Laboratory of Veterinary Pathology, Department of Veterinary Medicine, Faculty of Agriculture, Iwate University, 3–18–8 Ueda, Morioka, Iwate 020–8550, Japan

(Received 25 May 2013/Accepted 4 October 2013/Published online in J-STAGE 18 October 2013)

ABSTRACT. Maedi/visna (MV) is a lentiviral disease of sheep caused by the maedi/visna virus (MVV). Although MV is prevalent in many countries, it had not been reported in Japan. In 2011, however, three sheep in northern Japan were reported to be seropositive against the MVV antigen, indicating a persistent MVV infection. In the present study, we isolated MVV from one sheep to confirm MVV infection and conduct genomic classification of the virus. The co-culture of leukocytes from a seropositive sheep with fetal goat lung cells resulted in the formation of syncytial cells and the amplification of a long terminal repeat sequence of MVV by polymerase chain reaction. The isolate was confirmed as being MVV, rather than the caprine arthritis-encephalitis virus based on phylogenetic analysis of the gag gene sequence. Although the sheep was asymptomatic, nonpurulent meningitis and demyelination were found in the spinal cord. These were considered to be early lesions associated with pathogenic MVV infection. Therefore, the present study demonstrated that MVV is distributed in Japan.

KEY WORDS: maedi/visna, PCR, retrovirus, sheep, virus disease.

©2014 The Japanese Society of Veterinary Science

Maedi/visna (MV) is a lentiviral disease caused by the maedi/visna virus (MVV) that mainly affects domestic sheep. Infected sheep develop fatal and progressive pneumonia and encephalomyelitis after a latent period of several months to years [7, 12, 13]. Because most MVV infected sheep produce a specific antibody that persists for life, MVV infection can be diagnosed serologically by the agar gel immunodiffusion (AGID) test and enzyme-linked immunosorbent assay tests [5]. These sheep were asymptomatic, but the presence of an antibody indicated a persistent MVV infection. Therefore, in the present study, we tried to isolate MVV from one of the sheep on a farm in Iwate Prefecture, in order to confirm the infection and conduct genomic classification of the virus.

MATERIALS AND METHODS

Virus isolation: Peripheral blood was obtained from a Cheviot sheep (estimated to be 13 years old) that had already been diagnosed as seropositive against the MVV antigen [5]. The blood was mixed with an equal volume of 0.83% ammonium chloride to lyse erythrocytes for the isolation of whole leukocytes. The isolated leukocytes were then co-cultured with the primary cultured cells of fetal lamb lung (FLL) (passaged 10 times) and fetal goat lung (FGL) (passaged 2 times) in Eagle’s minimum essential medium supplemented with 10% fetal bovine serum, 100 unit/ml of penicillin, 100 μg/ml of streptomycin and 5 μg/ml of amphotericin B. The FLL and FGL cells were passaged approximately every five days. These cultured cells were subjected to Giemsa staining for the detection of multinucleated giant cells and to genomic DNA extraction for polymerase chain reaction (PCR), in order to analyze the integration and propagation of MVV.

PCR: PCR was performed to amplify a partial sequence of a long terminal repeat (LTR) and a complete sequence of the gag gene of MVV. The templates used for PCR were the genomic DNA extracted from the peripheral blood leukocytes of the seropositive sheep, those from the FLL or FGL cells co-cultured with infected sheep leukocytes or those from tissues taken at euthanasia of the sheep. The tissue samples
included the cerebrum, lung, spleen, turbinate and lymph nodes adjacent to the lung, intestine, spleen and spinal cord.

DNA was extracted using a Gentra Puregene Kit (QIA-GEN, Hilden, Germany) according to the manufacturer’s instructions. PCR was performed using a GoTaq Green Master Mix (Promega, Madison, WI, U.S.A.). Table 1 lists the primers used to amplify LTR and the gag gene, which were added at 0.5 µM. The primer pair used for LTR has been previously reported [1]. The amplification of LTR was performed as follows: initial denaturation at 94°C for two min, followed by 35 cycles at 94°C for 30 sec, at 58°C for 30 sec and then at 72°C for 30 sec. The gag gene was amplified as follows: initial denaturation at 94°C for two min, followed by 35 cycles at 94°C for 30 sec, at 52°C for 30 sec and then at 72°C for 1.5 min. The MVV positive control was kindly provided by Prof. Misao Onuma at Hokkaido University. Triplicate PCRs were performed for sequence determination to exclude PCR-generated errors. The amplified products were cloned into the pCR2.1-TOPO vector (Life Technologies, Gaithersburg, MD, U.S.A.). The LTR and gag gene sequences were analyzed using a BigDye Terminator v3.1 Cycle Sequencing Kit with an Applied Biosystems 3130 Genetic Analyzer (Life Technologies).

**Phylogenetic analysis of the virus**: Phylogenetic analysis was performed using Clustal W in MEGA5 software [21]. A phylogenetic tree based on gag gene nucleotide sequences was generated using the Neighbor-Joining method with a p-distance model. The analysis included MVV strain 1514 as the reference strain (M60610) [20], 85/34 (AY101611) [10], P1OLV (AF479638) [2], SA-OMVV (M31646) [16] and EV1 (S51392) [18]. The caprine arthritis-encephalitis virus (CAEV) strains included in the study were Cork as the reference strain (M33677) [17], Ov496 (FJ195346) [6], Gansu (AY900630) and No. 40, which was isolated from a goat in Japan [8]. The gag gene of equine infectious anemia virus (EIAV) (AF247394) was used as the outgroup.

**AGID**: The serum sample taken from the sheep used in the present study was analyzed by the AGID test to confirm the animal’s seropositivity against the MVV antigen. The reference MVV antigen and antiserum were provided from National Institute of Animal Health (Tsukuba, Japan). In addition, the antigen of the present isolate was prepared to confirm its antigenicity against the reference MVV antisera. Briefly, virus-infected FGL cells were cultured in tissue culture flasks and passaged every five days. The culture fluids collected from each passage were pooled and concentrated 100 times using ammonium sulfate, as previously described for the preparation of bovine leukemia virus antigen for the AGID test [9].

**Histopathology and immunostaining**: The sheep used in the present study was humanely sacrificed for pathological analysis. Tissue samples were fixed in 10% formalin and embedded in paraffin wax. Sections (4 µm) were stained with hematoxylin and eosin (HE), along with Klüver-Barrera staining for histological examination. Immunostaining was performed using the antibodies for MVV (1:100, VMRD Inc., Pullman, WA, U.S.A.), CD3 (1:50, Dako, Glostrup, Denmark) and CD20 (1:2,000, Thermo, Waltham, MA, U.S.A.).

### Table 1. Sequence of the primers used for LTR and gag gene amplification

<table>
<thead>
<tr>
<th>Name</th>
<th>Sequence (5’→3’)</th>
</tr>
</thead>
<tbody>
<tr>
<td>LTR 2s</td>
<td>CAGAAATCATAGTCAGGATGACAC</td>
</tr>
<tr>
<td>LTR 2a</td>
<td>CCACGTTGGGCGCCAGCTGCGAGA</td>
</tr>
<tr>
<td>Gag-F</td>
<td>AACTCCGGGAGCCCGTGAAG</td>
</tr>
<tr>
<td>Gag-Rab</td>
<td>WTCCCCATTTCYCTCCTCTA</td>
</tr>
</tbody>
</table>

a) W: A+T, b) Y: C+T.

### RESULTS

**Virus isolation from a seropositive sheep**: Although the presence of MVV in peripheral blood leukocytes and tissues from the MVV-seropositive sheep was investigated by genomic PCR, the amplification of LTR was not observed. However, the FGL cells cultivated with the leukocytes formed numerous multinucleated giant cells after the fourth passage (Fig. 1). In contrast, FLL cells did not show any syncytial cells. The MVV LTR fragment was amplified by genomic PCR from the FGL cells forming multinucleated giant cells (Fig. 2), and a 253-bp long sequence without 48 bp of primer sequences was determined (AB821356). The ampricon of the positive control was 346-bp long (data not shown).

---

**Fig. 1.** Representative syncytium (arrow) formed in FGL cells. The FGL cells co-cultured with leukocytes from a MVV-seropositive sheep were passaged four times and stained with Giemsa.

**Fig. 2.** Amplification of LTR fragments by genomic PCR. DNA from FGL cells passaged four times (P4) produced a band representing a 301 bp fragment, whereas DNA from seropositive sheep leukocytes before co-culture (Pre) did not undergo amplification. The positive control (P.C.) sample was 346-bp long. Autoclaved double-distilled water was used for negative control (N.C.). M: DNA marker.
Fig. 3. Multiple alignment of the nucleotide (A, this page) or amino acid (B, next page) sequences of gag genes. (A) MVV-1514 and CAEV-Co (CAEV-Co) were used for nucleotide comparison as the MVV and CAEV reference strains, respectively. Nine viruses including our isolate were compared in their amino acid sequences. Nucleotides or amino acids identical to the present isolate are shown as dots. Gaps are indicated by dashes.
Fig. 3. Continued.
shown). The shorter LTR length in the present isolate mainly resulted from the lack of a 36 bp sequence in the U3 and a 11 bp sequence in the R region, in addition to sparse single nucleotide insertions and deletions.

**Gag gene sequence of the isolated virus:** The complete gag gene sequence of the isolated virus was found to be 1,308-bp long and encoded 435 amino acid residues (AB818536). When compared with the MVV and CAEV reference strains, the nucleotide sequence showed 82.8 and 72.0% homology with MVV strain 1514 and the CAEV strain Cork, respectively (Fig. 3A). The homology of the gag protein N-terminal region was higher to MVV strains than to CAEV strains (Fig. 3B). However, a 21 bp sequence between nt 403 and nt 423 of MVV-1514 was lacking in the isolate. The seven corresponding amino acid residues were also found in the other four MVV strains, but not in all three CAEV strains. The homology with the reported partial gag gene of CAEV-No. 40 was 84.5% (Fig. 4).

**Phylogenetic analysis of the virus:** Phylogenetic analysis based on the gag gene sequence was performed to confirm that the isolated virus was MVV. The gag sequence obtained was compared with those of viruses that have already been reported as MVVs or CAEVs. The isolated virus was classified as MVV in the phylogenetic tree containing MVV and CAEV clusters (Fig. 5). In addition, the virus was similar to MVV strain 85/34, which was isolated in North America. In contrast, CAEV-No. 40 isolated in Japan was classified in the CAEV cluster, although its available gag gene sequence was as short as 129 bp.

**Confirmation of MVV infection in the seropositive sheep:** The sheep used for virus isolation in the present study was reconfirmed as being seropositive against the MVV antigen by the AGID test (Fig. 6A). In addition, the antigen prepared from the present isolate formed a clear precipitation line and connected with the line produced between the positive reference serum and the reference antigen (Fig. 6B). The prepared antigen, which was the culture fluid concentrated to about 1/100 of the original volume, formed a precipitation line even when the antigen was diluted to eight fold.

**Pathological examination:** The sheep had not shown any clinical manifestations. Gross examination revealed pulmonary atelectasis in a restricted region of the right cranial lobe, but prominent lesions were not observed in the cerebrum, cerebellum and spinal cord. However, mononuclear cell infiltration was identified in many areas of meninges between the cervical and lumbar cords (Fig. 7A and 7B). The infiltrates included CD3 or CD20 positive lymphocytes (Fig. 7C and 7D). The white matter of these segments also sparsely showed demyelination (Fig. 7E), vacuolation and the swelling of astrocytes. The MVV antigen was not detected by immunohistochemistry (data not shown).

**DISCUSSION**

MVV is a slow virus, and the latent period of MV is generally several months to years. Because infection with lentiviruses persists for life, serological surveillance is important to prevent the spread of disease from asymptomatic carrier animals. The three sheep reported in Japan as being seropositive against the MVV antigen were highly likely to be infected persistently with MVV. Therefore, the present study aimed to isolate MVV from one of those sheep.

---

**Fig. 4.** Alignment of the partial gag gene sequences from the isolated MVV (Isolate) and CAEV-No. 40 isolated from a goat in Japan.

**Fig. 5.** Phylogenetic analysis of the isolated virus (Isolate) and other MVV and CAEV strains. The phylogenetic tree was generated using the entire gag gene sequences of previously reported MVVs and CAEVs, except for CAEV-No. 40, the available sequence of which was 129-bp long.
and characterize the virus based on comparisons with other MVV and CAEV strains.

Although MVV and CAEV are now classified as small ruminant lentiviruses (SRLVs), both were considered to be distinct viral species restricted to their respective hosts. Therefore, the viruses isolated from sheep and goats were referred to as MVV and CAEV, respectively [11]. However, MVV in the present study was isolated from a co-culture of peripheral blood leukocytes taken from a seropositive sheep with primary cultured goat cells, but not with sheep cells, probably due to the effect of the low passage times of FGL cells as compared with the high passage times of FLL cells. It has become evident that MVV and CAEV can cross the species barrier [19]. An epidemic of CAEV in goats oc-

Fig. 6. A) Confirmation of MVV-seropositivity of the sheep analyzed in the present study by the AGID test. SS: Serum of the MVV-seropositive sheep. NS: Negative serum (NS1 and NS2). PS: Positive reference antiserum. PA: Positive reference antigen. Phosphate buffered saline (PBS) was used for a negative control reaction. B) Antigenicity of the isolated MVV. The prepared antigen was used without dilution (×1) or was diluted two (×2), four (×4) or eight (×8) times with PBS. The negative control reaction was carried out using PBS. PS: Positive reference antiserum. PA: Positive reference antigen.

Fig. 7. Spinal cord lesions in the MVV-seropositive sheep. (A) Infiltration of mononuclear cells into meninges of the thoracic spinal cord region (HE staining). Higher magnification is shown in (B). The infiltrates include CD3 positive T cells (C) and CD20 positive B cells (D). (E) Demyelinating lesion in white matter (Klüver-Barrera staining).
curred in 2002 on a farm in Nagano Prefecture, where a CAEV strain was isolated and characterized [8]. Thus, one possible source of the isolate’s origin was CAEV that may have spread from goats on the farm to sheep probably related to the sheep used in the present study. However, the gag gene and amino acid sequence of the isolate were closer to that of MVV than to that of the CAEV reference strain and apparently distinct from that of CAEV reported in Japan, although the analyzed partial gag gene sequence of which was as short as 129 bp. Moreover, our phylogenetic analysis revealed that the present isolate belonged to the MVV cluster. These data indicated that the present isolated virus was MVV, but not CAEV. It was also demonstrated that the present MVV (named MVV-Iwate) was similar to MVV strain 85/34 isolated in North America. This suggests that the present isolate might be related to the North American strain. Another point of view is that the present isolate might be a recombinant of MVV and CAEV, which could be attributed to the lack of a 21 bp nucleotide sequence in the isolate and in all three CAEV strains analyzed. The recombination of MVV and CAEV reportedly occurred due to their coinfection or superinfection, resulting in the generation of replication-competent chimerical viruses [15]. However, the sheep used in the present study were not considered simultaneously infected with MVV and CAEV, as the several gag gene clones amplified had a uniform sequence despite PCR using primers that were designed in highly conserved regions among several MVV and CAEV strains. The ancestral virus might have originated from a recombination of MVV and CAEV.

Although the sheep used in the present study had not shown any clinical signs, degenerative and nonpurulent inflammatory lesions were identified in the histological examination of the spinal cord. Benavides et al. classified spinal cord lesions associated with MVV into three patterns: the vascular pattern characterized by perivascular cuffs with minimal lesions in the adjacent neuroparenchyma; the malacic pattern (the most common type) characterized by severe white matter destruction and small numbers of macrophages; and the infiltrative pattern characterized by a severe infiltrate of histiocytes in the parenchyma [3]. However, the present spinal lesions did not match any of these patterns. Because Benavides’s classification is based on the spinal lesions of 12 sheep with clinically recognized neurologic signs, it was considered that the present findings were early lesions that formed during latent MVV infection. The negative result of MVV immunostaining might be attributed to limited viral antigens in the tissues or the reactivity of the primary antibody.

The MVV infection route to the sheep used in the present study remains unknown due to a lack of available information about this animal. The quarantine of imported sheep and goats against SRLVs that entails serological tests had not been performed in Japan before the outbreak of CAEV. Therefore, it was suggested that MVV-infected sheep had been imported prior to the SRLV quarantine and thus became a source of infection. The present study reconfirmed the importance of serological surveys to identify sheep with subclinical but persistent MVV infection. Further surveillance is required to elucidate the current distribution of MVV in Japan.

ACKNOWLEDGMENTS. We are grateful to Dr. Misako Konishi at the National Institute of Animal Health (Tsuchi, Japan) for her help in the AGID tests. This study was supported by a Grant-in-Aid for Scientific Research and a Grant-in Aid for the Academic Frontier Project for Private Universities from the Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan.

REFERENCES


