Sevoflurane is the most unstable modern volatile anesthetic molecule, which can be degraded in dry carbon dioxide (CO₂) absorbent to compound A (CA) (Fluoro-methyl1-2, 2-difluoro-1-(trifluolimethyl)) [19]. CA has been proven to be nephrotoxic in rats after exposures that have varied in duration from 1 to 3 hr [8, 11, 16] and could create a transient dysfunction in the human nephron [27]. In addition, the possibility of hepatic lesion has been suggested [17, 27]. It is preferable that a CO₂ absorbent with less reactivity to the possibility of hepatic lesion has been suggested [17, 27]. In addition, -duration from 1 to 3 hr [8, 11, 16] and could create a transient dysfunction in the human nephron [27]. Yabashi lime® did not produce CA. Control CO₂ absorbent generated CA, and its concentration was significantly higher in low-flow rate than a high-flow rate. CO was generated only in low-flow rate groups, but there was no significance between Yabashi lime® groups and control CO₂ absorbent groups. However, the CO concentration in the circuit could not be detected (≤5ppm), and no change was found in COHb level. Canister temperature was significantly higher in low-flow rate groups than high-flow rate groups. Furthermore, in low-flow rate groups, the lower layer of canister temperature in control CO₂ absorbent group was significantly higher than Yabashi lime® group. CA and CO productions are thought to be related to the composition of CO₂ absorbent, flow rate and canister temperature. Though CO concentration is equal, it might be safer to use Yabashi lime® with sevoflurane anesthesia in dogs than conventional CO₂ absorbent at the point of CA production.

KEY WORDS: canine, compound A, sevoflurane, Yabashi lime®
MATERIALS AND METHODS

Animal model: Four healthy adult dogs (3 females and 1 male beagle), weighing 10–13 kg, were used. Four anesthetic protocols were repeated in each animal with at least one week interval; Yabashi lime® with high-flow rate of oxygen (Group I), Sodasorb® as control CO₂ absorbent with high-flow rate of oxygen (Group II), Yabashi lime® with low-flow rate of oxygen (Group III) and control CO₂ absorbent with low-flow rate of oxygen (Group IV) (Table1). These dogs were treated in accordance with the guideline approved by the Animal Use Committee of Gifu University.

Experimental set-up: Forty-five min before inhalation anesthesia, all dogs were premedicated with atropine 0.04 mg/kg (i.v.). Fifteen min after the premedication, 0.2 mg/kg of butorphanol and 150 µg/kg of midazolam were injected (i.v.) for sedation. General anesthesia was induced by 7 mg/kg of propofol (i.v.). We used semi-closed circle anesthetic apparatus (Compact 15; KIMURA medical instrument CO., LTD., Tokyo, Japan) with a vaporizer (PPVΣ, Penlon, Oxford, U.K.) throughout the study. Immediately before induction of anesthesia, 1 kg of fresh absorbent was placed in the anesthetic canister. The absorbent was discarded after each case. After tracheal intubation, the animals were positioned in left recumbency and administered 100% oxygen at a flow rate of 3 l/min (Groups I and II) or 0.5 l/min (Groups III and IV) with 3 mg/kg/hr continuous infusion of propofol for induction of sevoflurane until starting of inhalation anesthesia. After induction of inhalation anesthesia, 24 G catheter was inserted into a femoral artery, and 1.5 ml of arterial blood was collected. Gas samples for measurement of CA and CO, and arterial blood samples were collected just before starting of inhalation anesthesia and thereafter 1, 2 and 3 hr, and temperature of the canister was recorded just before starting of inhalation and thereafter every 10 min. After every setting and sampling were finished, the dogs were subjected to inhalation anesthesia and were maintained after every setting and sampling were finished, the dogs were subjected to inhalation and thereafter every 10 min. After every setting and sampling were finished, the dogs were subjected to inhalation anesthesia and were maintained with 3.0–3.5% sevoflurane concentration according to the individual status and were breathing spontaneously. The anesthesia was continued with sevoflurane for 3 hr. Heart rate (HR), respiratory rate (RR), inspired CO₂ (CO₂), end tidal CO₂ (ETCO₂), sevoflurane concentrations, blood pressure, body temperature of animals and oxygen saturation (SpO₂) were monitored continuously as usually every 10 min throughout the experiment (BIO-SCOPE AM120, FUKUDA ME, Tokyo, Japan). In addition, arterial oxygen partial pressure (PaO₂) and arterial carbon dioxide partial pressure (PaCO₂) were measured every 1 hr throughout the experiment (i-stat, Fusio Pharmaceutical Industries, Osaka, Japan). The experiments were performed all procedures at room temperature of 25°C.

CA measurement: Sample gas for CA measurement was collected from the inspiratory limb of the circuit just before starting of sevoflurane anesthesia and every hr thereafter. A glass syringe (100 ml) was used for sampling, and a silicon grease (Non-absorbing Grease to Hydro Carbons, GL Sciences, Tokyo, Japan) was used to ensure an airtight seal. Immediately, 100 ml of the gas was transferred to a bottle that was kept under pressure −80 ± MPa. The sample bottles were stored in the icebox, and the samples were analyzed within a week. The concentrations of CA were measured by employing a gas chromatograph (model GC-7AG; Shimazu, Kyoto, Japan) [4]. The gas chromatograph column was 5 m in length and 3.0 mm in internal diameter, and it was filled with 20% diocetyl phthalate and Chromsorb WAW (GL Sciences) with 80/100 mesh. The injection temperature was 130°C, and the column temperature was 110°C. The carrier gas was nitrogen, and the carrier gas flow rate was 42 ml/min. The gas chromatograph was calibrated with standard calibration gas prepared from stock solutions of CA (Marubishi Pharmaceutical Co., Ltd., Osaka, Japan) [4].

CO measurement: For CO measurement, 100 ml of gas sample was aspirated from the inspiratory limb of the circuit to a CO detector tube (1LC, GASTEC, Ayase, Japan). The changing color was developed over 4 min. CO concentration was measured just before starting anesthesia and every hr thereafter.

HbCO measurement: 1.5 ml of arterial blood was collected from the catheter just before beginning of inhalation anesthesia and every hr thereafter. HbCO concentration was examined by using CO-oximeter (OML3; Radiometer, Copenhagen, Denmark).

Measurement of canister temperature: The carbon dioxide absorbent container of the circuit system was equipped with temperature probes in the upper and lower layers of the container, as described in the previous study [15]. Temperature data were continuously recorded every 10 min.

Statistical analysis: Results are presented by means and SD. All measurements, including CA, CO and HbCO concentrations, and increased temperature of the canister were compared by repeated-measures ANOVA. Significance was assigned at P<0.05.

RESULTS

No significant differences were found between the four groups of consecutive monitoring of HR, RR, inspired CO₂, ETCO₂, sevoflurane concentrations, blood pressure, body temperature of animals, SpO₂, PaO₂ and PaCO₂.

CA measurement: Yabashi lime® was totally lacking CA production throughout the experiment. CA production was significantly higher in control CO₂ absorbent groups (Groups II and IV) (P<0.05) at each measurement point (Fig. 1). Moreover, CA production was significantly higher in low-flow rate (Group IV) than in high-flow rate (Group II) in control CO₂ absorbent groups.

CO measurement: There were significant differences of CO production between high-flow rate groups (Groups
I and II) and low-flow rate groups (Groups III and IV) at each measurement point \((P<0.05)\) (Fig. 2). High-flow rate groups (Groups I and II) were totally lacking CO production throughout the experiment. CO production was significantly affected by flow rate, but was not affected by the type of absorbent.

**HbCO measurement:** No significant differences were found between the four groups in consecutive monitoring of the HbCO (Fig. 3).

**Measurement of canister temperature:** In the upper layer of canister temperature, it isn’t shown remarkable changes in the temperature. However, in the lower layer of canister temperature, there were significant differences between high-flow rate groups (Groups I and II) and low-flow rate groups (Groups III and IV) \((P<0.05)\). In addition, between low-flow rate groups (Groups III and IV), there was a significantly higher temperature in control CO2 absorbent than Yabashi lime® \((P<0.05)\) (Fig. 4).

**DISCUSSION**

The main finding of our study is that CA was not detected with Yabashi lime® by using semi-closed circle anesthetic apparatus even with low-flow rate (Group III) sevoflurane. On the other hand, CA was produced by Sodasorb®. CA is formed by the elimination of hydrogen fluoride from sevoflurane, which is initiated by proton abstraction [9]. The presence of strong bases, such as NaOH and KOH, in the CO2 absorbent may be a factor in the dehalogenation of sevoflurane to CA [25]. Sodasorb® is mainly made of Ca(OH)2, but contains a small amount of KOH and NaOH. On the other hand, Yabashi lime® mostly consists of Ca(OH)2 and does not contain any NaOH and KOH. Yabashi lime® therefore does not generate CA (Table 2). Other researchers also sug-
In this study, we found that CO concentration and increased temperature of the canister and CO concentration. In addition, it may be related to the accumulation with a low-flow rate of oxygen [22]. In other words, CO molecules are likely excess gas by carrier gas like CO2 above, CO molecules may also be diluted or exhausted as excess gas by carrier gas, so concentrations of CO2 in the circuit are declined in Yabashi lime® pellet original shape which makes lower density in the canister, and larger surface area than Sodasorb® as control CO2 absorbent in this study (Fig. 5) [26]. It may be easy to discharge heat of canister for its peculiar shape. When we used control CO2 absorbent with low-flow rate of oxygen (Group IV), increased temperature of the lower layer of the canister and concentration of CA is higher than other groups. However, there were no remarkable changes in the upper layer of the canister construction of anesthesia apparatus. It may be because of aspirated air from the animal flow bottom to top, and it touches with a lower layer of the absorbent in first in anesthetic circuit that we used in this experiment. Therefore, the more CO2 molecule is absorbed by the lower layer of the absorbent than the upper. Furthermore, when we used high-flow rate of oxygen (Groups I and II), there were no differences even between groups of the lower layer of the canister. We suggest that in the case of high-flow rate of oxygen, the accumulated CO2 molecules are diluted or exhausted as excess gas by carrier gas, so concentrations of CO2 in the circuit are declined [23]. Similar reason is thought for CA and may be the major reason why CA concentration with low-flow rate of oxygen in using control CO2 absorbent (Group IV) is higher than high-flow rate of oxygen (Group II).

CO is produced by the reaction between CO2 absorbent and the inhaled anesthetic [14]. Factors accelerating CO generation are similar to those that accelerate CA generation [13, 14]. We found that the CO concentration in the circuit is not affected by the component of absorbent, but by the flow rate. However, no differences were found in COHb between any groups of this experiment. As we mentioned above, CO molecules may also be diluted or exhausted as excess gas by carrier gas like CO2 or CA with high-flow rate of oxygen [22]. In other words, CO molecules are likely to accumulate with a low-flow rate of oxygen, so CO concentration is increased. In addition, it may be related to the increased temperature of the canister and CO concentration. In this study, we found that CO concentration and increased temperature of the canister were significantly higher in use of a low-flow rate of oxygen than a high-flow rate. Other researchers suggest that CO production increases in high temperature of the canister [6]. There were no obvious findings that Yabashi lime® is superior to control CO2 absorbent in the point of CO concentration. However, concentrations of CO in 4 groups were extremely low and not up to the level that affects HbCO level.

In the present study, CA and CO were diluted or exhausted as excess gas by carrier gas, because we used semi-closed circuit. CO concentrations are not up to affecting HbCO levels in any groups. However, it is said that sevoflurane generates less CO than isoflurane and desflurane [1, 6]. It is possible that CO concentrations are up to higher levels and affect the HbCO level by using isoflurane and desflurane. We need more studies by using other anesthetic circuits or other inhalation anesthetics.

The toxicity of CA remains controversial. Data from animal and human studies regarding the safety of CA during low-flow sevoflurane anesthesia are insufficient to prove safety [13]. Therefore, the application of absorbents that minimally or not degrade sevoflurane to CA would eliminate any potential hazard from this toxic compound [2]. Our results suggest that Yabashi lime® is safer than conventional CO2 absorbent in the point of not generating CA. More studies about the safety of CO2 absorbent products in dogs undergoing sevoflurane anesthesia are needed.

A limitation of this study was 3 hr setting experiment for measuring of CA generation after sevoflurane anesthesia. In other studies, the experimental setting for measuring CA production was 4 or 5 hr after sevoflurane anesthesia [9, 14]. In fact, the practical cases are forced for prolonged periods of anesthesia. Moreover, the toxicity of CA is defined by the product concentration and time [18]. Therefore, further studies should examine the CA production after a prolonged period of anesthesia.

In conclusion, though the CO concentration is equal, it is safer to use Yabashi lime® with semi-closed anesthetic circuit and sevoflurane than conventional CO2 absorbent that

| Table 2. Chemical composition of the carbon dioxide (CO2) absorbents* (weight%) |
| CO2 absorbent | Ca(OH)2 | KOH | NaOH | H2O |
| Sodasorb® | 89 | 5 | 2.68 | 12–19 |
| Yabashi lime® | 84 | – | – | 16 |

*Values were provided by the respective manufacturers. Sodasorb® (Grace, Epemon, France), Yabashi lime® (Yabashi product, Gifu, Japan). Ca(OH)2=calcium hydroxide, KOH=potassium hydroxide, NaOH=sodium hydroxide.
contains strong alkali like NaOH or KOH in the point of CA concentration.

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REFERENCES


