Prevalence and pattern of thoracolumbar caudal articular process anomalies and intervertebral disk herniations in pugs

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Running head: ARTICULAR PROCESS ANOMALIES IN PUGS
ABSTRACT. Thoracolumbar intervertebral disk herniation (TL-IVDH) with caudal articular process anomalies has been reported in Pugs. It currently remains unclear whether congenital caudal articular process aplasia/hypoplasia predisposes to the development of TL-IVDH. However, there are difficulties in proving the causal relationship between caudal articular process anomalies and TL-IVDH. The aim of this study was to describe the prevalence of TL-IVDH at the vertebral space containing anomalous and normal caudal articular processes in Pugs. Fifty-seven pugs were eligible to be included in this study. Caudal articular process aplasia/hypoplasia affected 52/57 (91.2%) dogs. The caudal articular process anomalies were most frequently located between T10 and T13. Colocalization of caudal articular process aplasia/hypoplasia and TL-IVDH was detected in 11 dogs (19.3%). The prevalence of TL-IVDH at vertebral spaces containing abnormal caudal articular processes was 12.3%, whereas the prevalence of TL-IVDH at vertebral spaces containing normal articular processes was 2.4%. With the increase in the number of vertebrae with caudal articular process anomalies, the prevalence of TL-IVDH also increased. The results of this study suggested the prevalence of caudal articular process anomalies was high in Pugs. The caudal articular process anomalies could be associated with TL-IVDH. A large cohort is needed to prove the causal relationship between caudal articular process anomalies and TL-IVDH.

KEY WORDS: caudal articular process anomalies, intervertebral disk herniation, pug
Articular processes are the projections of the vertebrae that fit with an adjacent vertebra. Caudal and cranial articular processes contribute to stabilization of the spine and restriction of the ventrodorsal and lateral bending [5]. Articular process malformations have been documented in Pugs, Pomeranians, Cavalier King Charles Spaniels, and German Shepherds [11, 16, 21]. Although the precise pathophysiology of these malformations is unknown, failure of ossification due to an insufficient blood supply during their fetal life has been suggested to underlie the condition [4, 18].

Constrictive myelopathy due to thoracolumbar intervertebral disk herniation (TL-IVDH), arachnoid diverticula, and fibroid constructs with caudal articular process anomalies has recently been reported in Pugs [11]. However, there have been difficulties in proving the causal relationship between caudal articular process anomalies and TL-IVDH. The aim of this retrospective study was to describe the prevalence of TL-IVDH at the vertebral spaces containing anomalous and normal caudal articular processes in Pugs.

MATERIALS AND METHODS

Animals

The imaging database of the Animal Medical Center of Gifu University was reviewed to identify Pugs that had undergone computed tomography (CT) between 2011 and 2016. Cases were included in this study if the vertebral region from T10 to L7 or thorax and abdomen were scanned. Magnetic resonance imaging (MRI) was performed in dogs with neurological deficits. Neurological deficits were determined and recorded. Acute presentations were defined as neurological signs progressed within 3 weeks. Chronic presentations were defined as neurological signs presented for > 3 weeks, as described by Moissonnier et al [14]. Exclusion criteria were as
follows: slice thickness of CT scans exceeding 2 mm and/or images of non-diagnostic quality. Dogs with vertebral fracture/luxation and vertebral/spinal tumors were also excluded.

**CT and MRI**

CT images (Asteion Super 4 CT Scanner, Canon Medical Systems, Tochigi, Japan) of each dog were retrieved. The standard protocol used was a contiguous slice thickness of 0.5 – 2.0 mm (120 kV, 40-80 mA), then reconstruction in a low spatial reconstruction algorithm and high spatial resolution. MRI was acquired with 0.4 Tesla scanner (APERTO Lucent Open MRI, Hitachi Healthcare Manufacturing, Kashiwa, Japan). MR images of spinal cord were acquired using T1 weighted sagittal sequences (TE = 13, TR = 400, slice thickness = 3 mm), T1 weighted transverse sequences (TE = 13, TR = 450, slice thickness = 3.5 mm), T2 weighted sagittal sequences (TE = 120, TR = 2500, slice thickness = 3 mm), T2 weighted transverse sequences (TE = 105, TR = 1600, slice thickness = 3.5 mm), and contrast enhanced T1 weighted sagittal and transverse sequences after intravenous injection of 0.1 mmol/kg of gadodiamide hydrate (OMNISCAN, Daiichi-Sankyo, Tokyo, Japan). Images were displayed using an open-source PACS Workstation DICOM viewer (Osirix Imaging Software, Pixmeo, Bernex, Switzerland).

**Image evaluation**

Two observers, who were unaware of the clinical signs and final diagnoses, independently evaluated images in the region from T10 to L7. The following parameters were evaluated: the location and number of caudal articular process anomalies using 3D volume rendering CT images, the occurrence of IVDH defined as the presence of disk material dorsal to the disk space or calcified materials in the vertebral canal using CT images, and occurrence of spondylosis deformans defined as
the presence of ventral osseous proliferation or enthesopathy at the margins of the endplates using CT images. IVDH were classified into disk extrusion and protrusion if MRI was performed. Disk herniations were classified as extrusion if the disks herniated through all layers of the anulus fibrosus and appeared as a focal epidural mass, as described by Besalti et al [3]. Disk herniations were classified as protrusion if the disk protruded in the central direction or to the left and right and appeared as a symmetrical extension of the disk beyond the margin of the intervertebral space. The observers had the option of manipulating images on PACS during evaluation, including but not limited to magnifying and windowing. If the two observers conflicted on their opinion, images were reevaluated together and a consensus was reached.

Each caudal articular process anomaly was classified as follows (Fig. 1): absent process (aplasia), incomplete formation (hypoplasia), and normal process formation [11]. Caudal articular processes were classified as normal if the caudal articular processes crossed the intervertebral space and have a rounded cursive W-shape when viewed from the dorsal side. The vertebrae were classified as vertebrae with abnormal or normal articular processes. Vertebral malformations were also evaluated using the previously reported classification [12]. Vertebral malformations included dorsal hemivertebra, lateral hemivertebra, dorso-lateral hemivertebra, butterfly vertebra, ventral wedge shape vertebra, and lateral wedge shape vertebra.

Statistical analysis

All analyses were performed with a statistical analysis software (Graphpad Prism software, La Jolla, CA, USA). The association between the total number of vertebrae with caudal articular process anomalies and the presence of TL-IVDH were examined with Cochran-Armitage trend test. The association between the prevalence of
TL-IVDH and the articular process anomalies was evaluated by Fisher’s exact test. P<0.05 was considered statistically significant.

RESULTS

Fifty-seven Pugs were eligible for inclusion to be included in this study. They consisted of 27 males and 30 females that were between 7 months old and 14 years old (median, 9 years) and weighed between 5.0 kg and 13.2 kg (median, 7.8 kg). Five dogs were imaged for neurological deficits. Thirty-seven dogs were imaged for thoracic and abdominal metastasis check. Fifteen dogs were imaged for screening of the thorax and abdomen. Thoracolumbar caudal articular process anomalies were found in 52/57 (91.2%) dogs. Caudal articular process anomalies were located at T10 (39 dogs, 68.4%), T11 (46 dogs, 80.7%), T12 (42 dogs, 73.7%), T13 (17 dogs, 29.8%), L1 (1 dog, 1.8%), L3 (1 dog, 1.8%), and L7 (1 dog, 1.8%, Fig. 2). One hundred forty-four out of 147 anomalous caudal articular processes (98.0%) were concentrated in the region between T10 and T13. There was no cranial articular process anomaly at the region from T10 to L7. Nineteen dogs had 6 lumbar vertebrae. No vertebral body malformations were found.

TL-IVDH was detected in 19 dogs. They were between 6 years and 14 years old (median, 10 years). Disk herniations were located at T10-11 in 1 dog (5.3%), T11-12 in 7 dogs (36.8%), T12-13 in 7 dogs (36.8%), T13-L1 in 2 dogs (10.5%), L1-2 in 1 dog (5.3%), L3-4 in 4 dogs (21.1%), L4-5 in 2 dogs (10.5%), and L5-6 in 4 dogs (13.6%, Fig. 2). Four of these dogs had a history of ataxia and paraparesis.

Forty out of 57 dogs (70.2%) had spondylosis deformans. They were located at T10-11 in 5 dogs (12.5%), T11-12 in 8 dogs (20.0%), T12-13 in 12 dogs (30.0%), T13-L1 in 8 dogs (20.0%), L1-2 in 6 dogs (15.0%), L2-3 in 20 dogs (50.0%), L3-4 in...
143 14 dogs (35.0%), L4-5 in 14 dogs (35.0%), L5-6 in 8 dogs (12.5%), and L6-7 in 2
dogs (5.0%, Fig. 2).

Colocalization of caudal articular process aplasia/hypoplasia and TL-IVDH was
detected in 11 dogs (Table 1). Six of these dogs had 2 or more sites of TL-IVDH. Four
out of the 6 dogs with TL-IVDH at multiple sites had histories of neurological signs
including chronic non-painful progressive paraparesis. The mean duration of
neurological signs was 7.5 months (range, 2 to 12 months). MRI was performed in
these dogs with neurological deficits. All the disk herniations of these dogs were
diagnosed with protrusions. Two of these dogs had dorsal constrictive compression
and 3 of these dogs had enlargement of the dorsal subarachnoid space. TL-IVDH
without colocalized caudal articular process anomalies were detected in 8 dogs. These
dogs had no neurological sign. Thirty-three dogs had caudal articular process
anomalies but did not accompany TL-IVDH. Another 5 dogs had neither caudal
articular process anomalies nor TL-IVDH. They were between 7 months and 14 years
old (median, 8.5 years).

The prevalence of TL-IVDH at the vertebral spaces containing anomalous caudal
articular processes and normal caudal articular processes was shown in Tables 2 and 3.
The prevalence of TL-IVDH at the vertebral spaces containing anomalous and normal
caudal articular processes was 12.3% (18/146 vertebrae) and 2.4% (10/424 vertebrae),
respectively. The prevalence of TL-IVDH at the vertebral spaces containing bilateral
and unilateral anomalous caudal articular processes was 15.0% (17/113) and 3.0%
(1/33), respectively. With the increase in the number of vertebrae with caudal articular
process anomalies, the prevalence of TL-IVDH also increased ($P<0.001$, Table 4).
The prevalence of TL-IVDH in dogs with more than 3 vertebrae with articular process
anomalies was 50% (18/36 dogs), and that containing 0-2 vertebrae with articular
process anomalies was 4.7% (1/21 dogs, $P<0.001$).

DISCUSSION

The overall prevalence of caudal articular process anomalies was 91.2% (52/57) in this study. Most anomalous caudal articular processes were concentrated in the region between T10 and T13. Cranial articular process anomaly was not observed in Pugs included in this study. The retrospective nature and the small number of cases in our study precluded the establishment of the exact prevalence of caudal articular process anomalies in the general population of Pugs; however, a large proportion of Pugs seemed to have caudal articular process anomalies which might underlie late onset myelopathies in this specific breed. The prevalence of IVDH in our study was 19.8% (11/57). Most cases received only CT scans in our study, therefore some disk herniations, fibrous hypertrophy and arachnoid diverticula might have been overlooked, possibly accounting for the lower prevalence of TL-IVDH. A report showed that the sensitivity of CT was 88.6% and that of MRI was 98.5% for the diagnosis of TL-IVDH [6].

Postmortem studies showed that the bilateral defects of articular facets were associated with 28% increase in its range of movement, 43% decrease in its strength [20], and 10% increase in its instability [19]. A previous study reported that the congenital absence of the lumbar facet joint triggered IVDH in humans [18]. The prevalence of TL-IVDH at the vertebral spaces containing bilateral anomalous caudal articular processes was higher than that containing unilateral anomalous caudal articular process in this study. This finding was consistent with the results of cadaver study [7]. Hemilaminectomy has proven not to significantly decrease the stability of the spine [7]. Chronic physicomechanical overload and trauma are known to cause
intervertebral disk degeneration [2]. Stress myelography was helpful for diagnosing
dynamic compression due to disk herniation and dorsal compression [1]. It might be
useful for the evaluation of the extent of spinal instability and further corroborate the
roles of caudal articular process anomalies on the development of TL-IVDH. The
majority of TL-IVDH in small breed dogs would be expected to locate between T10
and L2. Therefore, it was difficult to prove the causal relationship between caudal
articular process anomalies and TL-IVDH. A large cohort would be needed to prove
the causal relationship between caudal articular process anomalies and TL-IVDH.

The exact cause of spondylosis deformans is unknown, but is considered to be
associated with degenerative changes in the annulus fibrosus of intervertebral disks
due to chronic mild instability [8, 15]. The frequency of spondylosis deformans in
dogs was previously reported to be the highest in the intervertebral spaces between
L2-3 and L7-S1 [13, 14]. In the present study, L2-3 disk spaces had the highest
frequency of spondylosis deformans. There seemed no relationship between the
prevalence of caudal articular process anomalies and spondylosis deformans.

We targeted the vertebral region between T10 and L7. The cranial and
mid-thoracic regions were excluded because of the presence of intercapital ligaments
that prevent extrusion of the intervertebral disks. Articular processes from T1 to T9
have similar shapes and their orientations are in a corresponding caudoventral
direction [6]. The articular processes of L1-7 have similar shapes and restrict lateral
flexion. The vertebrae from T10 to T13 represent a transitional portion of the spine [5].
The shape and function of articular processes in the region of T10-T13 differ from
those of other regions and additional stabilizing structures of costovertebral
articulations are absent in the caudal thoracic spine [9, 10]. Disk herniations were
quite prevalent at thoracolumbar junction in aged dogs. Therefore, not only the
presence of caudal articular process anomalies, but also other factors could be involved with the occurrence of TL-IVDH.

No vertebral body malformations were found in the vertebral region between T10 and L7. However, recent research indicated that vertebral body malformation was found most commonly between T7 – T9 [17]. Further studies are needed to establish causal relationship between vertebral body malformations and TL-IVDH in dogs.

Our results indicated the prevalence of caudal articular process anomalies was high in Pugs. The caudal articular process anomalies could be associated with TL-IVDH. A large cohort is needed to further prove the causal relationship between caudal articular process anomalies and TL-IVDH.

ACKNOWLEDGEMENT

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CONFLICT OF INTEREST

The authors declare no conflict of interest to this report.

REFERENCES


FIGURE LEGENDS

Fig. 1. A dorsal view of a 3D volume rendering CT image of the thoracolumbar vertebrae of a Pug. The arrowheads denote hypoplasia of the caudal articular process, and arrows denote aplasia of the caudal articular facet. The other articular processes were normal.

Fig. 2. Locations of caudal articular process anomalies, intervertebral disk herniations, and spondylosis deformans in 57 Pugs.
<table>
<thead>
<tr>
<th>Dog</th>
<th>Age (years)</th>
<th>Sex</th>
<th>Weight (kg)</th>
<th>T10</th>
<th>T11</th>
<th>T12</th>
<th>T13</th>
<th>L1</th>
<th>L3</th>
<th>IVDH</th>
<th>Neurological Deficits</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>10</td>
<td>F</td>
<td>8.2</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>T11-12, T12-13</td>
<td>Chronic paraparesis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>12</td>
<td>F</td>
<td>7.1</td>
<td>B: aplasia</td>
<td>L: hypoplasia</td>
<td>R: hypoplasia</td>
<td>L: hypoplasia</td>
<td>T11-12, T12-13</td>
<td>Chronic ataxia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>9</td>
<td>M</td>
<td>7.8</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>T12-13, T13-L1</td>
<td>Chronic ataxia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>10</td>
<td>F</td>
<td>5</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>R: aplasia</td>
<td>B: aplasia</td>
<td>T12-13, T13-L1</td>
<td>Chronic ataxia</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>11</td>
<td>F</td>
<td>6.1</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>L: hypoplasia</td>
<td>R: aplasia</td>
<td>L-1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>10</td>
<td>F</td>
<td>7.7</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>T10-11, T11-12, T12-T13, L-1-4</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>8</td>
<td>F</td>
<td>8.1</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>T11-12</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>9</td>
<td>M</td>
<td>7.7</td>
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<td>B: aplasia</td>
<td>B: hypoplasia</td>
<td></td>
<td>T11-12</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>8</td>
<td>M</td>
<td>9.4</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: hypoplasia</td>
<td></td>
<td>T12-13</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>11</td>
<td>12</td>
<td>M</td>
<td>5.7</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td></td>
<td>T11-12</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>7</td>
<td>M</td>
<td>6.9</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td></td>
<td>L-3-4</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>9</td>
<td>M</td>
<td>11</td>
<td>R: aplasia</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>L-5-6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>14</td>
<td>11</td>
<td>M</td>
<td>7.6</td>
<td>B: hypoplasia</td>
<td>R: hypoplasia</td>
<td>L: hypoplasia</td>
<td>R: hypoplasia</td>
<td>L-5-6</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>15</td>
<td>11</td>
<td>M</td>
<td>8.5</td>
<td>L: Hypoplasia</td>
<td>R: Hypoplasia</td>
<td>R: Hypoplasia</td>
<td>R: Hypoplasia</td>
<td>L-5-6</td>
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<td></td>
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<tr>
<td>16</td>
<td>14</td>
<td>M</td>
<td>8.9</td>
<td>B: aplasia</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>L-4-5, L-5-6</td>
<td></td>
<td></td>
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<tr>
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<td>10</td>
<td>F</td>
<td>6.2</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td></td>
<td>L-3-4</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>18</td>
<td>6</td>
<td>M</td>
<td>5.5</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>L: aplasia</td>
<td>L-5-6</td>
<td></td>
<td></td>
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<tr>
<td>19</td>
<td>6</td>
<td>M</td>
<td>7.8</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td>B: aplasia</td>
<td></td>
<td>L-3-4</td>
<td></td>
<td></td>
<td></td>
</tr>
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</table>

M: male, F: female, B: bilateral, L: left, R: right
<table>
<thead>
<tr>
<th></th>
<th>T10-11</th>
<th>T11-12</th>
<th>T12-13</th>
<th>T13-L1</th>
<th>L1-2</th>
<th>L2-3</th>
<th>L3-4</th>
<th>L4-5</th>
<th>L5-6</th>
<th>L6-7</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Anomalous processes</strong> (n=146)</td>
<td>1/39</td>
<td>7/46</td>
<td>7/42</td>
<td>2/17</td>
<td>1/1</td>
<td>0/0</td>
<td>0/1</td>
<td>0/0</td>
<td>0/0</td>
<td>0/0</td>
<td>18/146 (12.3%)</td>
</tr>
<tr>
<td><strong>Normal processes</strong> (n=424)</td>
<td>0/18</td>
<td>0/11</td>
<td>0/15</td>
<td>0/40</td>
<td>0/56</td>
<td>0/57</td>
<td>4/56</td>
<td>2/57</td>
<td>4/57</td>
<td>0/57</td>
<td>10/424 (2.4%)</td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.0001</td>
<td></td>
<td></td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>

Table 2. The prevalence of IVDH at the vertebral spaces containing anomalous and normal caudal articular processes.
### Table 3. Relationship between the types of caudal articular process anomalies and the prevalence of colocalized IVDH

<table>
<thead>
<tr>
<th>Types of caudal articular process anomalies</th>
<th>Prevalence of IVDH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bilateral aplasia (n=87)</td>
<td>13/87 (14.9%)</td>
</tr>
<tr>
<td>Aplasia and hypoplasia (n=17)</td>
<td>3/17 (17.6%)</td>
</tr>
<tr>
<td>Bilateral hypoplasia (n=9)</td>
<td>1/9 (11.1%)</td>
</tr>
<tr>
<td>Unilateral aplasia (n=16)</td>
<td>1/16 (6.3%)</td>
</tr>
<tr>
<td>Unilateral hypoplasia (n=17)</td>
<td>0/17 (0%)</td>
</tr>
<tr>
<td>Normal (n=424)</td>
<td>10/424 (2.4%)</td>
</tr>
<tr>
<td>Total number of vertebrae with caudal articular process anomalies</td>
<td>Prevalence of IVDH</td>
</tr>
<tr>
<td>---------------------------------------------------------------</td>
<td>-------------------</td>
</tr>
<tr>
<td>0</td>
<td>0/5 (0%)</td>
</tr>
<tr>
<td>1</td>
<td>1/9 (11.1%)</td>
</tr>
<tr>
<td>2</td>
<td>0/7 (0%)</td>
</tr>
<tr>
<td>3</td>
<td>9/23 (39.1%)</td>
</tr>
<tr>
<td>4</td>
<td>8/12 (66.7%)</td>
</tr>
<tr>
<td>5</td>
<td>1/1 (100%)</td>
</tr>
</tbody>
</table>