Novel Technique for Growth Plate Analysis Based on the Superposition of $T_1$- and $T_2$-weighted MR Imaging of Adolescent Wrists

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In this article, a new method of information extraction on the basis of the differentiation of $T_1$- and $T_2$-weighted MR images is proposed. It relies on a technique of superposition of $T_1$- and $T_2$-weighted MR images with use of statistical dominance algorithm. On the basis of implemented image analysis, a reproducible extraction of growth zone of adolescent boys’ wrists is possible.

Keywords: bone age, children, magnetic resonance, statistical dominance algorithm

Introduction

Growth of the long bones is traditionally evaluated by means of radiographic techniques, which are based on differences in the radiolucency of uncalcified epiphyses. Assessment of the maturation process relies on the detection of calcification nuclei and measurement of the thickness of the uncalcified cartilage of the growth plate.¹ This reflects the calcification progress of primary and secondary ossification centers.

The area between the calcified anlages of the primary and secondary nuclei of calcification contains several histologically distinct zones where enchondral ossification occurs.²

Aforementioned zones cannot be visualized using radiographic techniques. As different zones in the growth plate differ according to the degree of calcification and water content, they are magnetically susceptible and can be detected by MRI. Jaramillo et al.³ used an experimental model and histological correlation to show that only $T_1$- and $T_2$-weighted pulse sequences obtained with a 1.5T scanner (GE Optima 360) allowed visualization of those layers as distinct morphological zones. They showed that physeal and epiphyseal cartilage presented distinct signal intensities; this opened the door for studies using signal postprocessing. A novel mathematical approach to signal interpretation—the statistical dominance algorithm (SDA)⁴—has been proposed to be able to delineate and possibly measure the areas showing signal differences.

The aim of this study was to evaluate different areas of the growth of plate using $T_1$- and $T_2$-weighted MR images, by implementing the SDA for the study of human radius in children.

Materials and Methods

This study was conducted in accordance with good medical practice outlines and was approved by the Ethics Committee of Jagiellonian University (permission no. 1072.6120.16.2017), and complied with the Declaration of Helsinki and good medical practice. Informed written consent for participation was obtained from the legal guardians of the examined children. We included the left hand of 10 healthy boys (age range 9–15 years), including two 9 years old, two 11 years old, three 12 years old, and two 15 years old without translation nor rotation between $T_1$-and $T_2$-weighted images.

Presented patients’ data were anonymized and described as $P_{01}$, $P_{02}$, …, $P_{07}$.

MR studies containing uncorrecatable motion artifacts or that were technically imperfect were rejected. A 1.5T system (GE Optima 360, Chicago, IL, USA) with a dedicated four-channel wrist coil were used. $T_1$- and $T_2$-coronal plane sequences were applied. The following parameters were used to create $T_1$-weighted images: slice thickness, 3 mm; TR, 2749 ms; TE, 106 ms; number of averages, 2; spacing, 3.5 mm; echo train length, 23; bandwidth, 97 MHz. The following parameters were used to create $T_2$-weighted images: slice thickness, 3 mm; TR, 2749 ms; TE, 106 ms; number of averages, 2; spacing, 3.5 mm; echo train length, 23; bandwidth, 81 MHz. For the comparison of $T_1$- and $T_2$-weighted images, overlap of their condition of equal matrices had to be met (286 × 286 pixels matrices were used for the study).
Growth plates were analyzed in coronal scans, separately, for the radius and ulna. The layers for the study were selected in the middle of the antero-posterior diameter of the examined epiphysis and therefore represented its widest part. Images were archived using a SIEMENS PACS (SYNGO, Siemens Healthineers, Erlangen, Germany). Anonymized studies were subsequently retrieved for mathematical postprocessing.

As the scanning time was relatively long (approximately 10 min for the whole study) and as the patients were children, an application test was compulsory. Using pixel-by-pixel positioning of the overlaid images, masks were developed that could compensate for the horizontal and vertical movements by a given number of pixels in case of minor movements. In the case of rotation, other transformations were applied.

Different luminance of the $T_1$- and $T_2$-weighted images was problematic. As the images from these sequences look quite similar, it was difficult to identify differences between the images reproducibly. Some of the images showed local changes in brightness, which hamper a uniform visualization of the differences between images, or comparison of different patients. Thus, $T_1$- and $T_2$-weighted images superposition required standardization.

**Input image standardization**

To standardize the superposition of $T_1$- and $T_2$-weighted images, we used a new image processing algorithm, the SDA, and its simplified version intended for image normalization, the statistical dominance transform (SDT). The SDA transformation converts the image from the brightness domain to the domain of statistical information about dependencies of objects in the image. The algorithm essentially aims to determine the number of neighbors for each pixel in an area with a radius $R$.

$$\text{for (} x = N; x < SX - N; x++ \text{)}$$
$$\text{for (} y = N; y < SY - N; y++ \text{)}$$
$$\quad \{$$
$$\quad \quad \text{imgout}[x, y] = 0;$$
$$\quad \quad \text{for (} i = -N; i <= N; i++ \text{)}$$
$$\quad \quad \quad \text{for (} j = -N; j <= N; j++ \text{)}$$
$$\quad \quad \quad \quad \text{if (} i \cdot i + j \cdot j <= R \cdot R \text{)}$$
$$\quad \quad \quad \quad \quad \text{if (} \text{imgin}[x + i, y + j] >= \text{imgin}[x, y] + \text{threshold} \text{)}$$
$$\quad \quad \quad \quad \quad \text{imgout}[x, y]++;$$
$$\quad \}$$

where, $\text{imgin} =$ input image (matrix), $\text{imgout} =$ output image, $SX, SY =$ width and height of input/output image, $R =$ radius of neighborhood, $N =$ size of neighborhood mask (also size of mirror margin used here), $N = \lceil R \rceil$, threshold = the threshold to be checked (especially for noisy images).

The SD transformation does not consider the additional threshold of dominance determination, which is assumed to be zero (SDA threshold = 0).

Such a procedure makes the picture independent of the dynamics of the luminance of the input image. As an example showing this key feature mentioned above, an analysis of the two different one-dimensional profiles could be provided (directional version assumed for simplicity) that have the same course (but not values) of valleys and peaks (Fig. 1A and 1B), thereby producing the same output from the algorithm (Fig. 1C and 1D, respectively).

![Fig. 1 An example of statistical dominance algorithm (SDA) normalization for one-dimensional images. Two different input profiles with the same course of valleys and peaks but different values, for radius $R = 1$ have the same output profiles. Margins were not considered to maintain clarity.](image-url)
For this study, the value of the radius was assumed to be $R = 50$ pixels, because we found that further changes to the radius value did not result in significant changes. Radius selection is discussed in Fig. 6d in Nurzynska et al.\textsuperscript{6}

Using the algorithm, the following goals were achieved:

(a) Input images are analyzed in a uniform way; irrespective of whether they differed in dynamics or resolution (e.g., 8 vs. 12 bit), important features are presented in the same way, allowing the interpretation to be standardized.

(b) Changes are emphasized, regardless of their brightness, making them easier to distinguish.

(c) Output images are standardized; each image is transferred to a new, strictly defined space, within a given range. Therefore, comparison of images is possible.

MR images subjected to SDA transformation (Fig. 2c and 2d) look very similar to the original images (Fig. 2a and 2b), while possessing features that allow further processing and analysis.

**Superposition of $T_1$- and $T_2$-weighted images**

To determine the resultant image, the difference in images after SDA transformation is calculated mathematically, as described by the following formula (1):

$$D_{out} [x, y] = (SDA(I_{T_1})) [x, y] - (SDA(I_{T_2})) [x, y]$$

where, $D_{out} = output$ matrix with difference values (superposition), $SDA(I_{T_1}) [x, y] = pixel (x, y)$ values of SDA transformation applied on Image ($T_1$), $SDA(I_{T_2}) [x, y] = pixel (x, y)$ values of SDA transformation applied on Image ($T_2$).

The differential image has positive values if the brightness of the objects in the $T_1$-weighted sequence is greater than the brightness of the objects in the $T_2$-weighted sequence; alternatively, the values are negative. A zero value means that both sequences have the same brightness.

Based on superposition outcomes of the transformed images, the following colors can be assigned to the corresponding areas:

- Source color (grayscale) indicates that $T_1$- or $T_2$-weighted images after SDA transformation have a value higher than 50% of the range, otherwise the following applied:
- Yellow indicates that the difference $D [x, y]$ is $>0$. The $T_1$-weighted image after the SDA transformation has a higher value than the $T_2$-weighted image after the transformation.
- Red indicates that the difference $D [x, y]$ is $>0$, and the value of the $T_2$-weighted image after transformation does not reach more than approximately 25% of the range.
- Blue indicates that the difference $D [x, y]$ is $<0$, and the $T_2$-weighted image after the SDA transformation has a higher value than the $T_1$-weighted image after the transformation.

\textbf{Fig. 2} The columns present transformation of $T_1$- and $T_2$-weighted source magnetic resonance images of patient P\textsubscript{04}. (a) and (b) represents input images, $T_1$- and $T_2$-weighted images, respectively. (c) and (d) represents SDA transformation. (e) and (f) expresses colored signal differences based on superposition on $T_1$- and $T_2$-weighted images. (g) and (h) shows surface plots made on the basis of images transformed by applying the SDA. (i) and (j) shows colors representing signal differences overlaid on the surface plot of the normalized source images. SDA, statistical dominance algorithm.
Green indicates that the difference $D(x, y)$ is $<0$, and the value of the $T_1$-weighted image after transformation does not reach more than approximately 25% of the range.

These descriptions are summarized in Table 1.

In the output obtained after image superposition using our proposed method, the zones of $T_1$- and $T_2$-weighted image value dominance were sharply delineated in the growth zones, in a repeatable, reproducible, and objective manners. $T_1$ image value dominance over $T_2$ image value was indicated in yellow and $T_2$ image value dominance over $T_1$ image value was indicated in blue color. The determination of the boundary between these zones of domination is free of the human factor. Based on the size and spacing of pixels in these color zones, the growth zone could be accurately measured. Moreover, subzones of markedly low brightness were considered. Low brightness areas in $T_2$-weighted images were marked in red color, visualized as separate spots on the yellow background (indicating general $T_1$ dominance). Low brightness areas in $T_1$-weighted images were marked in green color, visible as separate areas on the blue background (indicating general $T_2$ dominance). The thresholds for these subzones could be set manually, with a difference of ±3% from the initial threshold of 25%. Therefore, those zones were not used in isolation in growth zone measurement, as they were somewhat operator-dependent.

The obtained images were superimposed with special attention to pixel overlay in corresponding anatomical structures. Differences in the overlaid masks were highlighted where areas of non-mineralized zone with the highest volume of bounded and unbounded water were marked on the growth plate area on the image.

The colored regions obtained were used to determine the growth plate width. Based on pixel size and spacing in the particular image, the surface area was calculated, and the average thickness of the growth zones was computed (pixel spacing of 0.293 mm). The data obtained are included in Table 2.

### Results

Using the SDA, we were able to show repeatedly that the zones in the growth plate of bone could be indicated by different colors (Fig. 3). These differences represent the dominance of the signal arising from the $T_1$ and $T_2$ pulse sequences. As these color differences represent areas with markedly different water content, we assumed that this technique allowed distinction of the distribution of different histological areas in the growth plate (Fig. 3). This idea might be supported by the observation of the color zone distribution in the growth plates of children of different ages (Fig. 4). The wrist of a 9-year-old child presented a significantly broader green area than seen in the wrist of a 12-year-old child. This green area represents the higher $T_2$-weighted image brightness than $T_1$-weighted image, and is indicative of watery cartilage. This area was not present in a 15-year-old subject. The yellow area

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**Table 1** Color projection of the differences in the image brightness recorded in human growth plates

<table>
<thead>
<tr>
<th>Color</th>
<th>$SDA(T_1)$, $SDA(T_2)$ values of the range</th>
<th>$T_1-T_2$ difference</th>
<th>$T_1$ vs $T_2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Source</td>
<td>$SDA(T_1) &gt; 50%$ OR $SDA(T_2) &gt; 50%$</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Yellow</td>
<td>$SDA(T_1) \leq 50%$ AND $SDA(T_2) \leq 50%$</td>
<td>$&gt;0$</td>
<td>$T_1 &gt; T_2$</td>
</tr>
<tr>
<td>Red</td>
<td>$SDA(T_1) \leq 50%$ AND $SDA(T_2) \leq 25%$</td>
<td>$&lt;0$</td>
<td>$T_1 &lt; T_2$</td>
</tr>
<tr>
<td>Blue</td>
<td>$SDA(T_1) \leq 50%$ AND $SDA(T_2) \leq 50%$</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Green</td>
<td>$SDA(T_1) \leq 25%$ AND $SDA(T_2) \leq 50%$</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

SDA, statistical dominance algorithm.
represents a high $T_1$-weighted image area that reflects the presence of a provisional calcification zone, and was broader in 12-year-old patients than in 9-year-old patients. In 15-year-old patients, this area was dominant and reflects the ongoing maturation of the physis (Fig. 4).

Mapping of the different signal intensity distributions across growth plates was reproducible in the group of one 11-year-old, and two 12-year-old subjects, as shown in Fig. 5. The proximal high signal zone, indicated in green, represents area of hydrated tissue with a high signal intensity in $T_2$-weighted images; the yellow distal part represents the tissue of the calcification zone, which contains less water.

The proposed method of normalization allows to compare $T_1$- and $T_2$-weighted images with different

![Fig. 4](image-url)
illuminations (Fig. 6, $T_1$ source values are dominating over $T_2$ source values; seen as a red line of profile). The comparison of different superposition outputs for source images and normalized outputs for SDA transformed images is presented in Fig. 7. It shows that using SDA as normalization method the high reproducibility of ROI pattern was achieved, in opposite to unnormalized source image.

**Discussion**

Radiographic technique based bone age assessment is the most accepted and widely used approaches for estimation of bone age in pediatric populations. Much effort has been expended to develop methods that allow automatic evaluation of radiographic images by means of computerized graphic analysis. The BoneExpert software described by Martin et al.
Novel Technique Based on Superposition of $T_1$- and $T_2$-weighted pulse sequences.

Analysis of uncalcified growth plate width as a whole was implemented with a focus on the chondro-progenitors and proliferating cells, as described by Hochberg. To the best of our knowledge, our work is the first attempt to delineate precisely and directly measure uncalcified growth plate in the basis of the signal differences in the MR pulse sequences.

Our study had some limitations. Histological verification of our finding was not possible in the current model for ethical reasons, but we plan to continue this further in a cadaveric study. Additionally, we used a small group of patients. Nevertheless, this preliminary study illustrated the feasibility of extracting chondrous tissue from MR images with this technique; it should be further verified in a larger group of patients. An error in the segmentation of the growth plate has to be taken into account, given the small area of interest and the averaging effect due to

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**Fig. 6** Profile plots for superposition on source input data and SDA-transformed input data. Red line represents $T_1$ values and blue $T_2$ values. SDA, statistical dominance algorithm.
Fig. 7 Comparison of $T_1$- and $T_2$-weighted image superposition for source images and transformed images for five patients – reproducibility of pattern for normalized images. SDA, statistical dominance algorithm.

different proportions of water, proteoglycans, and collagen in the analyzed tissue. Further studies are needed to evaluate this problem.

Conclusion

Using our method, we were able to develop new indexes describing the size of the uncalcified layer, expressed as ratio of diameter of uncalcified epiphyseal area vs. growth plate area. This approach might be clinically useful, because it utilizes only two sequences, namely $T_1$- and $T_2$-weighted images, which are short and available in all MR systems. The SDA method applied in this study is used in a program that allows reproducible normalization to a standardized threshold across all examined studies as opposed to operations on source data (Fig. 6). This allows the use of MR images for further comparative analyses and extraction of non-calcified areas of different intensity that represent the different areas of the growth plate. This could facilitate detection of lesions in the growth plate by means of MRI.

Ethical Statement

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Informed Consent

Informed consent was obtained from the legal guardians of all individual participants included in the study.

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Conflicts of Interest

The authors declare no conflicts of interest.

References