Cloning and Sequencing of cDNA from *Oryza sativa* encoding a homolog to non-ATPase subunit, MBP1, of 26S Proteasome in *Arabidopsis thaliana*

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In eukaryotes, the intracellular degradation of proteins is a precisely controlled process. This control is ensured partly by an enzyme system capable of selectively marking proteins intended for rapid intracellular degradation by the covalent attachment of ubiquitins, and partly by a multicomponent ATP-dependent protease, the 26S proteasome which clusively degrades ubiquitinated proteins [1-5].

Target specificity within the ubiquitin pathway is determined by two recognition steps [6, 7]. The first selects appropriate substrates for ubiquitination. The second step in defining the specificity of ubiquitin-dependent proteolysis involves the recognition of multiubiquitinated proteins by the 26S proteasome. Substrate selection by the 26S proteasome is presumably mediated by the interaction of specific components of the 19S/PA700 regulatory complex with multiubiquitinated proteins. Although little is known for this essential step, a subunit of the human 19S/PA700 regulatory complex, designated S5a, that can bind ubiquitin-lysosome conjugates *in vitro* was recently identified [6]. The corresponding gene was subsequently identified in *Arabidopsis thaliana* [7]. Sequence analysis revealed that homologs to this gene, designed AtMBP1 for *A. thaliana* multiubiquitin-binding protein, are present in a wide variety of other eukaryotes as well, including *Saccharomyces cerevisiae*, *Drosophila melanogaster*, *Coenorhabditis elegans*, rice and castor beans [7].

Multiubiquitin chain binding is one of the key functions of the 26S proteasomal-selective degradation process. We tried to clone and analyze a full length cDNA encoding a subunit of proteasome, a homologous to *A. thaliana* MBP1, from rice (*Oryza sativa* L., cv. Nipponbare), which is a monocotyledonous plant.

Our large-scale sequencing of randomly chosen rice cDNAs has produced many cDNAs which exhibit significant sequence similarities to previously established sequences [8]. Among them, a cDNA clone from 8-day-old rice seedling mRNAs, *OsS5a* (*Oryza sativa* Subunit 5 a), exhibited strong homology to *A. thaliana* MBP1.

It is likely that the cDNA insert represents the complete coding region of the gene. The clone contains an ORF from an ATG codon located at position 71, until a stop codon at position 1279, followed by a 256-bp 3' untranslated region with a poly(A) tail.

The ORF encoded a protein of 402 amino acids. Using the Smith and Waterman homology search [9], significant homology was observed with potential products of ORFs from *A. thaliana* MBP1 (64.9% identity) [7], human S5a (42.1% identity) [10], *D. melanogaster* S5a (38.3% identity) [11] (**Fig. 1**). This amino acid sequence had a highly conserved N-terminal region and had some characteristic motifs. We detected a nuclear localization signal (NLS) sequence, as well as α -type proteasomal subunits [12]. Recently it was shown that the highly conserved region (residue 213 to 242) with a GVDP motif and a repeated hydrophobic sequence (LALAL/VorL) was necessary for the multiubiquitin chain binding [13, 14]. This region in Rice OsS5a was also highly conserved, and thus it is likely that rice OsS5a functions also as a multiubiquitin binding protein. But rice OsS5a had a DVDP motif instead of the GVDP motif in this region although second GVDP motif (residue 366 to 369) which was not considered as a significant motif was well conserved [13]. It will be required to investigate the ability of multiubiquitin chain binding of rice OsS5a. Rice OsS5a possessed an alanine(A)-rich region starting at residue 200, similar to *A. thaliana* MBP1. It may be a characteristic of plants because neither human nor *D. melanogaster* had this region [10, 11]. The C-terminal region was more poorly conserved than the N-terminal region. But interestingly, it had a KEKE-like motif in the C-terminal region similar to human S5a [10] and the KK at the C-terminus was especially well conserved. These KEKE motifs are present in

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The nucleotide sequence data reported this paper will appear in the DDBJ, EMBL and GenBank nucleotide sequence databases under the following accession number AB010740.

rice OsS5a	MVLEATMICI	DNSEWMRNGD	YSPSRFQAQA	DAVNLICGAK	TQSNPENTVG	50
Arabidopsis MBP1	A..ICI	...EWM....	YS.S.LQA.T	EAV.LL.GA.	TQ.....T..	50
human S5a	S..VCV	...EYM....	FL.T.LQA.Q	DAV.IV.HS.	TR.....N..	50
Drosophila S5a	S..ISF	...DFQ....	YF.T.LIV.R	DGI.LV.LT.	LR.....N..	50
Consensus	MVLE TMIC	DNSE MRNGD	Y P R QAQ	AVNL C K T	SNPEN VG	50
rice OsS5a	VMTMAGKGYR	VLVTPSDLG	KILACMHGLE	VGAEANLAAA	IQVAQLALKH	100
Arabidopsis MBP1	IL.MAGKGYR	..T.P.S.L.	K.LACM.GLD	VGGEINLTAA	.QI.Q.A...	100
human S5a	LI.LA-NDCE	..T.L.P.T.	R.LSKL.TVQ	PKGKITFCTG	.RV.H.A...	99
Drosophila S5a	LM.LS-NTVE	..A.L.S.A.	R.FSKM.LVQ	PKGEINLLTG	.RI.H.V...	99
Consensus	T A V	VL T TSD G	IL MH	E NL	I A LALKH	100
rice OsS5a	RONKRQQQRI	IAFIGSPVKY	DKKVLETIGK	KLKKNVVALD	IVDFGETDDD	150
Arabidopsis MBP1	..N.NQRQ..	IV.A...IKY	EKKA.EIVG.	R...NS.SL.	.VN..EDDDE	150
human S5a	..G.NHKM..	IA.V...VED	NEKD.VKLA.	R...EK.NV.	.IN..EEEVN	149
Drosophila S5a	..G.NHKM..	VV.V...INH	EEGD.VKQA.	R...EK.NV.	.VS..DHGNN	149
Consensus	RQ K	RI I F GSP	K L K	LKK V D	IV FGE	150
			*** **			
rice OsS5a	-KPEKLEALI	SAVNSSDSSH	IVHVPPGENA	LSDVLISTPI	FTGEEGGSGF	199
Arabidopsis MBP1	EKPQKLEALL	TAVNNNDG..	I.H..S.ANA	.S.V.L.T.V	FT.DE.ASGY	200
human S5a	TEKLTAFTNT	LNGKDG.TG.	L.T..P.P-S	.A.A.I.S.I	LA.EG.-AML	197
Drosophila S5a	NEILTAFINA	LNGKDG.TG.	L.S..R.S-V	.S.A.L.S.I	IQ.ED.MGGA	198
Consensus		SH V VP G	LSD L S PI	GE G G		200
rice OsS5a	AASAAAAAAT	GAAGFEFDVD	PNVDPELALA	LRLSMEEERA	RQEAIAKKAA	249
Arabidopsis MBP1	VSAAAAAATA	G-GD.D.G..	.NI.....	.V....E.A	...AAAKK.A	249
human S5a	GLGA-----	--SD.E.G..	.SA.....	.V....Q.Q	...EEARR.A	239
Drosophila S5a	GLGG-----	--NV.E.G..	.NE.....	.V....Q.Q	...SEQRR.N	240
Consensus	A	- FEF VD	PN DPELALA	LR SMEE R	RQE A AA	250
		+++ +	=====	=		
rice OsS5a	EESSGAENKD	HASSSNADSV	MAEAEPASNA	ADDKKDQPK	DDDAQLLQQA	299
Arabidopsis MBP1	DEAGQKDKDG	DTASASQETV	-----AR	TTDKNAEPM-	DEDSAL.DQA	290
human S5a	AASA---AEA	GIATTG----	-----TE---	-----	DSDDAL.KMT	264
Drosophila S5a	PDGA---PPT	GGDAGGGGGV	SGSGPGNEES	AGAENEA---	NTEEAM.QRA	284
Consensus		V			D D LL A	300
rice OsS5a	LAMSMEEGSS	GAAAAADAAMA	EAAVDDQDLA	LALQMSVQDA	GGSSQSDM-S	348
Arabidopsis MBP1	IAMSVGD----	-----VNMS	EADEDQDL.	L.L...MSG	ESSEATGAGN	331
human S5a	IS-QQEFGR-	----TGLPDL	SSMTEEEQI.	Y.M...LQA	EFQGAESADI	308
Drosophila S5a	LALSTETPE-	----DNLPDF	ANMT.EEQI.	F.M...MODA	P-DDSVTQQA	328
Consensus	A S E		A	A QMS Q A		350
rice OsS5a	K-----	-----	VFEDRSFVTS	ILNSLPGVDP	NDPSVKDLLA	379
Arabidopsis MBP1	N-----	-----	LLGNQA.IS.	V.SS.....	NDPAVKELLA	362
human S5a	DASSAMDTSE	PAKEEDDY-D	VMQDPE.LQ.	V.EN.....	NNEAIRNAMG	357
Drosophila S5a	KRPKTDEANA	PMDVDYSE	VIGDPA.LQ.	V.EN.....	OSEAVRDVAG	378
Consensus		- V D F S L	LPGVDP N V			400
			++++			
rice OsS5a	SLHGQGEQ-E	KKEDKSDKPE	DEKK			402
Arabidopsis MBP1	..PDESKRTE	EEESSKKGE	DE..			386
human S5a	..ASQATKDG	KKDKKEE---	-D..			377
Drosophila S5a	..NKDKDK--	----KSDGKD	SQ..			396
Consensus	SL	KS	KK			424

Fig. 1 The alignment of the amino acid sequence of the rice OsS5a subunit with those of equivalent proteasome subunits of other species, *Arabidopsis* MBP1, human S5a, and *Drosophila* S5a subunits. The asterisk (*) shows the nuclear localization signal (NLS). Box I indicates the alanine (A) rich region. A significant region for multiubiquitin chain binding was reported [13, 14]. Box II indicates the corresponding region in rice OsS5a. The crosses and double underlines indicate G(or D) VDP motifs and a hydrophobic region, respectively. Box III indicates the KEKE-like sequence. Dots (.) indicate identical amino acid residues to the corresponding rice sequence, and dashes (-) indicate blanks introduced to achieve maximum similarity.

PA28a, a-type subunits of the 20S proteasome, non-ATPase subunits of the 19S/PA700 complex, and other proteins such as chaperonins. Thus, these regions have been believed to be involved in the formation of protein complexes [15, 16]. Amino acids of rice OsS5a was larger than its homologs of other species. *A. thaliana* MBP1, exhibited the higher homology with rice OsS5a, was 16 residues shorter than the rice OsS5a. Amino acid components of this

product provided isoelectric point (pI) of about 4.2. It is likely that this pI value, like human S5a, was lower than those of other 19S/PA700 regulatory subunits of rice [17].

Genomic Sourthen hybridization under high-strin-gency conditions revealed that the gene for rice OsS5a as well as its homologs was a single copy (Fig. 2) [7, 10, 11]. This gene was deduced to have at least 1 intron, because *Eco*RI digestion caused the formation

Fig. 2 Result of Southern blot analysis of rice genomic DNA probed with the full length *OsS5a* cDNA (about 1.5 kbp). Genomic DNA was digested with the following restriction enzymes: *Bgl* II (lane 1), *Eco*RI (lane 2), *Hind* III (lane 3), *Xba* I (lane 4).

of 2 bands and cDNA for *OsS5a* has no *Eco*RI site. Under low-stringency conditions, several additional faint bands could be detected (data not shown). This result indicates that there are additional sequences similar to this gene in the rice genome.

RNA blot analysis revealed that its mRNA was expressed in both suspension cultures and rice seedlings (**Fig. 3**). The mRNA expression was especially high in suspension cultures that were proliferating rapidly. Ten-day-old seedlings are fully expanded cotyledons. Twenty-day-old seedlings develop true leaves and the cotyledons begin to turn yellow-green. In this 20-day-old seedlings, the mRNA was present at the lowest level. These results suggest that this subunit plays a role in cell proliferation or cell cycle progression.

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Fig. 3 Expression of *OsS5a* mRNA gene of rice during cell culture and seedling growth. Suspension cultures (lane 1) were collected in the logarithmic phase of growth. Seedlings were collected when 10 (lane 2) and 20 days old (lane 3) after sowing at 25°C.

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