A Further Study on the Actions of Phenethylguanidine on the Neuromuscular Junction in Frog Sartorius

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NISHIYAMA, A. A further Study on the Actions of Phenethylguanidine on the Neuromuscular Junction in Frog Sartorius. Tohoku J. exp. Med., 1973, 111 (1), 15-23 — The active phase (end-plate current) of the end-plate potential, and the relationship between the amplitude of the end-plate potential or the end-plate current and the membrane polarization produced by passing current across the muscle membrane were studied in frog sciatic nerve-sartorius treated with PG (phenethylguanidine), which seems to have both competitive and noncompetitive receptor-blocking actions. The falling phase of the end-plate current in PG was slightly but clearly prolonged as compared with that in dTc, a typical competitive receptor blocking agent. The equilibrium potential obtained by the collision experiment (del Castillo and Katz 1954) in PG was found to be at about -15 mV, suggesting that PG did not influence significantly the ratio P_Na/P_K raised by ACh at the end-plate membrane. However, the equilibrium potential was not determined by membrane polarization produced by passing the current across the membrane. Amplitudes of both end-plate potential and end-plate current were increased with muscle membrane depolarization and were decreased with hyperpolarization. Possible factors which may account for these relationships are discussed.

phenethylguanidine; active phase of end-plate potential; relation between end-plate potential or end-plate current and muscle membrane polarization

Electrically the motor end-plate can be described as an electromotive force (equilibrium potential, -15 mV inside) and a series resistance. The mode of action of ACh on the motor end-plate is to decrease the series resistance whereby electrical change is transferred in the inward direction through this shunting resistance causing outward current flow through the adjoining membrane resulting in its depolarization (Fatt and Katz 1951). Takeuchi and Takeuchi (1959) confirmed, using a voltage clamp technique, that the active phase of the frog endplate potential lasts 5 msec during normal transmission, and that the change in series resistance is independent of the membrane potential. Furthermore, Takeuchi and Takeuchi (1960) showed that the series resistance change produced by ACh at the end-plate membrane is due to both sodium and potassium permeability increase, with the ratio P_Na/P_K constant.

These figures mentioned above apply to the curarized muscle, dTc decreases the magnitude of the permeability increase to cations produced by ACh but has no influence on the ratio P_Na/P_K, and therefore also no influence on the equilibri-

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ium potential of the end-plate current or the end-plate potential (Takeuchi and Takeuchi 1960). Another interesting fact about the action of dTc is that dTc shortens the time course, especially the falling part of the active phase (Kuffler 1942; Takeuchi and Takeuchi 1959). In the previous paper (Nishiyama and Kuwabara 1973), PG (phenethylguanidine) was found to act on the end-plate partly by a competitive and partly by a noncompetitive receptor-blocking action. Since the mode of action of PG is somewhat different from that of dTc, it was decided in the present experiments to compare the active phase and the equilibrium potential of the end-plate potential in PG with those in dTc. The experimental results indicate that the active phase of the end-plate potential in PG is slightly longer than that in dTc, and that the end-plate potential in PG is decreased with membrane hyperpolarization and increased with depolarization, and thus the equilibrium potential cannot be extrapolated by this relationship. The explanation for these results has not been resolved.

**METHODS**

The experimental methods for measuring the end-plate potential in frog sartorius muscle were essentially the same as those of our previous report (Nishiyama and Kuwabara 1973).

For the measurement of the active phase of the end-plate potential, the end-plate current was recorded with a modified voltage clamp method similar to that of Takeuchi and Takeuchi (1959). The feed-back amplifier was composed of three stages. The first and second were DC coupled and the final stage was an AC coupled differential amplifier with a time constant of 2.0 sec. This feed-back amplifier was sufficient for the present experiment in which the clamping time was short (about 15 msec). The sciatic nerve was stimulated so that the end-plate current appeared within a few msec after the feed-back circuit was closed. As shown in Table 1, the time course of the end-plate current obtained with this circuit was similar to that of Takeuchi and Takeuchi (1959). Some slight discrepancies might be due to differences in the experimental conditions.

For the determination of the equilibrium potential of the end-plate potential, two different methods were used. In one, the equilibrium potential was determined by collision experiment, i.e., by superimposing the end-plate activity on the repolarization phase of muscle action potential (del Castillo and Katz 1954). In this experiment, the preparation was perfused with a hypertonic saline solution containing 230 mM-NaCl, 2.0 mM-KCl, 5.4 mM-CaCl₂, 0.9 mM-MgCl₂, and 1.0 mM-Na₂HPO₄, to avoid contraction of the muscle fiber during the action potential. In the other, the equilibrium potential was extrapolated from the relationship between the amplitude of the end-plate potential or the end-plate current and membrane polarization (Fatt and Katz 1951; Takeuchi and Takeuchi 1959).

PG was used in the concentrations of $1.0 \times 10^{-4}$ g/ml–$4.0 \times 10^{-4}$ g/ml. These concentrations were rather lower than those in the previous paper (Nishiyama and Kuwabara 1973), and a weak muscle contraction was still observed by neural stimulation in some preparations.

<table>
<thead>
<tr>
<th>Receptor-blocking agent</th>
<th>Rise time</th>
<th>50% fall time</th>
<th>Total time</th>
<th>n</th>
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<tbody>
<tr>
<td>PG</td>
<td>0.8</td>
<td>1.3</td>
<td>5.6</td>
<td>12</td>
</tr>
<tr>
<td>dTc</td>
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<td>0.7</td>
<td>4.2</td>
<td>18</td>
</tr>
<tr>
<td>SCh</td>
<td>0.8</td>
<td>0.9</td>
<td>4.6</td>
<td>7</td>
</tr>
</tbody>
</table>
RESULTS

Active phase of end-plate potential (end-plate current)

The time course of the end-plate current with three different types of receptor-blocking agents is shown in Table 1. The active phase obtained in the curarized muscle at 20–22°C was slightly shorter with respect to the 50% fall time than that at 17°C obtained by Takeuchi and Takeuchi (1959).

Fig. 1. End-plate potentials (upper records) and end-plate currents (lower records) before and after changing the external medium from a solution containing dTc in a concentration of $3.0 \times 10^{-6}$ g/ml to one containing PG in a concentration of $4.0 \times 10^{-5}$ g/ml. A, B, in dTc and C, D, E, F, in PG. The amplitudes of both the end-plate potential and end-plate current declined initially for 2–3 min after changing the solution to the PG one as shown in C and D, but then recovered gradually leading to steady sizes (E, F). Voltage scale in upper record, 10 mV. Current scale in lower record, $10^{-7}$A. Time scale, 2 msec. Resting potential, -80 mV. Ca$^{2+}$, 5.4 mM.

Fig. 2. Time courses of changes in amplitude of end-plate potential (open circle) and end-plate current (filled circle), rise time (open triangle) and 50% fall time (filled triangle) before and after changing the external medium from dTc to PG. The same preparation as in Fig. 1. Note a progressive increase in 50% fall time in PG.
Fig. 3. End-plate potentials (upper records) and end-plate currents (lower records) before and after changing the concentration of dTc from $2.4 \times 10^{-6}$ g/ml to $1.2 \times 10^{-8}$ g/ml. A, B, before and C, D, E, F, after changing the concentration. Note that the time course of each end-plate current was essentially the same nevertheless amplitudes of end-plate potential and end-plate current were different. Voltage scale in upper record, 5 mV. Current scale in lower record, $10^{-7}$ A. Time scale, 2 msec. Resting potential, -70 mV. Ca$^{2+}$ 5.4 mM.

Fig. 4. Time courses of changes in the amplitude of the end-plate potential (open circle) and the end-plate current (filled circle), rise time (open triangle) and 50% fall time (filled triangle) before and after changing the concentration of dTc from $2.4 \times 10^{-6}$ g/ml to $1.2 \times 10^{-8}$ g/ml. The same preparation as in Fig. 3.

The active phase of the end-plate potential in PG was somewhat longer than that in dTc. Both the 50% fall time and total duration were slightly prolonged as compared with those in dTc, while the rise time was the same as in dTc. This was confirmed by recording the end-plate current before and after application of PG as shown in Figs. 1 and 2. After changing the solution from a dTc containing saline to a PG containing one, both the 50% fall time and total duration were prolonged. It was clear from Fig. 1 and 2 that this prolongation was not due to a change in the size of the end-plate potential or the end-plate current. Further-
more, the time course of the end-plate current in various concentrations of dTc was fairly constant and did not vary with the size of the end-plate potential or the end-plate current as shown in Figs. 3 and 4.

Membrane polarization and end-plate potential or end-plate current

The amplitudes of both end-plate potential and end-plate current in dTc were decreased with depolarization and were increased with hyperpolarization, and the

Fig. 5. End-plate potentials from an end-plate at various membrane potentials in dTc (1.5\times10^{-6} g/ml) and in PG (3.0\times10^{-4} g/ml). Left, in dTc and right, in PG. The membrane potentials were altered by passing current through the second microelectrode. Voltage scale, 25 mV. Time scale, 40 msec. Resting potential, -90 mV. Ca^{2+}, 5.4 mM.

Fig. 6. Relation between the amplitude of end-plate potential and membrane potential in dTc and in PG. Open circles were obtained from an end-plate in dTc and filled circles were obtained from the same end-plate in PG. The same preparation as in Fig. 5.
Fig. 7. End-plate potentials recorded from an end-plate at various membrane potentials in PG at a concentration of $3.0 \times 10^{-5}$ g/ml. A similar experiment with that of Figs. 1 and 2, but the duration of the passed current through were far longer than that of Figs. 1 and 2. Voltage scale, 25 mV. Time scale, $2.0 \times$ sec. Resting potential, $-80$ mV. Ca$^{2+}$, 3.6 mM.

Fig. 8. Relation between amplitude of the end-plate current and membrane potential in dTc at a concentration of $1.2 \times 10^{-6}$ g/ml and in PG at a concentration of $4.0 \times 10^{-5}$ g/ml. Open circles were obtained from an end-plate in dTc. Resting potential, $-90$ mV. Filled circles were obtained from another end-plate in PG. Resting potential, $-72$ mV. Ca$^{2+}$, 5.4 mM.

relationships were linear. If the lines were extrapolated, both lines crossed at about $15$ mV negative to the outer saline solution (Fig. 5 left, Fig. 6 open circle, and Fig. 8 open circle). These results agree to those of Takeuchi and Takeuchi (1959). The same relationships were obtained in the end-plates treated with SCh, a depolarizing receptor-blocking agent.
In contrast to the results obtained in dTc or in SCh, the relation between the end-plate potential and the membrane potential in PG was inversely proportional, i.e., the end-plate potential was increased with depolarization and was decreased with hyperpolarization (Fig. 5 right, and Fig. 6 closed circle). This change in the size of the end-plate potential appeared immediately after the polarization began and remained almost constant during the period of polarization and returned to the control level as soon as the polarization was turned off (Fig. 7). Furthermore, the relation between the amplitude of the end-plate current and the membrane potential, in which it was unnecessary to consider the electrical characteristics of the muscle membrane around the end-plate (Takeuchi and Takeuchi 1959), was the same as that between the end-plate potential and membrane potential as shown in Fig. 8 (closed circle).

![Figure 9](image_url)

Fig. 9. Direct muscle action potentials with and without end-plate potential in PG at a concentration of $2.0 \times 10^{-5}$ g/ml. Voltage scale, 50 mV. Time scale, 2 msec. Resting potential, -90 mV. Equilibrium potential, about -13 mV. NaCl, 230 mM, Ca$^{2+}$, 5.4 mM.

Therefore, the equilibrium potentials of both the end-plate potential and end-plate current could not be extrapolated by these lines. However, comparing the action potential set up by stimulation alone with the one, in which the end-plate potential was superimposed, the equilibrium potential of the end-plate potential in PG was found not to be significantly different from those in dTc or those in normal saline solution (Fig. 9). The evidence suggests that PG might act somewhere in the end-plate, leading to a reduction in the increase in permeability to both sodium and potassium ions, but does not affect the ratio of $P_{Na}/P_{K}$ which is increased by ACh.

**DISCUSSION**

There are several factors which determine the time course of the active phase as discussed by Takeuchi and Takeuchi (1959). They described that the falling
phase of the normal end-plate current was somewhat slower than that of the
curarized end-plate current and that ΔTC might shorten the time course of the
end-plate current perhaps by competing with the receptor in the end-plate for
ACh. The time course of the end-plate current in PG was rather similar to that
of the normal one reported by Takeuchi and Takeuchi (1959). Therefore, the
slower time course of the end-plate current in PG may be due to a non-competitive
action of PG, which is supposed to interact with a receptor other than that at
which the transmitter acts.

Another possibility is that PG acts on the process of liberation of the trans-
mitter from the nerve terminals. Guanidine itself increases the quantity of
transmitter released per nerve impulse (Otsuka and Endo 1960), and some guani-
dino-derivatives also potentiate the neuromuscular transmission (Ozawa and
Takeda 1965). Therefore, it is possible that PG acts on the presynaptic site and
increases the transmitter output. Tetraethylammonium (TEA), which prolongs
the duration of the presynaptic nerve action potential, increases the size of the
end-plate potential with a decrease in postsynaptic sensitivity to ACh (Koketsu
1958). Thesleff and Quastel (1965) described that the prolongation of the dura-
tion of the action potential might increase fractional release and thus the number
of quanta per impulse. Takeuchi and Takeuchi (1959) also suggested an intimate
relationship between the time course of the end-plate current and that of the nerve
impulse. It is not clear whether PG influences the time course of the action
potential of nerve terminals, since it did not influence the nerve action potential
recorded with a sucrose-gap method although PG prolonged slightly the duration
of the muscle action potential (Nishiyama and Kuwabara 1973).

Although a reasonable explanation which accounts for the inverse relationship
between the membrane polarization and end-plate potential or end-plate current
in PG has not been obtained, this might be due to either change in liberation of
the transmitter from the nerve terminals or in some step of the postsynaptic
process, which begins with the interaction of ACh with its receptor and ends by
the increase in the membrane permeability to cations.

Takeuchi and Takeuchi (1961) reported a reduction of transmitter output
with hyperpolarization in high potassium media. Recently, Maeno, Edward and
Hashimura (1970) have observed that the change in amplitude of the end-plate
potential with hyperpolarization was smaller than that of depolarization in a
muscle treated with procaine. They concluded that the nonlinear relationship
was due to a reduction of ACh output with hyperpolarization, and that the post-
synaptic process might be independent of the muscle membrane potential.

On the other hand, Kordas (1969) suggested the possibility that the relation-
ship obtained in glycerol treated muscle, which is similar to that in procaine,
might be explained by a change in the receptor-ACh interaction. It is possible
that the polarizing current changes the interaction of positively charged PG with
some receptor in the end-plate directly or by altering the concentration of PG at
the surface of the end-plate.
Acknowledgment

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References