Original Article

Systemic Histopathology of Infant Rats Exposed to Busulfan

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Abstract: Busulfan is an antineoplastic bifunctional alkylating agent. We previously reported the busulfan-induced systemic histopathological changes in fetal rats and the sequence of brain lesions in fetal and infant rats. In the present study, in order to clarify the nature and sequence of busulfan-induced systemic histopathological changes in infant rats, 6-day-old male infant rats were subcutaneously administered 20 mg/kg of busulfan and histopathologically examined at 1, 2, 4, 7 and 14 days after treatment (DAT). As a result, histopathological changes characterized by pyknosis of component cells were observed in the heart, lungs, stomach, intestines, liver, kidneys, testes, epididymides, hematopoietic and lymphoid tissues, dorsal skin and femur as well as in the brain and eyes (data not shown in this paper). Such pyknosis transiently appeared until 7 DAT with prominence at 2 and/or 4 DAT in each tissue, except for the thymus, in which pyknosis peaked at 1 DAT. Most of the pyknotic nuclei were immunohistochemically positive for cleaved caspase-3, indicating that pyknotic cells were apoptotic. Different from the reports of fetal and adult rats, apoptosis was also found in cardiomyocytes and osteoblasts in infant rats. (DOI: 10.1293/tox.2013-0043; J Toxicol Pathol 2014; 27: 25–29)

Key words: busulfan, systemic histopathology, apoptosis, infant rat

Introduction

Busulfan, a bifunctional alkylating agent, has been used for the treatment of chronic myeloid leukemia and for myeloablative-conditioning regimens before stem cell transplantation. In children, there are several reports of diverse effects of busulfan treatment such as pulmonary fibrosis and acute clinical neurotoxicity (spasm)^{1–3}.

Busulfan has teratogenic and cytotoxic potentials⁴, and it is reported that rat fetuses exposed to busulfan developed microencephaly and microphthalmia⁵. Our previous studies clarified the systemic histopathological changes⁶ and the sequence of the central nervous system (CNS) lesions characterized by neural progenitor cell apoptosis⁷ in rat fetuses transplacentally exposed to busulfan on gestation day 13. It is also reported that busulfan induces histopathological changes in the lungs^{8–11} in adult humans and in gastrointestinal tissues¹², lymphoid tissues¹³ and gonadal tissues^{14–18} in adult rats. On the other hand, there are few reports of systemic histopathological changes in infant animals induced by busulfan except for our previous report of busulfan-induced CNS lesions in infant rats¹⁹.

In the present study, we examined the busulfan-induced systemic histopathological changes in infant rats mainly

from the viewpoints of the distribution and sequence of pyknosis of component cells, except for brain¹⁹ and eye lesions, which will be described elsewhere in the near future.

Materials and Methods

Animals

Male newborn rats were obtained in our laboratory by mating females with males of the same colony of specific pathogen-free rats of the Sprague-Dawley strain purchased from Charles River Laboratories Japan, Inc. (Kanagawa, Japan). One foster mother with 8 male newborns were housed together in plastic Econ cages (W 340 mm × D 450 mm × H 185 mm) with bedding (White flakes: Charles River Laboratories Japan, Inc.) in an environmentally controlled animal room (temperature, 23 ± 3°C; relative humidity, 50 ± 20%; air ventilation rate, 10–15 times per hour; lighting, 12 h/12 h light/dark cycle) and fed an irradiation-sterilized pelleted diet (NMF, Oriental Yeast Co., Ltd., Tokyo, Japan) and tap water *ad libitum*. Finally, a total of fifty 6-day-old male rats were subjected to the experiment. The protocol of this study was reviewed and approved by the Animal Care and Use Committee of BoZo Research Center.

Experimental designs

Busulfan was obtained from Sigma Chemicals (St. Louis, MO, USA) and was suspended with olive oil.

Fifty 6-day-old male rats were equally divided into the control and busulfan groups. The animals of the busulfan group were subcutaneously administered 20 mg/kg (10 mL/kg body weight) of busulfan, and those of the control group

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were subcutaneously administered 10 mL/kg of olive oil, respectively. The dose of busulfan was decided based on the results of our preliminary study. Five animals each of the busulfan and control groups were euthanized at 1, 2, 4, 7 and 14 days after treatment (DAT), respectively. At necropsy, all organs and tissues were collected from each animal for histopathological examination.

Histopathology and immunohistochemistry for cleaved caspase-3

Collected organs and tissues were fixed with 10% neutral buffered formalin. After fixation, the femur was decalcified in formic acid solution. Four- μ m paraffin sections were stained with hematoxylin and eosin (HE) and subjected to histopathological examination.

Some of the paraffin sections were also subjected to immunohistochemical examination for cleaved caspase-3. In brief, sections were reacted with rabbit anti-cleaved caspase-3 polyclonal antibody (1:200, Cell Signaling Technology, Beverly, MA, USA) at 4°C overnight after pretreatment. Then, the sections were reacted with an EnVision+kit (Dako Japan) at room temperature for 40 min. These sections were visualized by peroxidase-diaminobenzidine (DAB, Dojindo Laboratories, Kumamoto, Japan) reaction and then counterstained with hematoxylin.

Histopathological examination was performed on tissues such as the heart, lungs, stomach, intestines, liver, pancreas, kidneys, testes, epididymides, thymus, spleen, mesenteric lymph node, bone marrow, skin (dorsal) and bone (femur) based on the results of our preliminary study.

Results

No deaths occurred in any group until 7 DAT. Thereafter, one animal died with severe myelosuppression at 13 DAT in the busulfan group.

In the control group, there were no histopathological changes observed in any tissues. On the other hand, in the busulfan group, histopathological changes mainly characterized by pyknosis of component cells were observed in many tissues as listed in Table 1. Histopathological changes other than pyknosis are shown in Table 2. Histopathological changes were also detected in the brain and eyes, but their data were excluded from the present paper as mentioned above.

In the cardiopulmonary system, pyknosis was observed in a small number of cardiomyocytes (Fig. 1a) and alveolar and terminal bronchiolar epithelial cells at 2 and 4 DAT (Fig. 1b). In the digestive system, pyknosis was found in a small number of hematopoietic cells in the liver at 2 DAT, glandular epithelial cells in the stomach (Fig. 1c) from 1 to 7 DAT, and crypt epithelial cells in the intestines from 1 to 4 DAT. Hematopoietic cells in the liver mildly decreased from 4 to 14 DAT, and glandular epithelial cells in the stomach showed vacuolation at 4 DAT.

In the urogenital system, pyknosis was found in a small number of proximal and distal tubule epithelial cells in the

kidneys (Fig. 1d) at 2 and 4 DAT. Pyknotic changes in spermatogonia started at 1 DAT and became moderate at 2 and 4 DAT in the testes (Fig. 1e). Thereafter, seminiferous tubules showed atrophy with depletion of germ cells at 7 and 14 DAT, at which point only Sertoli cells were left in the germinal epithelium of markedly atrophied seminiferous tubules (Fig. 1f). Pyknosis was also found in a small number of epithelial cells in the epididymides from 2 to 7 DAT.

In the hematopoietic and lymphoid system, the thymus showed moderate cortical atrophy at 2 and 4 DAT following moderate or mild pyknotic changes in cortical lymphocytes at 1 and 2 DAT (Fig. 1g). Similar but less severe changes were observed in mesenteric lymph nodes at 4 and 7 DAT. In the spleen, a minimal or mild decrease in the number of hematopoietic cells was detected from 2 to 14 DAT. In the bone marrow, mild or moderate pyknotic changes of hematopoietic cells were found from 1 to 7 DAT. A decrease in the number of hematopoietic cells with fat cell infiltration started at 2 DAT, progressed thereafter and became prominent at 14 DAT (Fig. 1h). In the other tissues, pyknosis was found in a small number of hair follicle epithelial cells (Fig. 1i) in the dorsal skin and osteoblasts (Fig. 1j) in the femur at 2 and 4 DAT. Most of the pyknotic nuclei were immunohistochemically positive for cleaved caspase-3 (Fig. 1e, inset), indicating that pyknotic cells were apoptotic.

Discussion

In the present study, we examined the nature and sequence of systemic histopathological changes observed in infant rats exposed to busulfan (20 mg/kg) at 6 days of age. As mentioned above, those in the CNS have been previously reported¹⁹, and those in the eyes will be published elsewhere in the near future.

Pyknosis of component cells was detected in many tissues (Table 1). Among them, the thymus was moderately affected by pyknosis at 1 DAT, and the bone marrow and testes were moderately affected by pyknosis at 2 and 4 DAT. Most of the pyknotic nuclei were immunohistochemically positive for cleaved caspase-3. This strongly indicates that pyknotic cells are apoptotic. In addition, moderate cortical atrophy was observed simultaneously with moderate pyknosis of cortical lymphocytes in the thymus, a moderate to marked decrease in the number of hematopoietic cells with infiltration of fat cells was found from 4 to 14 DAT in the bone marrow, and moderate or marked atrophy due to depletion of germ cells developed at 7 and 14 DAT in the testes. Thus, histopathological changes remained until 14 DAT in the bone marrow and testes, and whether or not the rats could recover from such lesions in the bone marrow and testes thereafter was not clear in the present study. On the other hand, histopathological changes observed in tissues other than the bone marrow and testes were considered to be transient in nature.

Although there were no reports of cardiac lesions in fetal⁶ or adult rats¹² following exposure to busulfan, apoptosis of cardiomyocytes was detected in infant rats in the pres-

Table 1. Distribution and Sequence of Pyknotic Cells in Rat Infant Tissues Exposed to Busulfan

	Dose of busulfan	0 mg/kg					20 mg/kg				
		Days after treatment									
	No. of animals examined	1	2	4	7	14	1	2	4	7	14
Heart											
Cardiomyocytes		–	–	–	–	–	–	±	±	–	–
Lungs											
Epithelial cells of alveoli or terminal bronchioles		–	–	–	–	–	–	±	±	–	–
Stomach											
Glandular epithelial cells		–	–	–	–	–	±	+	+	±	–
Intestines											
Crypt cells		–	–	–	–	–	±	±	±	–	–
Liver											
Hematopoietic cells		–	–	–	–	–	–	±	–	–	–
Kidneys											
Tubular cells		–	–	–	–	–	–	±	±	–	–
Testes											
Spermatogonia		–	–	–	–	–	±	++	++	±	–
Epididymides											
Epithelial cells		–	–	–	–	–	–	+	+	±	–
Thymus											
Lymphocytes of the cortex		–	–	–	–	–	++	+	–	–	–
Mesenteric lymph node											
Lymphocytes of the cortex		–	–	–	–	–	–	–	+	–	–
Bone marrow											
Hematopoietic cells		–	–	–	–	–	+	++	++	+	–
Skin (dorsal)											
Epithelial cells of hair follicles		–	–	–	–	–	–	±	±	–	–
Bone (femur)											
Osteoblasts		–	–	–	–	–	–	±	±	–	–

No. of pyknotic cells/No. of cells counted: –, almost absent; ±, minimal <25%; +, mild 25%–50%; ++, moderate 50%–75%; +++, marked >75%.

Table 2. Summary of the Histopathological Findings for Rat Infant Tissues Exposed to Busulfan

	Dose of busulfan	0 mg/kg					20 mg/kg				
		Days after treatment									
	No. of animals examined	1	2	4	7	14	1	2	4	7	14
Stomach											
Vacuolation of glandular epithelial cells		–	–	–	–	–	–	–	+	–	–
Liver											
Decreased hematopoietic cells		–	–	–	–	–	–	–	+	+	+
Testes											
Atrophy with depletion of germ cells		–	–	–	–	–	–	–	–	++	+++
Thymus											
Atrophy of the cortex		–	–	–	–	–	–	++	++	–	–
Mesenteric lymph node											
Atrophy of the cortex		–	–	–	–	–	–	–	±	+	–
Spleen											
Decreased hematopoietic cells		–	–	–	–	–	–	+	+	+	±
Bone marrow											
Decreased hematopoietic cells with fat cells infiltration		–	–	–	–	–	–	+	++	++	+++

Lesion area/Tissue area observed: –, almost absent; ±, minimal <25%; +, mild 25%–50%; ++, moderate 50%–75%; +++, marked >75%.

ent study, suggesting a susceptibility of the infant rat heart to busulfan. Regarding pulmonary lesions, it has been reported in humans that long-term and/or high-dose busulfan therapy brought about such pulmonary lesions as broncho-pulmonary dysplasia and diffuse interstitial pulmonary fibrosis in adults^{9–11} and children^{1,2}. These lesions are known as “busulfan lungs.” In the lungs of fetal⁶ and infant rats,

only transient apoptotic changes were detected in alveolar and terminal bronchiolar epithelial cells.

With regard to histopathological changes in the gastrointestinal tissues, apoptotic changes were common in fetal⁶ and infant rats. Namely, they were milder in the intestine than in the stomach in fetal and infant rats, while they were reported to be milder in the stomach than in the intestine

Fig. 1. Histopathological changes in infant rat tissues. In the busulfan group, pyknosis (arrowhead) was observed in cardiomyocytes (a), alveolar or bronchiolar epithelial cells (b), glandular epithelial cells in the stomach (c), uriniferous tubule epithelial cells in the kidneys (d), spermatogonia (e), lymphocytes in the thymus (g), hair follicle epithelial cells (i) and osteoblasts in the femur (j). Pyknotic nuclei were immunohistochemically positive for cleaved caspase-3 (e, inset). At 14 DAT, only Sertoli cells were left in the germinal epithelium of markedly atrophied seminiferous tubules (f), and marked depletion of hematopoietic cells with prominent infiltration of fat cells was observed in the femur bone marrow (h). HE stain, inset of (e) cleaved caspase-3 immunostaining. (a–f, i and j) Bar=50 µm. (g) Bar=100 µm. (h) Bar=500 µm.

in adult rats¹². In humans⁸, although there were no reports of histopathological changes in the gastrointestinal tissues, clinical signs of nonspecific gastroenteritis were reported. In the kidneys, although there were no reports of apoptosis in tubular epithelial cells in adult rats and humans, apoptosis of tubular epithelial cells was observed in fetal⁶ and infantile rats, suggesting that tubular epithelial cells of infant rats still remain susceptible to busulfan. The outline of the testicular lesions in infant rats was similar to those in adult rats^{14–18}, while there have been no reports of testicular lesions in humans.

Histopathological changes in the thymus and mesenteric lymph nodes were similar between infant and adult rats^{12,13}. On the other hand, atrophy of the splenic white pulp, reported in adult rats¹³, was not clear in infant rats. In the bone marrow of infant rats, as mentioned above, the number of hematopoietic cells decreased with time and became marked at 14 DAT with prominent infiltration of fat cells. This corresponded well to depressed bone marrow cellularity reported in adult rats^{12,13,20}.

In our previous study on histopathological changes in fetal rats⁶, we observed apoptosis of component cells in mesenchymal tissues such as craniofacial tissues, the mandible, limb buds and the tail bud. In the present study on histopathological changes in infant rats, apoptosis was found in hair follicle epithelial cells in the dorsal skin and osteoblasts in the femur, which were not reported in adult rats.

In conclusion, the present study showed that busulfan-induced histopathological changes were characterized by apoptosis of component cells and that the distribution and sequence of apoptosis showed some differences, especially between infant and adult rats, probably reflecting the difference in susceptibility of component cells to busulfan between them.

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References

1. Oakhill A, Green ID, Knowlson GT, Cameron AH, Shah KJ, Hill FGH, and Mann JR. Busulphan lung in childhood. *J Clin Pathol*. **34**: 495–500. 1981. [Medline] [CrossRef]
2. Fauroux B, Clement A, and Toumier G. Pulmonary toxicity of drugs and thoracic irradiation in children. *Rev Mal Respir*. **13**: 235–242. 1996. [Medline]
3. Vassal G, Deroussent A, Hartmann O, Challine D, Benhamou E, Valteau-Couanet D, Brugieres L, Kalifa C, Gouyette A, and Lemerle J. Dose-dependent neurotoxicity of high-dose busulfan in children: A clinical and pharmacological study. *Cancer Res*. **50**: 6203–6207. 1990. [Medline]
4. Bishop JB, and Wassom JS. Toxicological review of busulfan (Myleran). *Mutat Res*. **168**: 15–45. 1986. [Medline] [CrossRef]
5. Furukawa S, Usuda K, Abe M, and Ogawa I. Microencephaly and microphthalmia in rat fetuses by busulfan. *Histol Histopathol*. **22**: 389–397. 2007. [Medline]
6. Ohira T, Ando R, Andoh R, Nakazawa T, Nishihara K, Yamamoto S, Nakamura N, and Tamura K. Distribution and sequence of pyknotic cell in rat fetuses exposed to busulfan. *J Toxicol Pathol*. **22**: 167–171. 2009. [Medline] [CrossRef]
7. Ohira T, Ando R, Okada Y, Suzuki H, Saito T, Nakazawa T, Nishihara K, Yamamoto S, Nakamura N, and Tamura K. Sequence of busulfan-induced neural progenitor cell damage in the fetal rat brain. *Exp Toxicol Pathol*. **65**: 523–530. 2013. [Medline] [CrossRef]
8. Gureli N, Denham SW, and Root SW. Cytologic dysplasia related to Busulfan (Myleran) therapy. *Obstet Gynecol*. **21**: 466–470. 1963. [Medline]
9. Littler WA, Kay JM, and Hasleton PS. Busulfan lung. *Thorax*. **24**: 639–655. 1969. [Medline] [CrossRef]
10. Burns WA, MacFarland W, and Matthews MJ. Busulfan-induced pulmonary disease: report of a case and review of the literature. *Am Rev Respir Dis*. **101**: 408–413. 1970. [Medline]
11. Buggia I, Locatelli F, Regazzi MB, and Zecca M. Busulfan. *Ann Pharmacother*. **28**: 1055–1062. 1994. [Medline]
12. Sternberg SS, Philips FS, and Scholler J. Pharmacological and pathological effects of alkylating agents. *Ann N Y Acad Sci*. **68**: 811–825. 1958. [Medline] [CrossRef]
13. Tange T. Significance of the bone marrow lymphocyte hematopoietic regeneration following acute injury due to cobalt-60 irradiation and cytotoxic drugs. *Acta Pathol Jpn*. **24**: 93–117. 1974. [Medline]
14. Kasuga F, and Takahashi M. The endocrine function of rat gonads with reduced number of germ cells following busulphan treatment. *Endocrinol Jpn*. **33**: 105–115. 1986. [Medline] [CrossRef]
15. Udagawa K, Ogawa T, Watanabe T, Yumura Y, Takeda M, and Hosaka M. GnRH analog, leuporelin acetate, promotes regeneration of rat spermatogenesis after severe chemical damage. *Int J Urol*. **8**: 615–622. 2001. [Medline] [CrossRef]
16. Horn MM, Paz AH, Duarte ME, Baldo G, Belardinelli MC, Matte U, Lima EO, and Passos EP. Germinative testicular cells and bone marrow mononuclear cells transplanted to a rat model of testicular degeneration. *Cloning Stem Cells*. **10**: 543–546. 2008. [Medline] [CrossRef]
17. Hodel C, Ettlin RA, and Zschauer A. Morphological changes produced in rat testis by anticancer drugs. *Arch Toxicol Suppl*. **7**: 147–150. 1984. [Medline] [CrossRef]
18. Linder RE, Strader LF, Slott VL, and Suarez JD. Endpoints of spermatotoxicity in the rat after short duration exposures to fourteen reproductive toxicants. *Reprod Toxicol*. **6**: 491–505. 1992. [Medline] [CrossRef]
19. Ohira T, Ando R, Saito T, Yahata M, Oshima Y, and Tamura K. Busulfan-induced pathological changes of the cerebellar development in infant rats. *Exp Toxicol Pathol*. **65**: 789–797. 2013. [Medline] [CrossRef]
20. Dunjic A, and Cuvelier AM. Survival of rat bone marrow cells after treatment with myleran and endoxan. *Exp Hematol*. **1**: 11–21. 1973. [Medline]